

US009422528B2

(12) United States Patent

Suphaphiphat et al.

(10) Patent No.: US 9,422,528 B2

(45) **Date of Patent:** Aug. 23, 2016

(54) INFLUENZA VIRUS REASSORTMENT

(71) Applicant: Novartis AG, Basel (CH)

(72) Inventors: Pirada Suphaphiphat, Brookline, MA

(US); Peter Mason, Somerville, MA (US); Bjoern Keiner, Basel (CH); Philip Dormitzer, Weston, MA (US); Heidi Trusheim, Apex, NC (US)

(73) Assignee: Novartis AG, Basel (CH)

(*) Notice: Subject to any disclaimer, the term of this

patent is extended or adjusted under 35

U.S.C. 154(b) by 0 days.

(21) Appl. No.: 13/909,013

(22) Filed: Jun. 3, 2013

(65) **Prior Publication Data**

US 2014/0030291 A1 Jan. 30, 2014

Related U.S. Application Data

- (63) Continuation of application No. PCT/EP2013/054227, filed on Mar. 2, 2013.
- (60) Provisional application No. 61/605,922, filed on Mar. 2, 2012, provisional application No. 61/685,766, filed on Mar. 23, 2012.
- (51) **Int. Cl.**A61K 39/145 (2006.01)
 C12N 7/00 (2006.01)
- (52) **U.S. CI.** CPC *C12N 7/00* (2013.01); *C12N 2760/16121* (2013.01); *C12N 2760/16134* (2013.01)
- (58) Field of Classification Search
 NoneSee application file for complete search history.

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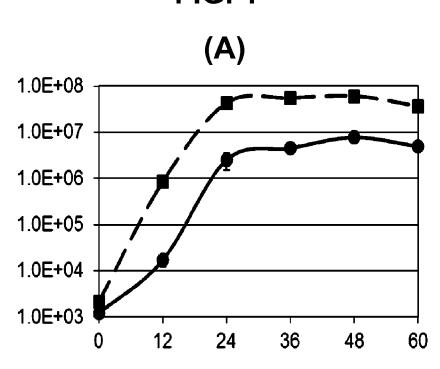
Primary Examiner — Nick Zou (74) Attorney, Agent, or Firm — Finnegan, Henderson, Farabow, Garrett & Dunner, LLP

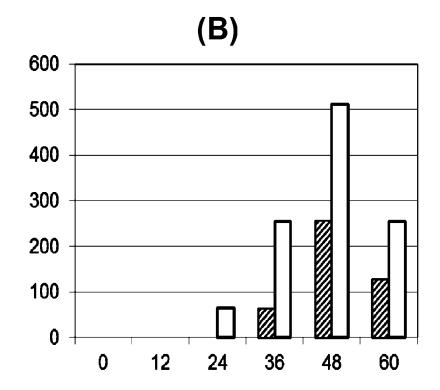
(57) ABSTRACT

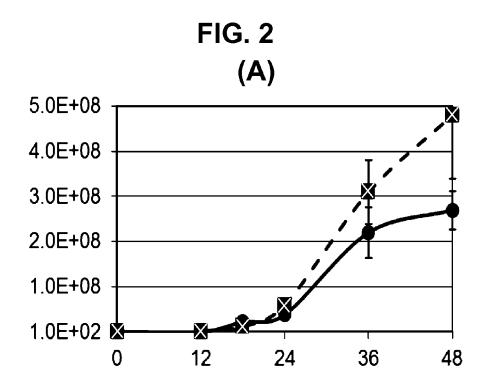
New influenza donor strains for the production of reassortant influenza A viruses are provided.

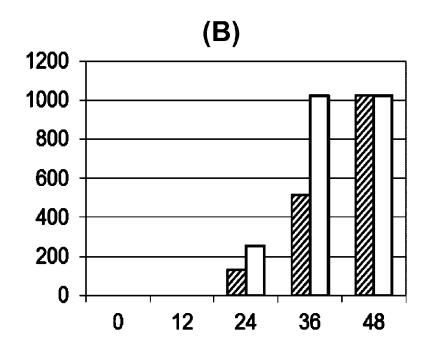
29 Claims, 18 Drawing Sheets

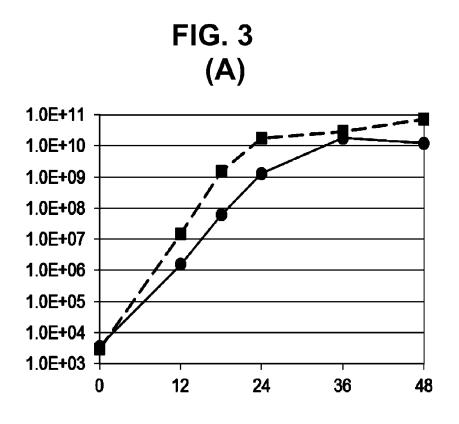
FIG. 1

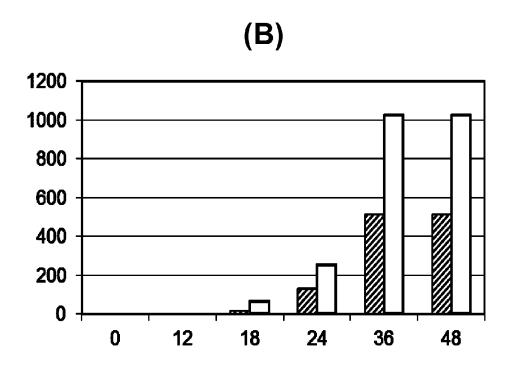


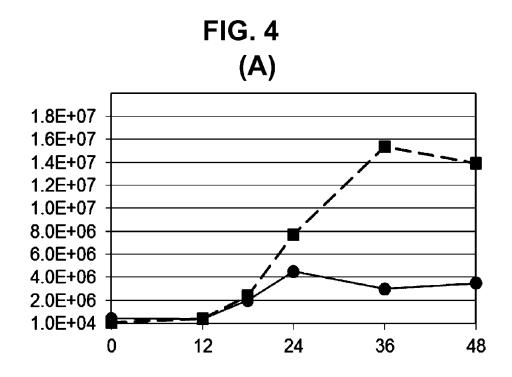


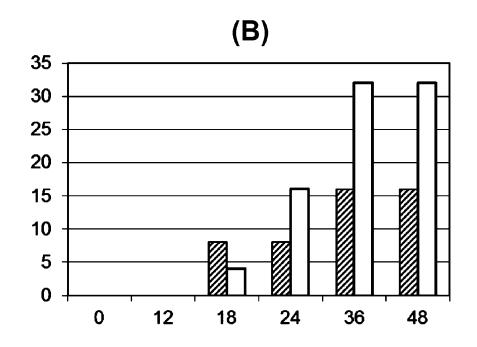


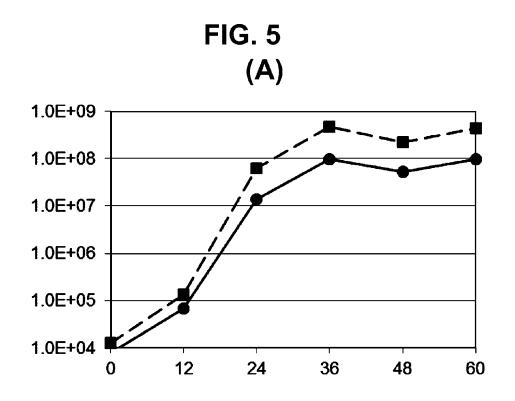












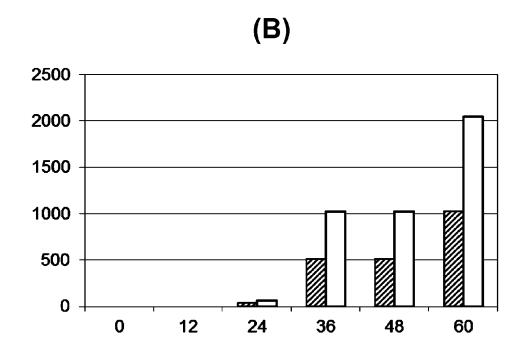


FIG. 6 (A) 1.0E+10 1.0E+09 1.0E+08 1.0E+07 1.0E+06 1.0E+05 1.0E+04 1.0E+03 1.0E+02 1.0E+01 12 24 0 36 48 60

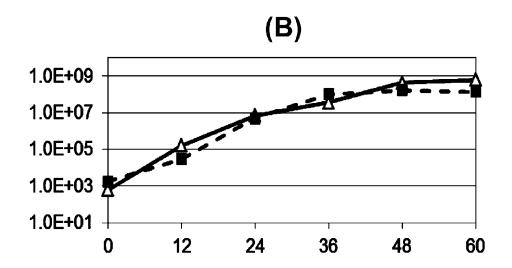


FIG. 7 (A) 10.00 9.00 8.00-7.00-6.00 5.00 4.00-3.00-2.00-1.00-0.00 12hpi 36hpi 60hpi 1hpi

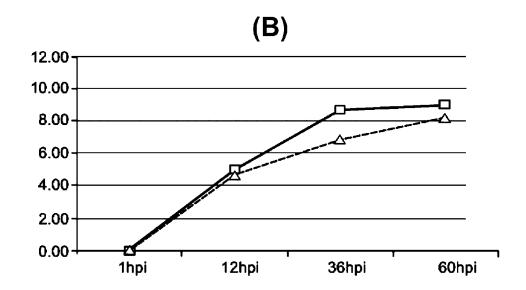
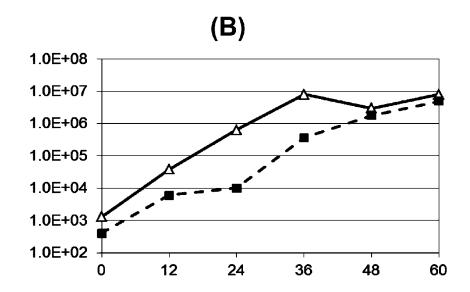
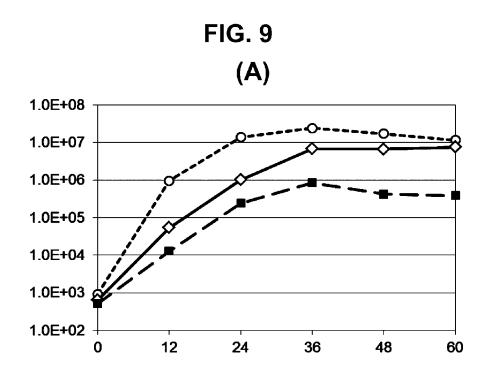
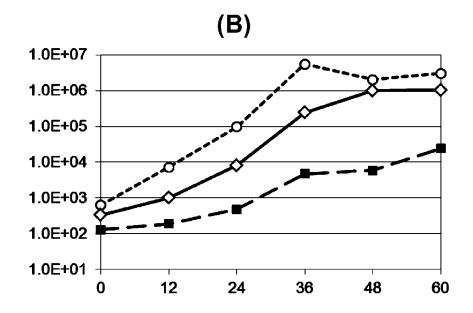
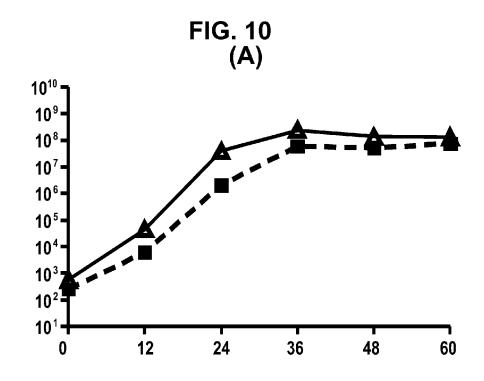


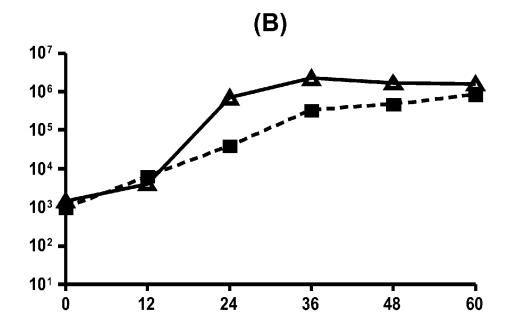
FIG. 8 (A) 1.0E+08 1.0E+07 1.0E+06 1.0E+05 1.0E+04 1.0E+03 1.0E+02 1.0E+01 1.0E+00 12 0 24 36 48 60











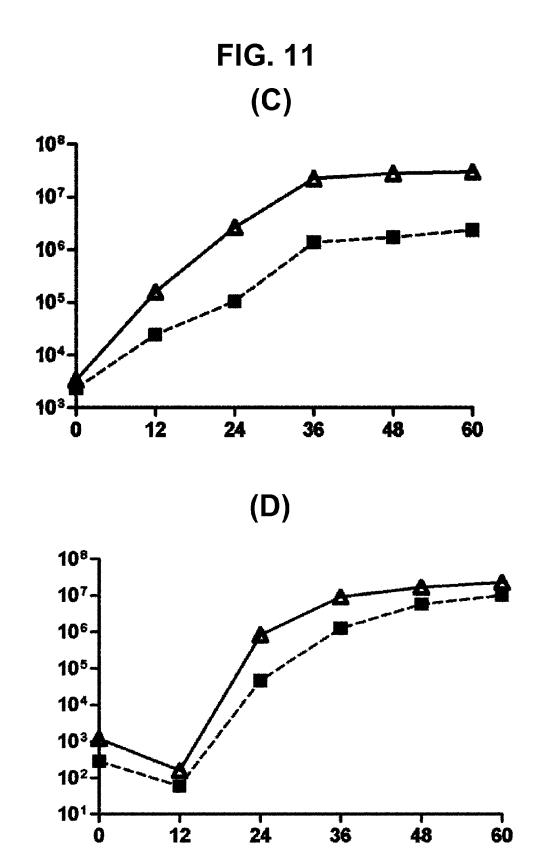
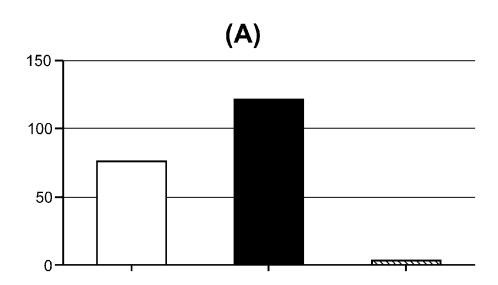


FIG. 12

10⁷
10⁶
10³
0
12
24
36
48
60

FIG. 13



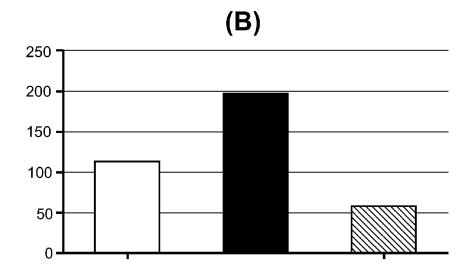
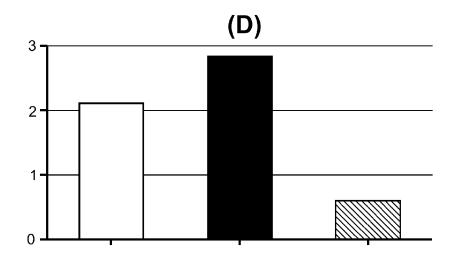


FIG. 14
(C)



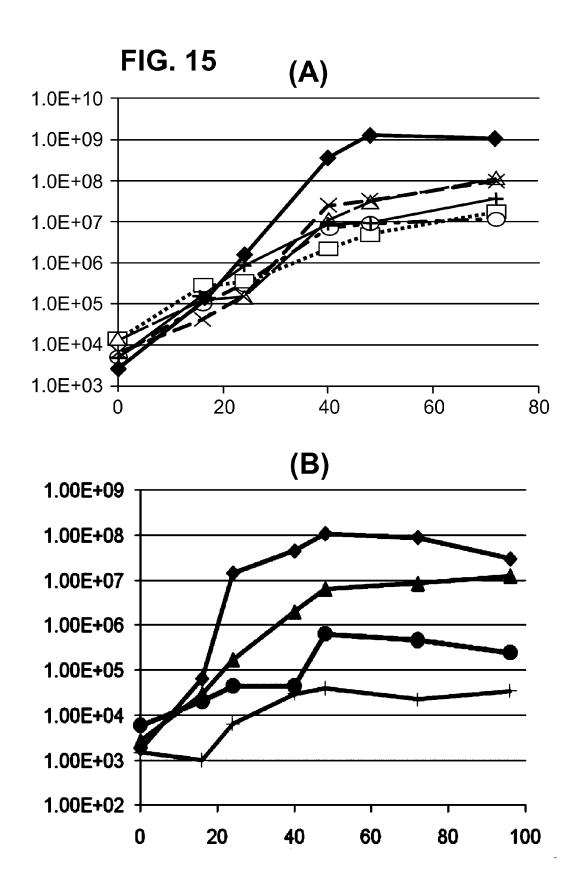
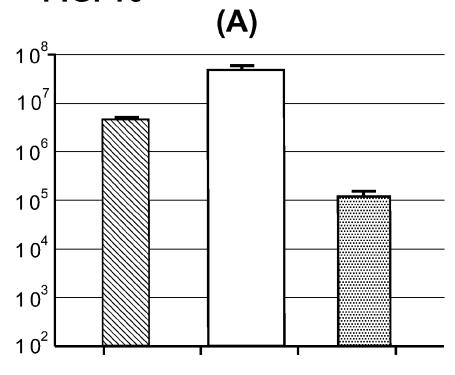


FIG. 16



(B)

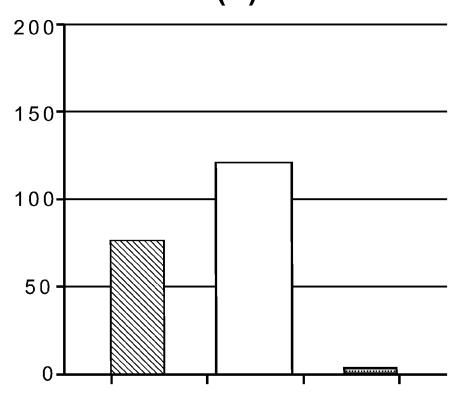
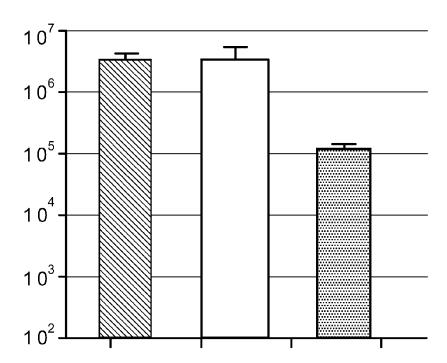
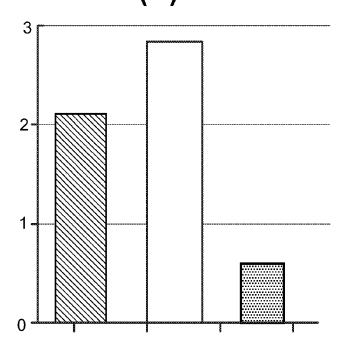


FIG. 17





(D)



105p30	\leftarrow	mslltevetyvlsivpsgplkaeiaqrlenvfagkntdlealmewlktrp	20
A/NC/20/66	ᅮ	ms1ltevetyv1sivpsgplkaeiaqrlenvfagkntdlealmewlktrp	50
105p30	51	ilspltkgilgfvftltvpserglgrrrfvgnalngngdpnnmdravkly	100
A/NC/20/66	51	ilspltkgilgfvftltvpserglqrrrfvqnalngngdpnnmdkavkly	100
105p30	101	rklkreitfhgakeialsysagalascmgliynrmgavttesafglicat	150
A/NC/20/66	101	rklkreitfhgakeialsysagalascmgliynrmgavttesafglicat	150
105p30	151	ceqiadsqhkshrqmvtttnplirhenrmvlasttakameqmagsseqaa	200
A/NC/20/66	151	ceqiadsqhkshrqmvtttnplirhenrmvlasttakameqmagsseqaa	200
105p30	201	eamevasqarqmvqamraigthpssstglkndllenlqayqkrmgvqmqr	250
A/NC/20/66	201	eamevasqarqmvqamraigthpssstglkndllenlqayqkrmgvqmqr	250
105p30	251	fk 252 HG 18	
A/NC/20/66	251	fk 252	

INFLUENZA VIRUS REASSORTMENT

This patent application is a continuation of International Application No. PCT/EP2013/054227, filed Mar. 2, 2013, which claims priority from U.S. provisional patent applications 61/605,922, filed Mar. 2, 2012 and 61/685,766 filed Mar. 23, 2012, the complete contents of which are incorporated herein by reference.

SUBMISSION OF SEQUENCE LISTING ON ASCII TEXT FILE

The content of the following submission on ASCII text file is incorporated herein by reference in its entirety: a computer readable form (CRF) of the Sequence Listing (file name: PAT055008_ST25.txt, date recorded: May 22, 2013, size: 161 KB).

TECHNICAL FIELD

This invention is in the field of influenza A virus reassortment. Furthermore, it relates to manufacturing vaccines for protecting against influenza A viruses.

BACKGROUND ART

The most efficient protection against influenza infection is vaccination against circulating strains and it is important to produce influenza viruses for vaccine production as quickly as possible.

Wild-type influenza viruses often grow to low titres in eggs and cell culture. In order to obtain a better-growing virus strain for vaccine production it is currently common practice to reassort the circulating vaccine strain with a faster-growing high-yield donor strain. This can be achieved by co-infecting a culture host with the circulating influenza strain (the vaccine strain) and the high-yield donor strain and selecting for reassortant viruses which contain the hemagglutinin (HA) and neuraminidase (NA) segments from the vaccine strain and the other viral segments (i.e. those encoding PB1, PB2, PA, NP, 40 M₁, M₂, NS₁ and NS₂) from the donor strain. Another approach is to reassort the influenza viruses by reverse genetics (see, for example references 1 and 2).

Reference 3 reports that a reassortant influenza virus containing a PB1 gene segment from A/Texas/1/77, the HA and 45 NA segments from A/New Caledonia/20/99, a modified PA segment derived from A/Puerto Rico/8/34 and the remaining viral segments from A/Puerto Rico/8/34 shows increased growth in cells.

There are currently only a limited number of donor strains 50 for reassorting influenza viruses for vaccine manufacture, and the strain most commonly used is the A/Puerto Rico/8/34 (A/PR/8/34) strain. However, reassortant influenza viruses comprising A/PR/8/34 backbone segments do not always grow sufficiently well to ensure efficient vaccine manufacture. Thus, there is a need in the art to provide further and improved donor strains for influenza virus reassortment.

SUMMARY OF PREFERRED EMBODIMENTS

The inventors have now surprisingly discovered that influenza viruses which comprise backbone segments from two or more influenza donor strains can grow faster in a culture host compared with reassortant influenza A viruses which contain all backbone segments from the same donor strain. In particular, the inventors have found that influenza viruses which comprise backbone segments derived from two high-yield

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donor strains can produce higher yield reassortants with target vaccine-relevant HA/NA genes than reassortants made with either of the two original donor strains.

In principle, all segments of closely related influenza A viruses can be specifically reassorted to produce viable viruses, but only a small fraction of these viruses will be high-growth reassortants, due to inefficient activities of the resulting viral components. The inventors have provided backbone combinations that produce the high yield strains.

Reassortant influenza A viruses comprising backbone segments from two or more influenza donor strains may contain the PB 1 and the PB2 viral segments from the same donor strain, in particular the A/New Caledonia/20/1999-like strain, referred to herein as the 105p30 strain. The PB1 and PB2 viral segments may have at least 95% identity or 100% identity with the sequence of SEQ ID NO: 2 and/or SEQ ID NO: 3.

Where the reassortant influenza A virus comprises backbone segments from two or three donor strains, each donor strain may provide more than one of the backbone segments
of the reassortant influenza A virus, but one or two of the donor strains can also provide only a single backbone segment.

Where the reassortant influenza A virus comprises backbone segments from two, three, four or five donor strains, one or two of the donor strains may provide more than one of the backbone segments of the reassortant influenza A virus. In general the reassortant influenza A virus cannot comprise more than six backbone segments. Accordingly, for example, if one of the donor strains provides five of the viral segments, the reassortant influenza A virus can only comprise backbone segments from a total of two different donor strains.

Where a reassortant influenza A virus comprises the PB1 segment from A/Texas/1/77, it preferably does not comprise the PA, NP or M segment from A/Puerto Rico/8/34. Where a reassortant influenza A virus comprises the PA, NP or M segment from A/Puerto Rico/8/34, it preferably does not comprise the PB1 segment from A/Texas/1/77. In some embodiments, the invention does not encompass reassortant influenza A viruses which have the PB1 segment from A/Texas/1/77 and the PA, NP and M segments from A/Puerto Rico/8/34. The PB1 segment from A/Texas/1/77 may have the sequence of SEQ ID NO: 46 and the PA, NP or M segments from A/Puerto Rico/8/34 may have the sequence of SEQ ID NOs 47, 48 or 49, respectively.

The inventors have also discovered that variants of known donor strains can grow to higher viral titres compared to the original donor strain and can therefore be better donor strains for reassorting influenza viruses. Examples of such strains are PR8-X and 105p30.

Influenza A virus strains of the invention can grow to higher viral titres in MDCK cells in the same time and under the same growth conditions compared with A/Puerto Rico/8/34 and/or have a higher rescue efficiency compared with reassortant influenza strains that comprise all backbone segments from the same influenza donor strain. Further provided is a reassortant influenza A virus comprising at least one backbone viral segment from such an influenza strain.

The invention also provides a reassortant influenza A virus comprising at least one backbone viral segment from a donor strain, wherein the donor strain is selected from the group consisting of 105p30 and PR8-X. When the at least one backbone viral segment is the PA segment it may have a sequence having at least 95% or at least 99% identity with a sequence selected from the group consisting of SEQ ID NOs: 9 and 17. When the at least one backbone viral segment is the PB1 segment, it may have a sequence having at least 95% or at least 99% identity with a sequence selected from the group

consisting of SEQ ID NOs 10 and 18. When the at least one backbone viral segment is the PB2 segment, it may have a sequence having at least 95% or at least 99% identity with a sequence selected from the group consisting of or SEQ ID NOs: 11 and 19. When the at least one backbone viral segment is the M segment it may have a sequence having at least 95% or at least 99% identity with a sequence selected from the group consisting of SEQ ID NOs: 13 and 21. When the at least one backbone viral segment is the NP segment it may have a sequence having at least 95% or at least 99% identity with a sequence selected from the group consisting of SEQ ID NOs: 12 and 20. When the at least one backbone viral segment is the NS segment it may have a sequence having at least 95% or at least 99% identity with a sequence selected from the group consisting of SEQ ID NOs: 14 and 22.

In embodiments where the reassortant influenza A virus comprises backbone segments from at least two influenza donor strains, at least one backbone segment may be derived from a donor strain selected from the group consisting of 105p30 and PR8-X, as discussed in the previous paragraph. 20 Preferred reassortant influenza A viruses comprise 1, 2, 3 or 4 viral segments from the 105p30 donor strain wherein the PA segment may have at least 95% identity or 100% identity with SEQ ID NO: 17, the NP segment may have at least 95% identity or 100% identity with SEQ ID NO: 20, the M seg- 25 ment may have at least 95% identity or 100% identity with SEQ ID NO: 21, and/or the NS segment may have at least 95% identity or 100% identity with SEQ ID NO: 22. In some embodiments such influenza A viruses may also comprise at least one backbone viral segment from the PR8-X donor 30 strain. Where the at least one viral segment is the PA segment it may have at least 95% identity or 100% identity with SEQ ID NO: 9. Where the at least one viral segment is the NP segment it may have at least 95% identity or 100% identity with SEQ ID NO: 12. Where the at least one viral segment is 35 the M segment it may have at least 95% identity or 100% identity with SEQ ID NO: 13. Where the at least one viral segment is the NS segment it may have at least 95% identity or 100% identity with SEQ ID NO: 9. The inventors have backbone segments grow well in cell culture. In general a reassortant influenza virus will contain only one of each backbone segment. For example, when the influenza virus comprises the PA segment from 105p30 it will not at the same time comprise the PA segment of PR8-X.

In preferred embodiments, the virus comprises viral segments having at least 95% identity or 100% identity with the sequence of (a) the PB2 segment of SEQ ID NO: 19, the PB 1 segment of SEQ ID: NO 18 and the NS segment of SEQ ID NO: 22; or (b) the PB2 segment of SEQ ID NO: 19, the PB1 50 segment of SEQ ID NO: 18 and the M segment of SEQ ID NO: 21; or (c) the PB2 segment of SEQ ID NO: 19, the PB1 segment of SEQ ID NO: 18 and the NP segment of SEQ ID NO: 20; or (d) the PB2 segment of SEQ ID NO 19, the PB1 segment of SEQ ID NO 18 and the PA segment of SEQ ID NO 55 17. These embodiments are preferred because the inventors have found that such reassortant influenza A viruses grow particularly well in cell culture.

The invention provides a method of preparing the reassortant influenza A viruses of the invention. These methods 60 comprise steps of (i) introducing into a culture host one or more expression construct(s) which encode(s) the viral segments required to produce an influenza A virus wherein the backbone viral segments are from two or more influenza strains; and (ii) culturing the culture host in order to produce 65 reassortant virus and optionally (iii) purifying the virus obtained in step (ii).

The method may comprise the steps of (i) introducing into a culture host one or more expression construct(s) which encode(s) the viral segments required to produce an influenza A virus wherein the backbone viral segments are from two or more influenza strains and the PB1 and PB2 segments are from the same donor strain; and (ii) culturing the culture host in order to produce reassortant virus and optionally (iii) purifying the virus obtained in step (ii).

Also provided is a method of preparing a reassortant influenza A virus of the invention comprising the steps of (i) introducing into a culture host one or more expression construct(s) which encode(s) the viral segments required to produce an influenza A virus wherein the backbone viral segments are from two or more influenza strains and the HA and the PB 1 segment are from different influenza strains which have the same influenza HA subtype; and (ii) culturing the culture host in order to produce reassortant virus and optionally (iii) purifying the virus obtained in step (ii).

The invention also provides a method of preparing a reassortant influenza A virus of the invention comprising steps of (i) introducing into a culture host one or more expression construct(s) which encode(s) the viral segments required to produce an influenza A virus wherein one or more backbone viral segment(s) is/are from a 105p30 and/or a PR8-X influenza strain and wherein at least one viral segment is derived from a second influenza strain; and (ii) culturing the culture host in order to produce reassortant virus and optionally (iii) purifying the virus obtained in step (ii).

The methods may further comprise steps of: (iv) infecting a culture host with the virus obtained in step (ii) or step (iii); (v) culturing the culture host from step (iv) to produce further virus; and optionally (vi) purifying the virus obtained in step (v).

The invention also provides a method for producing influenza viruses comprising steps of (a) infecting a culture host with a reassortant virus of the invention; (b) culturing the host from step (a) to produce the virus; and optionally (c) purifying the virus obtained in step (b).

The invention also provides a method of preparing a vacshown that reassortant influenza A viruses comprising such 40 cine, comprising steps of (d) preparing a virus by the methods of any one of the embodiments described above and (e) preparing vaccine from the virus.

> In a further embodiment, the invention provides influenza strains PR8-X and 105p30.

> The invention also encompasses variant H1N1 influenza virus strains in which the M genome segment has lysine in the position corresponding to amino acid 95 of SEO ID NO: 33 when aligned to SEQ ID NO: 33 using a pairwise alignment algorithm. The variant H1N1 influenza virus strains according to the invention may further have a HA segment which has glycine in the position corresponding to amino acid 225 of SEQ ID NO: 35 when aligned to SEQ ID NO: 35 and/or has asparagine in the position corresponding to amino acid 231 of SEQ ID NO: 35 when aligned to SEQ ID NO: 35 using a pairwise alignment algorithm. The variant H1N1 influenza virus strain may also have a NA segment which has histidine in the position corresponding to amino acid 70 of SEQ ID NO: 31 when aligned to SEQ ID NO: 31 using a pairwise alignment algorithm.

> The preferred pairwise alignment algorithm is the Needleman-Wunsch global alignment algorithm [4], using default parameters (e.g. with Gap opening penalty=10.0, and with Gap extension penalty =0.5, using the EBLOSUM62 scoring matrix). This algorithm is conveniently implemented in the needle tool in the EMBOSS package [5].

> The invention provides an expression system comprising one or more expression construct(s) comprising the vRNA

encoding segments of an influenza A virus wherein the expression construct(s) encode(s) the backbone viral segments from two or more influenza donor strains. The expression construct(s) may encode the PB1 and PB2 segments from the same donor strain.

The invention also provides an expression system comprising one or more expression construct(s) comprising the vRNA encoding segments of a 105p30 or PR8-X strain wherein the expression construct(s) comprise(s) at least one backbone viral segment from the 105p30 or PR8-X, or strain. 10 The expression construct(s) may further comprise the vRNAs which encode the PB2, NP, NS, M and PA segments from PR8-X.

The invention also provides a host cell comprising the expression systems of the invention. These host cells can 15 express an influenza A virus from the expression construct(s) in the expression system.

Expression constructs which can be used in the expression systems of the invention are also provided. For example, the invention provides an expression construct which encodes the 20 backbone segments of the reassortant influenza strains according to the invention on the same construct. Donor Strains

Influenza donor strains are strains which typically provide the backbone segments in a reassortant influenza virus, even 25 though they may sometimes also provide the HA or NA segment, but not both, of the virus. Usually, however, both the HA and the NA segment in a reassortant influenza virus will be from the vaccine strain.

The inventors have surprisingly discovered that reassortant influenza A viruses comprising backbone segments from two or more influenza donor strains can grow to higher titres in cell culture compared with reassortant influenza viruses which contain all backbone segments from the same donor strain. The inventors have shown that this effect is due to the presence of backbone segments from two donor strains and does not require the presence of viral segments with specific mutations. Particularly good results are achieved, however, with influenza A strains in which the M genome segment has lysine in the position corresponding to amino acid 95 of SEQ 40 ID NO: 33 when aligned to SEQ ID NO: 33.

Reassortant influenza A viruses comprising the PB1 and PB2 segments from the same influenza strain (for example 105p30) are also advantageous because they showed a better rescue efficiency compared with influenza viruses in which 45 the PB1 and PB2 segments are from different viruses. The PB1 and PB2 segments of 105p30 have the sequence of SEQ ID NOs 18 and 19, respectively.

The inventors have also shown that some influenza virus strains can grow to higher viral titres in MDCK cells in the 50 same time and under the same growth conditions compared with A/Puerto Rico/8/34.

Variants of influenza donor strains which are derived from the donor strains of the invention or other known donor strains such A/PR/8/34 (wt PR8) can also be useful as donor strains. 55 These donor strains can grow to higher viral titres (in the same time and under the same growth conditions) compared to the donor strain from which they are derived. For example, the inventors have surprisingly discovered that passaging the A/PR/8/34 influenza strain several times in cell culture results in a virus strain (PR8-X) which grows to much higher viral titres compared to the original A/PR8/34 strain. Likewise, the inventors have found that passaging the A/New Caledonia/20/1999 strain several times in cells results in a strain (105p30) which grows to even higher viral titres compared to 65 the unpassaged A/New Caledonia/20/1999 strain in the same time and under the same growth conditions. Donor strain

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variants of the present invention will typically achieve viral titres which are at least 10%, at least 20%, at least 50%, at least 100%, at least 200%, at least 500% or at least 1000% higher under the same growth conditions and for the same time (for example 12 hours, 24 hours, 48 hours or 72 hours) compared to the viral titres obtained with the donor strain from which the variant was derived.

The segments of PR8-X have the sequences of SEQ ID NO: 11 (PB2), SEQ ID NO: 10 (PB1), SEQ ID NO: 9 (PA), SEQ ID NO: 12 (NP), SEQ ID NO: 13 (M), SEQ ID NO: 14 (NS), SEQ ID NO: 15 (HA) or SEQ ID NO: 16 (NA).

The segments of 105p30 have the sequences of SEQ ID NO: 19 (PB2), SEQ ID NO: 18 (PB1), SEQ ID NO: 17 (PA), SEQ ID NO: 20 (NP), SEQ ID NO: 21 (M), SEQ ID NO: 22 (NS), SEQ ID NO: 23 (HA) or SEQ ID NO: 24 (NA).

Influenza strains which contain one, two, three, four five, six or seven of the segments of the 105p30 or PR8-X strains can also be used as donor strains.

The invention can be practised with donor strains having a viral segment that has at least about 70%, at least about 75%, at least about 80%, at least about 85%, at least about 90%, at least about 95% or at least about 99% identity to a sequence of SEQ ID NOs 11-14 or 18-22. For example, due to the degeneracy of the genetic code, it is possible to have the same polypeptide encoded by several nucleic acids with different sequences. Thus, the invention may be practised with viral segments that encode the same polypeptides as the sequences of SEQ ID NOs 11-14 or 18-22. For example, the nucleic acid sequences of SEQ ID NOs: 3 and 28 have only 73% identity even though they encode the same viral protein.

The invention may also be practised with viral segments that encode polypeptides that have at least 80%, at least 85%, at least 90%, at least 95% or at least 99% identity to the polypeptide sequences encoded by SEQ ID NOs 11-14 or 18-22.

Variations in the DNA and the amino acid sequence may also stem from spontaneous mutations which can occur during passaging of the viruses. Such variant influenza strains can also be used in the invention.

Reassortant Viruses

The invention provides reassortant influenza viruses which comprise backbone segments from two or more influenza donor strains. The PB 1 and PB2 segments may be from the same donor strain.

The invention also provides reassortant influenza viruses comprising at least one backbone viral segment from an influenza virus strain that can grow to higher viral titres in MDCK cells in the same time and under the same growth conditions compared with A/Puerto Rico/8/34.

The invention provides reassortant influenza viruses comprising at least one backbone viral segment from the donor strains of the invention, e.g. a PR8-X or 105p30 strain. The reassortant influenza viruses of the invention can be reassortants between two, three or more different influenza strains provided that at least one viral segment is derived from a donor strain of the invention.

Influenza viruses are segmented negative strand RNA viruses. Influenza A and B viruses have eight segments (NP, M, NS, PA, PB1, HA and NA) whereas influenza C virus has seven. The reassortant viruses of the invention contain the backbone segments from two or more donor strains, or at least one (i.e. one, two, three, four, five or six) backbone viral segment from the donor strains of the invention. The backbone viral segments are those which do not encode HA or NA. Thus, backbone segments will typically encode the PB1, PB2, PA, NP, M1, M2, NS1 and NS2 polypeptides of the influenza virus. The reassortant viruses will not typically

contain the segments encoding HA and NA from the donor strains even though reassortant viruses which comprise either the HA or the NA but not both from the donor strains of the invention are also envisioned.

When the reassortant viruses of the invention are reassortants comprising the backbone segments from a single donor strain, the reassortant viruses will generally include segments from the donor strain and the vaccine strain in a ratio of 1:7, 2:6, 3:5, 4:4, 5:3, 6:2 or 7:1. Having a majority of segments from the donor strain, in particular a ratio of 6:2, is typical. When the reassortant viruses comprise backbone segments from two donor strains, the reassortant virus will generally include segments from the first donor strain, the seconds donor strain and the vaccine strain in a ratio of 1:1:6, 1:2:5, $_{15}$ 1:3:4, 1:4:3, 1:5:2, 1:6:1, 2:1:5, 2:2:4, 2:3:3, 2:4:2, 2:5:1, 3:1:2, 3:2:1, 4:1:3, 4:2:2, 4:3:1, 5:1:2, 5:2:1 or 6:1:1.

Preferably, the reassortant viruses do not contain the HA segment of the donor strain as this encodes the main vaccine antigens of the influenza virus and should therefore come 20 from the vaccine strain. The reassortant viruses of the invention therefore preferably have at least the HA segment and typically the HA and NA segments from the vaccine strain.

The invention also encompasses reassortant viruses which contain viral segments from more than one, for example two 25 or three different, donor strain(s) wherein at least one viral segment, preferably not HA, is derived from the PR8-X or 105p30 influenza strains. Such reassortant influenza viruses will typically contain the HA and/or NA segment from a vaccine strain. Where the reassortants contain viral segments 30 from more than one influenza donor strain, the further donor strain(s) can be any donor strain including the donor strains of the invention. For example, some of the viral segments may be derived from the A/PR/8/34 or AA/6/60 (A/Ann Arbor/6/ 60) influenza strains. Reassortants containing viral segments 35 from the AA/6/60 strain may be advantageous, for example, where the reassortant virus is to be used in a live attenuated influenza vaccine.

The invention also encompasses reassortants which comvided that the reassortant comprises a backbone according to the present invention. For example, the reassortant influenza viruses may comprise the HA segment from one donor strain and the NA segment from a different donor strain.

The reassortant viruses of the invention can grow to higher 45 viral titres than the wild-type vaccine strain from which some of the viral segment(s) of the reassortant virus are derived in the same time (for example 12 hours, 24 hours, 48 hours or 72 hours) and under the same growth conditions. The viral titre can be determined by standard methods known to those of 50 skill in the art. The reassortant viruses of the invention can achieve a viral titre which is at least 10% higher, at least 20% higher, at least 50% higher, at least 100% higher, at least 200% higher, at least 500% higher, or at least 1000% higher than the viral titre of the wild type vaccine strain in the same 55 time frame and under the same conditions.

The invention is suitable for reassorting pandemic as well as inter-pandemic (seasonal) influenza vaccine strains. The reassortant influenza strains may contain the influenza A virus HA subtypes H1, H2, H3, H4, H5, H6, H7, H8, H9, H10, 60 H11, H12, H13, H14, H15 or H16. They may contain the influenza A virus NA subtypes N1, N2, N3, N4, N5, N6, N7, N8 or N9. Where the vaccine strain used in the reassortant influenza viruses of the invention is a seasonal influenza strain, the vaccine strain may have a H1 or H3 subtype. In one 65 aspect of the invention the vaccine strain is a H1N1 or H₃N₂ strain.

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The vaccine strains for use in the invention may also be pandemic strains or potentially pandemic strains. The characteristics of an influenza strain that give it the potential to cause a pandemic outbreak are: (a) it contains a new hemagglutinin compared to the hemagglutinins in currently-circulating human strains, i.e. one that has not been evident in the human population for over a decade (e.g. H2), or has not previously been seen at all in the human population (e.g. H5, H6 or H9, that have generally been found only in bird populations), such that the human population will be immunologically naïve to the strain's hemagglutinin; (b) it is capable of being transmitted horizontally in the human population; and (c) it is pathogenic to humans. A vaccine strain with H5 hemagglutinin type is preferred where the reassortant virus is used in vaccines for immunizing against pandemic influenza, such as a H5N1 strain. Other possible strains include H5N3, H9N2, H2N2, H7N1 and H7N7, and any other emerging potentially pandemic strains. The invention is particularly suitable for producing reassortant viruses for use in vaccine for protecting against potential pandemic virus strains that can or have spread from a non-human animal population to humans, for example a swine-origin H1N1 influenza strain.

The reassortant influenza strain of the invention may comprise the HA segment and/or the NA segment from an A/California/4/09 strain. Thus, for instance, the HA gene segment may encode a H1 hemagglutinin which is more closely related to SEQ ID NO: 32 than to SEQ ID NO: 25 (i.e. has a higher degree sequence identity when compared to SEO ID NO: 32 than to SEQ ID NO: 25 using the same algorithm and parameters). SEQ ID NOs: 32 and 25 are 80% identical. Similarly, the NA gene may encode a N1 neuraminidase which is more closely related to SEQ ID NO: 27 than to SEQ ID NO: 26. SEQ ID NOs: 27 and 26 are 82% identical.

Strains which can be used as vaccine strains include strains which are resistant to antiviral therapy (e.g. resistant to oseltamivir [6] and/or zanamivir), including resistant pandemic strains [7].

The choice of donor strain for use in the methods of the prise viral segments from more than one vaccine strain pro- 40 invention can depend on the vaccine strain which is to be reassorted. As reassortants between evolutionary distant strains might not replicate well in cell culture, it is possible that the donor strain and the vaccine strain have the same HA and/or NA subtype. In other embodiments, however, the vaccine strain and the donor strain can have different HA and/or NA subtypes, and this arrangement can facilitate selection for reassortant viruses that contain the HA and/or NA segment from the vaccine strain. Therefore, although the 105p30 and PR8-X strains contain the H1 influenza subtype these donor strains can be used for vaccine strains which do not contain the H1 influenza subtype.

Reassortants of the donor strains of the invention wherein the HA and/or NA segment has been changed to another subtype can also be used. The H1 influenza subtype of the 105p30 or PR8-X strain may be changed, for example, to a H3 or H5 subtype.

Thus, the invention encompasses an influenza A virus which comprises one, two, three, four, five, six or seven viral segments from the 105p30 or PR8-X strains of the invention and a HA segment which is not of the H1 subtype. The reassortant donor strains may further comprise an NA segment which is not of the N1 subtype. Suitable techniques for reassorting the donor strains will be evident to those of skill in the art.

The invention also encompasses reassortant donor strains which comprise at least one, at least two, at least three, at least four, at least five, at least six or at least seven viral segments

from the 105p30 or PR8-X strains of the invention and a H1 HA segment which is derived from a different influenza

Reassortant viruses which contain an NS segment that does not encode a functional NS protein are also within the scope 5 of the present invention. NS 1 knockout mutants are described in reference 8. These NS1-mutant virus strains are particularly suitable for preparing live attenuated influenza vaccines.

The 'second influenza strain' used in the methods of the invention is different to the donor strain which is used. Reverse Genetics

The invention is particularly suitable for producing reassortant influenza virus strains through reverse genetics techniques. In these techniques, the viruses are produced in culture hosts using an expression system.

In one aspect, the expression system may encode the PB1 and PB2 segments from the same donor strain. In this aspect of the invention, the system may encode at least one (i.e. one, two three or four) of the segments NP, M, NS and/or PA from another influenza donor strain.

In another aspect, the system may encode 1 or more (e.g. 1, 2, 3, 4, 5 or 6) genome segments from the PR8-X strain, but usually this/these will not include the PR8-X HA segment and usually will not include the PR8-X NA segment. Thus the PB1 and/or PB2 (possibly all six) from PR8-X.

The system may encode 1 or more (e.g. 1, 2, 3, 4, 5 or 6) genome segments from the 105p30 strain, but usually this/ these will not include the 105p30 HA segment and usually will not include the 105p30 NA segment. Thus the system 30 may encode at least one of segments NP, M, NS, PA, PB1 and/or PB2 (possibly all six) from 105p30.

Reverse genetics for influenza A and B viruses can be practised with 12 plasmids to express the four proteins required to initiate replication and transcription (PB 1, PB2, 35 PA and nucleoprotein) and all eight viral genome segments. To reduce the number of constructs, however, a plurality of RNA polymerase I transcription cassettes (for viral RNA synthesis) can be included on a single plasmid (e.g. sequences encoding 1, 2, 3, 4, 5, 6, 7 or all 8 influenza vRNA segments), 40 and a plurality of protein-coding regions with RNA polymerase II promoters on another plasmid (e.g. sequences encoding 1, 2, 3, 4, 5, 6, 7 or 8 influenza mRNA transcripts) [9]. It is also possible to include one or more influenza vRNA segments under control of a pol I promoter and one or more 45 influenza protein coding regions under control of another promoter, in particular a pol II promoter, on the same plasmid. This is preferably done by using bi-directional plasmids.

Preferred aspects of the reference 9 method involve: (a) PB 1, PB2 and PA mRNA-encoding regions on a single expres- 50 sion construct; and (b) all 8 vRNA encoding segments on a single expression construct. Including the neuraminidase (NA) and hemagglutinin (HA) segments on one expression construct and the six other viral segments on another expression construct is particularly preferred as newly emerging 55 influenza virus strains usually have mutations in the NA and/ or HA segments. Therefore, the advantage of having the HA and/or NA segments on a separate expression construct is that only the vector comprising the HA and NA sequence needs to be replaced. Thus, in one aspect of the invention the NA 60 and/or HA segments of the vaccine strain may be included on one expression construct and the vRNA encoding segments from the donor strain(s) of the invention, excluding the HA and/or NA segment(s), are included on a different expression construct. The invention thus provides an expression con- 65 struct comprising one, two, three, four, five or six vRNA encoding backbone viral segments of a donor strain of the

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invention. The expression construct may not comprise HA and/or NA viral segments that produce a functional HA and/ or NA protein.

Known reverse genetics systems involve expressing DNA molecules which encode desired viral RNA (vRNA) molecules from pol I promoters, bacterial RNA polymerase promoters, bacteriophage polymerase promoters, etc. As influenza viruses require the presence of viral polymerase to complete the life cycle, systems may also provide these proteins e.g. the system further comprises DNA molecules that encode viral polymerase proteins such that expression of both types of DNA leads to assembly of a complete infectious virus. It is also possible to supply the viral polymerase as a protein.

Where reverse genetics is used for the expression of influenza vRNA, it will be evident to the person skilled in the art that precise spacing of the sequence elements with reference to each other is important for the polymerase to initiate replication. It is therefore important that the DNA molecule 20 encoding the viral RNA is positioned correctly between the pol I promoter and the termination sequence, but this positioning is well within the capabilities of those who work with reverse genetics systems.

In order to produce a recombinant virus, a cell must express system may encode at least one of segments NP, M, NS, PA, 25 all segments of the viral genome which are necessary to assemble a virion. DNA cloned into the expression constructs of the present invention preferably provides all of the viral RNA and proteins, but it is also possible to use a helper virus to provide some of the RNA and proteins, although systems which do not use a helper virus are preferred. As the influenza virus is a segmented virus, the viral genome will usually be expressed using more than one expression construct in the methods of the invention. It is also envisioned, however, to combine one or more segments or even all segments of the viral genome on a single expression construct.

> In some embodiments an expression construct will also be included which leads to expression of an accessory protein in the host cell. For instance, it can be advantageous to express a non-viral serine protease (e.g. trypsin) as part of a reverse genetics system.

Expression Constructs

Expression constructs used in the expression systems of the invention may be uni-directional or bi-directional expression constructs. Where more than one transgene is used in the methods (whether on the same or different expression constructs) it is possible to use uni-directional and/or bi-directional expression.

As influenza viruses require a protein for infectivity, it is generally preferred to use bi-directional expression constructs as this reduces the total number of expression constructs required by the host cell. Thus, the method of the invention may utilise at least one bi-directional expression construct wherein a gene or cDNA is located between an upstream pol II promoter and a downstream non-endogenous pol I promoter. Transcription of the gene or cDNA from the pol II promoter produces capped positive-sense viral mRNA which can be translated into a protein, while transcription from the non-endogenous pol I promoter produces negativesense vRNA. The bi-directional expression construct may be a bi-directional expression vector.

Bi-directional expression constructs contain at least two promoters which drive expression in different directions (i.e. both 5' to 3' and 3' to 5') from the same construct. The two promoters can be operably linked to different strands of the same double stranded DNA. Preferably, one of the promoters is a pol I promoter and at least one of the other promoters is a pol II promoter. This is useful as the pol I promoter can be

used to express uncapped vRNAs while the pol II promoter can be used to transcribe mRNAs which can subsequently be translated into proteins, thus allowing simultaneous expression of RNA and protein from the same construct. Where more than one expression construct is used within an expression system, the promoters may be a mixture of endogenous and non-endogenous promoters.

The pol I and pol II promoters used in the expression constructs may be endogenous to an organism from the same taxonomic order from which the host cell is derived. Alternatively, the promoters can be derived from an organism in a different taxonomic order than the host cell. The term "order" refers to conventional taxonomic ranking, and examples of orders are primates, rodentia, carnivora, marsupialia, cetacean, etc. Humans and chimpanzees are in the same taxonomic order (primates), but humans and dogs are in different orders (primates vs. carnivora). For example, the human pol I promoter can be used to express viral segments in canine cells (e.g. MDCK cells).

The expression construct will typically include an RNA 20 transcription termination sequence. The termination sequence may be an endogenous termination sequence or a termination sequence which is not endogenous to the host cell. Suitable termination sequences will be evident to those of skill in the art and include, but are not limited to, RNA 25 polymerase I transcription termination sequence, RNA polymerase II transcription termination sequence, and ribozymes. Furthermore, the expression constructs may contain one or more polyadenylation signals for mRNAs, particularly at the end of a gene whose expression is controlled by a pol II 30 promoter.

An expression system may contain at least two, at least three, at least four, at least five, at least six, at least seven, at least eight, at least nine, at least ten, at least eleven or at least twelve expression constructs.

An expression construct may be a vector, such as a plasmid or other episomal construct. Such vectors will typically comprise at least one bacterial and/or eukaryotic origin of replication. Furthermore, the vector may comprise a selectable marker which allows for selection in prokaryotic and/or 40 eukaryotic cells. Examples of such selectable markers are genes conferring resistance to antibiotics, such as ampicillin or kanamycin. The vector may further comprise one or more multiple cloning sites to facilitate cloning of a DNA sequence.

As an alternative, an expression construct may be a linear expression construct. Such linear expression constructs will typically not contain any amplification and/or selection sequences. However, linear constructs comprising such amplification and/or selection sequences are also within the 50 scope of the present invention. Reference 10 describes a linear expression construct which describes individual linear expression constructs for each viral segment. It is also possible to include more than one, for example two, three four, five or six viral segments on the same linear expression construct. Such a system has been described, for example, in reference 11.

Expression constructs can be generated using methods known in the art. Such methods were described, for example, in reference 12. Where the expression construct is a linear 60 expression construct, it is possible to linearise it before introduction into the host cell utilising a single restriction enzyme site. Alternatively, it is possible to excise the expression construct from a vector using at least two restriction enzyme sites. Furthermore, it is also possible to obtain a linear expression 65 construct by amplifying it using a nucleic acid amplification technique (e.g. by PCR).

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The expression constructs used in the systems of the invention may be non-bacterial expression constructs. This means that the construct can drive expression in a eukaryotic cell of viral RNA segments encoded therein, but it does not include components which would be required for propagation of the construct in bacteria. Thus the construct will not include a bacterial origin of replication (ori), and usually will not include a bacterial selection marker (e.g. an antibiotic resistance marker). Such expression constructs are described in reference 13 which is incorporated by reference.

The expression constructs may be prepared by chemical synthesis. The expression constructs may either be prepared entirely by chemical synthesis or in part. Suitable methods for preparing expression constructs by chemical synthesis are described, for example, in reference 13 which is incorporated by reference.

The expression constructs of the invention can be introduced into host cells using any technique known to those of skill in the art. For example, expression constructs of the invention can be introduced into host cells by employing electroporation, DEAE-dextran, calcium phosphate precipitation, liposomes, microinjection, or microparticle-bombardment.

Cells

The culture host for use in the present invention can be any eukaryotic cell that can produce the virus of interest. The invention will typically use a cell line although, for example, primary cells may be used as an alternative. The cell will typically be mammalian. Suitable mammalian cells include, but are not limited to, hamster, cattle, primate (including humans and monkeys) and dog cells. Various cell types may be used, such as kidney cells, fibroblasts, retinal cells, lung cells, etc. Examples of suitable hamster cells are the cell lines having the names BHK21 or HKCC. Suitable monkey cells are e.g. African green monkey cells, such as kidney cells as in the Vero cell line [14-15]. Suitable dog cells are e.g. kidney cells, as in the CLDK and MDCK cell lines.

Further suitable cells include, but are not limited to: CHO; 293T; BHK; MRC 5; PER.C6 [16]; FRhL2; WI-38; etc. Suitable cells are widely available e.g. from the American Type Cell Culture (ATCC) collection [17], from the Coriell Cell Repositories [18], or from the European Collection of Cell Cultures (ECACC). For example, the ATCC supplies various different Vero cells under catalogue numbers CCL 81, CCL 81.2, CRL 1586 and CRL-1587, and it supplies MDCK cells under catalogue number CCL 34. PERC6 is available from the ECACC under deposit number 96022940.

Preferred cells for use in the invention are MDCK cells [19-20], derived from Madill Darby canine kidney. The original MDCK cells are available from the ATCC as CCL 34. It is preferred that derivatives of MDCK cells are used. Such derivatives were described, for instance, in reference 19 which discloses MDCK cells that were adapted for growth in suspension culture ('MDCK 33016' or '33016-PF', deposited as DSM ACC 2219; see also ref. 19). Furthermore, reference 21 discloses MDCK-derived cells that grow in suspension in serum free culture ('B-702', deposited as FERM BP-7449). In some embodiments, the MDCK cell line used may be tumorigenic. It is also envisioned to use non-tumorigenic MDCK cells. For example, reference 22 discloses non tumorigenic MDCK cells, including 'MDCK-S' (ATCC PTA-6500), 'MDCK-SF101' (ATCC PTA-6501), 'MDCK-SF102' (ATCC PTA-6502) and 'MDCK-SF103' (ATCC PTA-6503). Reference 23 discloses MDCK cells with high susceptibility to infection, including 'MDCK.5F1' cells (ATCC CRL 12042).

It is possible to use a mixture of more than one cell type to practise the methods of the present invention. However, it is preferred that the methods of the invention are practised with a single cell type e.g. with monoclonal cells. Preferably, the cells used in the methods of the present invention are from a single cell line. Furthermore, the same cell line may be used for reassorting the virus and for any subsequent propagation of the virus.

Preferably, the cells are cultured in the absence of serum, to avoid a common source of contaminants. Various serum-free 10 media for eukaryotic cell culture are known to the person skilled in the art (e.g. Iscove's medium, ultra CHO medium (BioWhittaker), EX-CELL (JRH Biosciences)). Furthermore, protein-free media may be used (e.g. PF-CHO (JRH Biosciences)). Otherwise, the cells for replication can also be 15 cultured in the customary serum-containing media (e.g. MEM or DMEM medium with 0.5% to 10% of fetal calf serum).

The cells may be in adherent culture or in suspension. Conventional Reassortment

Traditionally, influenza viruses are reassorted by co-infecting a culture host, usually eggs, with a donor strain and a vaccine strain. Reassortant viruses are selected by adding antibodies with specificity for the HA and/or NA proteins of the donor strain in order to select for reassortant viruses that 25 contain the vaccine strain's HA and/or NA proteins. Over several passages of this treatment one can select for fast growing reassortant viruses containing the vaccine strain's HA and/or NA segments.

The invention is suitable for use in these methods. It can be 30 easier to use vaccine strains with a different HA and/or NA subtype compared to the donor strain(s) as this facilitates selection for reassortant viruses. It is also possible, however, to use vaccine strains with the same HA and/or NA subtype as the donor strain(s) and in some aspects of the invention this preferred. In this case, antibodies with preferential specificity for the HA and/or NA proteins of the donor strain(s) should be available.

Virus Preparation

In one embodiment, the invention provides a method for 40 producing influenza viruses comprising steps of (a) infecting a culture host with a reassortant virus of the invention; (b) culturing the host from step (a) to produce the virus; and optionally (c) purifying the virus obtained in step (b).

The culture host may be cells or embryonated hen eggs. 45 Where cells are used as a culture host in this aspect of the invention, it is known that cell culture conditions (e.g. temperature, cell density, pH value, etc.) are variable over a wide range subject to the cell line and the virus employed and can be adapted to the requirements of the application. The following information therefore merely represents guidelines.

As mentioned above, cells are preferably cultured in serum-free or protein-free media.

Multiplication of the cells can be conducted in accordance with methods known to those of skill in the art. For example, 55 the cells can be cultivated in a perfusion system using ordinary support methods like centrifugation or filtration. Moreover, the cells can be multiplied according to the invention in a fed-batch system before infection. In the context of the present invention, a culture system is referred to as a fedbatch system in which the cells are initially cultured in a batch system and depletion of nutrients (or part of the nutrients) in the medium is compensated by controlled feeding of concentrated nutrients. It can be advantageous to adjust the pH value of the medium during multiplication of cells before infection 65 to a value between pH 6.6 and pH 7.8 and especially between a value between pH 7.2 and pH 7.3. Culturing of cells pref-

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erably occurs at a temperature between 30 and 40° C. When culturing the infected cells (step ii), the cells are preferably cultured at a temperature of between 30° C. and 36° C. or between 32° C. and 34° C. or at 33° C. This is particularly preferred, as it has been shown that incubation of infected cells in this temperature range results in production of a virus that results in improved efficacy when formulated into a vaccine [241].

Oxygen partial pressure can be adjusted during culturing before infection preferably at a value between 25% and 95% and especially at a value between 35% and 60%. The values for the oxygen partial pressure stated in the context of the invention are based on saturation of air. Infection of cells occurs at a cell density of preferably about 8–25×10⁵ cells/mL in the batch system or preferably about 5–20×10⁶ cells/mL in the perfusion system. The cells can be infected with a viral dose (MOI value, "multiplicity of infection"; corresponds to the number of virus units per cell at the time of infection) between 10⁻⁸ and 10, preferably between 0.0001 and 0.5.

Virus may be grown on cells in adherent culture or in suspension. Microcarrier cultures can be used. In some embodiments, the cells may thus be adapted for growth in suspension.

The methods according to the invention also include harvesting and isolation of viruses or the proteins generated by them. During isolation of viruses or proteins, the cells are separated from the culture medium by standard methods like separation, filtration or ultrafiltration. The viruses or the proteins are then concentrated according to methods sufficiently known to those skilled in the art, like gradient centrifugation, filtration, precipitation, chromatography, etc., and then purified. It is also preferred according to the invention that the viruses are inactivated during or after purification. Virus inactivation can occur, for example, by β -propiolactone or formaldehyde at any point within the purification process.

The culture host may be eggs. The current standard method for influenza virus growth for vaccines uses embryonated SPF hen eggs, with virus being purified from the egg contents (allantoic fluid). It is also possible to passage a virus through eggs and subsequently propagate it in cell culture and vice versa.

Vaccine

The invention utilises virus produced according to the method to produce vaccines.

Vaccines (particularly for influenza virus) are generally based either on live virus or on inactivated virus. Inactivated vaccines may be based on whole virions, 'split' virions, or on purified surface antigens. Antigens can also be presented in the form of virosomes. The invention can be used for manufacturing any of these types of vaccine.

Where an inactivated virus is used, the vaccine may comprise whole virion, split virion, or purified surface antigens (for influenza, including hemagglutinin and, usually, also including neuraminidase). Chemical means for inactivating a virus include treatment with an effective amount of one or more of the following agents: detergents, formaldehyde, β -propiolactone, methylene blue, psoralen, carboxyfullerene (C60), binary ethylamine, acetyl ethyleneimine, or combinations thereof. Non-chemical methods of viral inactivation are known in the art, such as for example UV light or gamma irradiation.

Virions can be harvested from virus-containing fluids, e.g. allantoic fluid or cell culture supernatant, by various methods. For example, a purification process may involve zonal centrifugation using a linear sucrose gradient solution that

includes detergent to disrupt the virions. Antigens may then be purified, after optional dilution, by diafiltration.

Split virions are obtained by treating purified virions with detergents (e.g. ethyl ether, polysorbate 80, deoxycholate, tri-N-butyl phosphate, Triton X-100, Triton N101, cetyltrimethylammonium bromide, Tergitol NP9, etc.) to produce subvirion preparations, including the 'Tween-ether' splitting process. Methods of splitting influenza viruses, for example are well known in the art e.g. see refs. 25-26, etc. Splitting of the virus is typically carried out by disrupting or fragmenting whole virus, whether infectious or non-infectious with a disrupting concentration of a splitting agent. The disruption results in a full or partial solubilisation of the virus proteins, altering the integrity of the virus. Preferred splitting agents are non-ionic and ionic (e.g. cationic) surfactants e.g. alkylglycosides, alkylthioglycosides, acyl sugars, sulphobetaines, betains, polyoxyethylenealkylethers, N,N-dialkyl-Glucamides, Hecameg, alkylphenoxy-polyethoxyethanols, NP9, quaternary ammonium compounds, sarcosyl, CTABs (cetyl 20 trimethyl ammonium bromides), tri-N-butyl phosphate, Cetavlon, myristyltrimethylammonium salts, lipofectin, lipofectamine, and DOT-MA, the octyl- or nonylphenoxy polyoxyethanols (e.g. the Triton surfactants, such as Triton X-100 or Triton N101), polyoxyethylene sorbitan esters (the Tween 25 surfactants), polyoxyethylene ethers, polyoxyethlene esters, etc. One useful splitting procedure uses the consecutive effects of sodium deoxycholate and formaldehyde, and splitting can take place during initial virion purification (e.g. in a sucrose density gradient solution). Thus a splitting process can involve clarification of the virion-containing material (to remove non-virion material), concentration of the harvested virions (e.g. using an adsorption method, such as CaHPO₄ adsorption), separation of whole virions from non-virion material, splitting of virions using a splitting agent in a density gradient centrifugation step (e.g. using a sucrose gradient that contains a splitting agent such as sodium deoxycholate), and then filtration (e.g. ultrafiltration) to remove undesired materials. Split virions can usefully be resuspended in sodium 40 phosphate-buffered isotonic sodium chloride solution. Examples of split influenza vaccines are the BEGRIVACTM, FLUARIXTM, FLUZONETM and FLUSHIELDTM products.

Purified influenza virus surface antigen vaccines comprise the surface antigens hemagglutinin and, typically, also 45 neuraminidase. Processes for preparing these proteins in purified form are well known in the art. The FLUVIRINTM, AGRIPPALTM and INFLUVACTM products are influenza subunit vaccines.

Another form of inactivated antigen is the virosome [27] (nucleic acid free viral-like liposomal particles). Virosomes can be prepared by solubilization of virus with a detergent followed by removal of the nucleocapsid and reconstitution of the membrane containing the viral glycoproteins. An alternative method for preparing virosomes involves adding viral membrane glycoproteins to excess amounts of phospholipids, to give liposomes with viral proteins in their membrane.

The methods of the invention may also be used to produce live vaccines. Such vaccines are usually prepared by purifying virions from virion-containing fluids. For example, the fluids may be clarified by centrifugation, and stabilized with buffer (e.g. containing sucrose, potassium phosphate, and monosodium glutamate). Various forms of influenza virus vaccine are currently available (e.g. see chapters 17 & 18 of reference 28). Live virus vaccines include MedImmune's FLUMISTTM product (trivalent live virus vaccine).

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The virus may be attenuated. The virus may be temperature-sensitive. The virus may be cold-adapted. These three features are particularly useful when using live virus as an antigen.

HA is the main immunogen in current inactivated influenza vaccines, and vaccine doses are standardised by reference to HA levels, typically measured by SRID. Existing vaccines typically contain about 15 μg of HA per strain, although lower doses can be used e.g. for children, or in pandemic situations, or when using an adjuvant. Fractional doses such as $^{1}\!/_{2}$ (i.e. $7.5\,\mu g$ HA per strain), $^{1}\!/_{4}$ and $^{1}\!/_{8}$ have been used, as have higher doses (e.g. $3\times$ or $9\times$ doses [29,30]). Thus vaccines may include between 0.1 and 150 μg of HA per influenza strain, preferably between 0.1 and 50 μg e.g. 0.1-20 μg , 0.1-15 μg , 0.1-10 μg , 0.5-5 μg , etc. Particular doses include e.g. about 45, about 30, about 15, about 10, about 7.5, about 5, about 3.8, about 3.75, about 1.9, about 1.5, etc. per strain.

For live vaccines, dosing is measured by median tissue culture infectious dose ($TCID_{50}$) rather than HA content, and a $TCID_{50}$ of between 10^6 and 10^8 (preferably between $10^{6.5}$ - $10^{7.5}$) per strain is typical.

Influenza strains used with the invention may have a natural HA as found in a wild-type virus, or a modified HA. For instance, it is known to modify HA to remove determinants (e.g. hyper-basic regions around the HA1/HA2 cleavage site) that cause a virus to be highly pathogenic in avian species. The use of reverse genetics facilitates such modifications.

As well as being suitable for immunizing against interpandemic strains, the compositions of the invention are particularly useful for immunizing against pandemic or potentially-pandemic strains. The invention is suitable for vaccinating humans as well as non-human animals.

Other strains whose antigens can usefully be included in the compositions are strains which are resistant to antiviral therapy (e.g. resistant to oseltamivir [31] and/or zanamivir), including resistant pandemic strains [32].

Compositions of the invention may include antigen(s) from one or more (e.g. 1, 2, 3, 4 or more) influenza virus strains, including influenza A virus and/or influenza B virus provided that at least one influenza strain is a reassortant influenza strain of the invention. Compositions wherein at least two, at least three or all of the antigens are from reassortant influenza strains of the invention are also envisioned. Where a vaccine includes more than one strain of influenza, the different strains are typically grown separately and are mixed after the viruses have been harvested and antigens have been prepared. Thus a process of the invention may include the step of mixing antigens from more than one influenza strain. A trivalent vaccine is typical, including antigens from two influenza A virus strains and one influenza B virus strain. A tetravalent vaccine is also useful [33], including antigens from two influenza A virus strains and two influenza B virus strains, or three influenza A virus strains and one influenza B virus strain. Pharmaceutical Compositions

Vaccine compositions manufactured according to the invention are pharmaceutically acceptable. They usually include components in addition to the antigens e.g. they typically include one or more pharmaceutical carrier(s) and/or excipient(s). As described below, adjuvants may also be included. A thorough discussion of such components is available in reference 34.

Vaccine compositions will generally be in aqueous form. However, some vaccines may be in dry form, e.g. in the form of injectable solids or dried or polymerized preparations on a patch.

Vaccine compositions may include preservatives such as thiomersal or 2-phenoxyethanol. It is preferred, however, that

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the vaccine should be substantially free from (i.e. less than 5 μg/ml) mercurial material e.g. thiomersal-free [Error! Bookmark not defined, 35]. Vaccines containing no mercury are more preferred. An α -tocopherol succinate can be included as an alternative to mercurial compounds [Error! Bookmark not 5 defined.]. Preservative-free vaccines are particularly pre-

To control tonicity, it is preferred to include a physiological salt, such as a sodium salt. Sodium chloride (NaCl) is preferred, which may be present at between 1 and 20 mg/ml. 10 Other salts that may be present include potassium chloride, potassium dihydrogen phosphate, disodium phosphate dehydrate, magnesium chloride, calcium chloride, etc.

Vaccine compositions will generally have an osmolality of between 200 mOsm/kg and 400 mOsm/kg, preferably 15 between 240-360 mOsm/kg, and will more preferably fall within the range of 290-310 mOsm/kg. Osmolality has previously been reported not to have an impact on pain caused by vaccination [36], but keeping osmolality in this range is nevertheless preferred.

Vaccine compositions may include one or more buffers. Typical buffers include: a phosphate buffer; a Tris buffer; a borate buffer; a succinate buffer; a histidine buffer (particularly with an aluminum hydroxide adjuvant); or a citrate buffer. Buffers will typically be included in the 5-20 mM 25

The pH of a vaccine composition will generally be between 5.0 and 8.1, and more typically between 6.0 and 8.0 e.g. 6.5 and 7.5, or between 7.0 and 7.8. A process of the invention may therefore include a step of adjusting the pH of the bulk 30 vaccine prior to packaging.

The vaccine composition is preferably sterile. The vaccine composition is preferably non-pyrogenic e.g. containing <1 EU (endotoxin unit, a standard measure) per dose, and preferably <0.1 EU per dose. The vaccine composition is prefer- 35 ably gluten-free.

Vaccine compositions of the invention may include detergent e.g. a polyoxyethylene sorbitan ester surfactant (known as 'Tweens'), an octoxynol (such as octoxynol-9 (Triton X-100) or t-octylphenoxypolyethoxyethanol), a cetyl trim- 40 ethyl ammonium bromide ('CTAB'), or sodium deoxycholate, particularly for a split or surface antigen vaccine. The detergent may be present only at trace amounts. Thus the vaccine may include less than 1 mg/ml of each of octoxynol-10 and polysorbate 80. Other residual components in trace 45 amounts could be antibiotics (e.g. neomycin, kanamycin, polymyxin B).

A vaccine composition may include material for a single immunisation, or may include material for multiple immunisations (i.e. a 'multidose' kit). The inclusion of a preservative 50 is preferred in multidose arrangements. As an alternative (or in addition) to including a preservative in multidose compositions, the compositions may be contained in a container having an aseptic adaptor for removal of material.

Influenza vaccines are typically administered in a dosage 55 volume of about 0.5 ml, although a half dose (i.e. about 0.25 ml) may be administered to children.

Compositions and kits are preferably stored at between 2° C. and 8° C. They should not be frozen. They should ideally be kept out of direct light. Host Cell DNA

Where virus has been isolated and/or grown on a cell line, it is standard practice to minimize the amount of residual cell line DNA in the final vaccine, in order to minimize any potential oncogenic activity of the DNA.

Thus a vaccine composition prepared according to the invention preferably contains less than 10 ng (preferably less

than 1 ng, and more preferably less than 100 pg) of residual host cell DNA per dose, although trace amounts of host cell DNA may be present.

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It is preferred that the average length of any residual host cell DNA is less than 500 bp e.g. less than 400 bp, less than 300 bp, less than 200 bp, less than 100 bp, etc.

Contaminating DNA can be removed during vaccine preparation using standard purification procedures e.g. chromatography, etc. Removal of residual host cell DNA can be enhanced by nuclease treatment e.g. by using a DNase. A convenient method for reducing host cell DNA contamination is disclosed in references 37 & 38, involving a two-step treatment, first using a DNase (e.g. Benzonase), which may be used during viral growth, and then a cationic detergent (e.g. CTAB), which may be used during virion disruption. Treatment with an alkylating agent, such as β -propiolactone, can also be used to remove host cell DNA, and advantageously may also be used to inactivate virions [39]. Adjuvants

Compositions of the invention may advantageously include an adjuvant, which can function to enhance the immune responses (humoral and/or cellular) elicited in a subject who receives the composition. Preferred adjuvants comprise oil-in-water emulsions. Various such adjuvants are known, and they typically include at least one oil and at least one surfactant, with the oil(s) and surfactant(s) being biodegradable (metabolisable) and biocompatible. The oil droplets in the emulsion are generally less than 5 µm in diameter, and ideally have a sub-micron diameter, with these small sizes being achieved with a microfluidiser to provide stable emulsions. Droplets with a size less than 220 nm are preferred as they can be subjected to filter sterilization.

The emulsion can comprise oils such as those from an animal (such as fish) or vegetable source. Sources for vegetable oils include nuts, seeds and grains. Peanut oil, soybean oil, coconut oil, and olive oil, the most commonly available, exemplify the nut oils. Jojoba oil can be used e.g. obtained from the jojoba bean. Seed oils include safflower oil, cottonseed oil, sunflower seed oil, sesame seed oil and the like. In the grain group, corn oil is the most readily available, but the oil of other cereal grains such as wheat, oats, rye, rice, teff, triticale and the like may also be used. 6-10 carbon fatty acid esters of glycerol and 1,2-propanediol, while not occurring naturally in seed oils, may be prepared by hydrolysis, separation and esterification of the appropriate materials starting from the nut and seed oils. Fats and oils from mammalian milk are metabolizable and may therefore be used in the practice of this invention. The procedures for separation, purification, saponification and other means necessary for obtaining pure oils from animal sources are well known in the art. Most fish contain metabolizable oils which may be readily recovered. For example, cod liver oil, shark liver oils, and whale oil such as spermaceti exemplify several of the fish oils which may be used herein. A number of branched chain oils are synthesized biochemically in 5-carbon isoprene units and are generally referred to as terpenoids. Shark liver oil contains a branched, unsaturated terpenoids known as squalene, 2,6,10,15,19,23-hexamethyl-2,6,10,14,18,22-tetracosahexaene, which is particularly preferred herein. Squalane, the saturated analog to squalene, is also a preferred oil. Fish oils, including squalene and squalane, are readily available from commercial sources or may be obtained by methods known in the art. Another preferred oil is α -tocopherol (see below).

Mixtures of oils can be used.

Surfactants can be classified by their 'HLB' (hydrophile/ lipophile balance). Preferred surfactants of the invention have

a HLB of at least 10, preferably at least 15, and more preferably at least 16. The invention can be used with surfactants including, but not limited to: the polyoxyethylene sorbitan esters surfactants (commonly referred to as the Tweens), especially polysorbate 20 and polysorbate 80; copolymers of 5 ethylene oxide (EO), propylene oxide (PO), and/or butylene oxide (BO), sold under the DOWFAXTM tradename, such as linear EO/PO block copolymers; octoxynols, which can vary in the number of repeating ethoxy(oxy-1,2-ethanediyl) groups, with octoxynol-9 (Triton X-100, or t-octylphenoxy- 10 polyethoxyethanol) being of particular interest; (octylphenoxy)polyethoxyethanol (IGEPAL CA-630/NP-40); phospholipids such as phosphatidylcholine (lecithin); nonylphenol ethoxylates, such as the Tergitol™ NP series; polyoxyethylene fatty ethers derived from lauryl, cetyl, 15 stearyl and oleyl alcohols (known as Brij surfactants), such as triethyleneglycol monolauryl ether (Brij 30); and sorbitan esters (commonly known as the SPANs), such as sorbitan trioleate (Span 85) and sorbitan monolaurate. Non-ionic surfactants are preferred. Preferred surfactants for including in 20 the emulsion are Tween 80 (polyoxyethylene sorbitan monooleate), Span 85 (sorbitan trioleate), lecithin and Triton X-100.

Mixtures of surfactants can be used e.g. Tween 80/Span 85 mixtures. A combination of a polyoxyethylene sorbitan ester 25 such as polyoxyethylene sorbitan monooleate (Tween 80) and an octoxynol such as t-octylphenoxypolyethoxyethanol (Triton X-100) is also suitable. Another useful combination comprises laureth 9 plus a polyoxyethylene sorbitan ester and/or an octoxynol.

Preferred amounts of surfactants (% by weight) are: polyoxyethylene sorbitan esters (such as Tween 80) 0.01 to 1%, in particular about 0.1%; octyl- or nonylphenoxy polyoxyethanols (such as Triton X-100, or other detergents in the Triton series) 0.001 to 0.1%, in particular 0.005 to 0.02%; polyoxy-35 ethylene ethers (such as laureth 9) 0.1 to 20%, preferably 0.1 to 10% and in particular 0.1 to 1% or about 0.5%.

Where the vaccine contains a split virus, it is preferred that it contains free surfactant in the aqueous phase. This is advantageous as the free surfactant can exert a 'splitting effect' on 40 the antigen, thereby disrupting any unsplit virions and/or virion aggregates that might otherwise be present. This can improve the safety of split virus vaccines [40].

Preferred emulsions have an average droplets size of <1 μ m e.g. \leq 750 nm, \leq 500 nm, \leq 400 nm, \leq 300 nm, \leq 250 nm, \leq 220 45 nm, \leq 200 nm, or smaller. These droplet sizes can conveniently be achieved by techniques such as microfluidisation.

Specific oil-in-water emulsion adjuvants useful with the invention include, but are not limited to:

A submicron emulsion of squalene, Tween 80, and Span 85. The composition of the emulsion by volume can be about 5% squalene, about 0.5% polysorbate 80 and about 0.5% Span 85. In weight terms, these ratios become 4.3% squalene, 0.5% polysorbate 80 and 0.48% Span 85. This adjuvant is known as 'MF59' [41-42], as described in more detail in Chapter 10 of ref. 43 and chapter 12 of ref. 44. The MF59 emulsion advantageously includes citrate ions e.g. 10 mM sodium citrate buffer

An emulsion comprising squalene, a tocopherol, and 60 polysorbate 80. The emulsion may include phosphate buffered saline. These emulsions may have by volume from 2 to 10% squalene, from 2 to 10% tocopherol and from 0.3 to 3% polysorbate 80, and the weight ratio of squalene:tocopherol is preferably <1 (e.g. 0.90) as this 65 can provide a more stable emulsion. Squalene and polysorbate 80 may be present volume ratio of about 5:2

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or at a weight ratio of about 11:5. Thus the three components (squalene, tocopherol, polysorbate 80) may be present at a weight ratio of 1068:1186:485 or around 55:61:25. One such emulsion ('AS03') can be made by dissolving Tween 80 in PBS to give a 2% solution, then mixing 90 ml of this solution with a mixture of (5 g of DL a tocopherol and 5 ml squalene), then microfluidising the mixture. The resulting emulsion may have submicron oil droplets e.g. with an average diameter of between 100 and 250 nm, preferably about 180 nm. The emulsion may also include a 3-de-O-acylated monophosphoryl lipid A (3d MPL). Another useful emulsion of this type may comprise, per human dose, 0.5-10 mg squalene, 0.5-11 mg tocopherol, and 0.1-4 mg polysorbate 80 [45] e.g. in the ratios discussed above.

An emulsion of squalene, a tocopherol, and a Triton detergent (e.g. Triton X-100). The emulsion may also include a 3d-MPL (see below). The emulsion may contain a phosphate buffer.

An emulsion comprising a polysorbate (e.g. polysorbate 80), a Triton detergent (e.g. Triton X-100) and a tocopherol (e.g. an α-tocopherol succinate). The emulsion may include these three components at a mass ratio of about 75:11:10 (e.g. 750 μg/ml polysorbate 80, 110 μg/ml Triton X-100 and 100 μg/ml α-tocopherol succinate), and these concentrations should include any contribution of these components from antigens. The emulsion may also include squalene. The emulsion may also include a 3d-MPL (see below). The aqueous phase may contain a phosphate buffer.

An emulsion of squalane, polysorbate 80 and poloxamer 401 ("PluronicTM L121"). The emulsion can be formulated in phosphate buffered saline, pH 7.4. This emulsion is a useful delivery vehicle for muramyl dipeptides, and has been used with threonyl-MDP in the "SAF-1" adjuvant [46] (0.05-1% Thr-MDP, 5% squalane, 2.5% Pluronic L121 and 0.2% polysorbate 80). It can also be used without the Thr-MDP, as in the "AF" adjuvant [47] (5% squalane, 1.25% Pluronic L121 and 0.2% polysorbate 80). Microfluidisation is preferred.

An emulsion comprising squalene, an aqueous solvent, a polyoxyethylene alkyl ether hydrophilic nonionic surfactant (e.g. polyoxyethylene (12) cetostearyl ether) and a hydrophobic nonionic surfactant (e.g. a sorbitan ester or mannide ester, such as sorbitan monoleate or 'Span 80'). The emulsion is preferably thermoreversible and/ or has at least 90% of the oil droplets (by volume) with a size less than 200 nm [48]. The emulsion may also include one or more of: alditol; a cryoprotective agent (e.g. a sugar, such as dodecylmaltoside and/or sucrose); and/or an alkylpolyglycoside. The emulsion may include a TLR4 agonist [49]. Such emulsions may be lyophilized.

An emulsion of squalene, poloxamer 105 and Abil-Care [50]. The final concentration (weight) of these components in adjuvanted vaccines are 5% squalene, 4% poloxamer 105 (pluronic polyol) and 2% Abil-Care 85 (Bis-PEG/PPG-16/16 PEG/PPG-16/16 dimethicone; caprylic/capric triglyceride).

An emulsion having from 0.5-50% of an oil, 0.1-10% of a phospholipid, and 0.05-5% of a non-ionic surfactant. As described in reference 51, preferred phospholipid components are phosphatidylcholine, phosphatidylethanolamine, phosphatidylserine, phosphatidylinositol, phosphatidylglycerol, phosphatidic acid, sphingomyelin and cardiolipin. Submicron droplet sizes are advantageous.

A submicron oil-in-water emulsion of a non-metabolisable oil (such as light mineral oil) and at least one surfactant (such as lecithin, Tween 80 or Span 80). Additives may be included, such as QuilA saponin, cholesterol, a saponin-lipophile conjugate (such as GPI-0100, described in 5 reference 52, produced by addition of aliphatic amine to desacylsaponin via the carboxyl group of glucuronic acid), dimethyldioctadecylammonium bromide and/or N,N-dioctadecyl-N,N-bis(2-hydroxyethyl)propanediamine.

An emulsion in which a saponin (e.g. QuilA or QS21) and a sterol (e.g. a cholesterol) are associated as helical micelles [53].

An emulsion comprising a mineral oil, a non-ionic lipophilic ethoxylated fatty alcohol, and a non-ionic hydro- 15 philic surfactant (e.g. an ethoxylated fatty alcohol and/or polyoxyethylene-polyoxypropylene block copolymer) [54].

An emulsion comprising a mineral oil, a non-ionic hydrophilic ethoxylated fatty alcohol, and a non-ionic lipo- 20 philic surfactant (e.g. an ethoxylated fatty alcohol and/or polyoxyethylene-polyoxypropylene block copolymer)

In some embodiments an emulsion may be mixed with antigen extemporaneously, at the time of delivery, and thus 25 the adjuvant and antigen may be kept separately in a packaged or distributed vaccine, ready for final formulation at the time of use. In other embodiments an emulsion is mixed with antigen during manufacture, and thus the composition is packaged in a liquid adjuvanted form. The antigen will gen- 30 erally be in an aqueous form, such that the vaccine is finally prepared by mixing two liquids. The volume ratio of the two liquids for mixing can vary (e.g. between 5:1 and 1:5) but is generally about 1:1. Where concentrations of components are given in the above descriptions of specific emulsions, these 35 concentrations are typically for an undiluted composition, and the concentration after mixing with an antigen solution will thus decrease.

Packaging of Vaccine Compositions

kit components) include vials, syringes (e.g. disposable syringes), nasal sprays, etc. These containers should be sterile.

Where a composition/component is located in a vial, the vial is preferably made of a glass or plastic material. The vial 45 is preferably sterilized before the composition is added to it. To avoid problems with latex-sensitive patients, vials are preferably sealed with a latex-free stopper, and the absence of latex in all packaging material is preferred. The vial may include a single dose of vaccine, or it may include more than 50 one dose (a 'multidose' vial) e.g. 10 doses. Preferred vials are made of colourless glass.

A vial can have a cap (e.g. a Luer lock) adapted such that a pre-filled syringe can be inserted into the cap, the contents of the syringe can be expelled into the vial (e.g. to reconstitute 55 lyophilised material therein), and the contents of the vial can be removed back into the syringe. After removal of the syringe from the vial, a needle can then be attached and the composition can be administered to a patient. The cap is preferably located inside a seal or cover, such that the seal or 60 cover has to be removed before the cap can be accessed. A vial may have a cap that permits aseptic removal of its contents, particularly for multidose vials.

Where a component is packaged into a syringe, the syringe may have a needle attached to it. If a needle is not attached, a 65 separate needle may be supplied with the syringe for assembly and use. Such a needle may be sheathed. Safety needles

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are preferred. 1-inch 23-gauge, 1-inch 25-gauge and 5%-inch 25-gauge needles are typical. Syringes may be provided with peel-off labels on which the lot number, influenza season and expiration date of the contents may be printed, to facilitate record keeping. The plunger in the syringe preferably has a stopper to prevent the plunger from being accidentally removed during aspiration. The syringes may have a latex rubber cap and/or plunger. Disposable syringes contain a single dose of vaccine. The syringe will generally have a tip cap to seal the tip prior to attachment of a needle, and the tip cap is preferably made of a butyl rubber. If the syringe and needle are packaged separately then the needle is preferably fitted with a butyl rubber shield. Preferred syringes are those marketed under the trade name "Tip-Lok"TM.

Containers may be marked to show a half-dose volume e.g. to facilitate delivery to children. For instance, a syringe containing a 0.5 ml dose may have a mark showing a 0.25 ml volume.

Where a glass container (e.g. a syringe or a vial) is used, then it is preferred to use a container made from a borosilicate glass rather than from a soda lime glass.

A kit or composition may be packaged (e.g. in the same box) with a leaflet including details of the vaccine e.g. instructions for administration, details of the antigens within the vaccine, etc. The instructions may also contain warnings e.g. to keep a solution of adrenaline readily available in case of anaphylactic reaction following vaccination, etc.

Methods of Treatment, and Administration of the Vaccine

The invention provides a vaccine manufactured according to the invention. These vaccine compositions are suitable for administration to human or non-human animal subjects, such as pigs or birds, and the invention provides a method of raising an immune response in a subject, comprising the step of administering a composition of the invention to the subject. The invention also provides a composition of the invention for use as a medicament, and provides the use of a composition of the invention for the manufacture of a medicament for raising an immune response in a subject.

The immune response raised by these methods and uses Suitable containers for compositions of the invention (or 40 will generally include an antibody response, preferably a protective antibody response. Methods for assessing antibody responses, neutralising capability and protection after influenza virus vaccination are well known in the art. Human studies have shown that antibody titers against hemagglutinin of human influenza virus are correlated with protection (a serum sample hemagglutination-inhibition titer of about 30-40 gives around 50% protection from infection by a homologous virus) [55]. Antibody responses are typically measured by hemagglutination inhibition, by microneutralisation, by single radial immunodiffusion (SRID), and/or by single radial hemolysis (SRH). These assay techniques are well known in the art.

> Compositions of the invention can be administered in various ways. The most preferred immunisation route is by intramuscular injection (e.g. into the arm or leg), but other available routes include subcutaneous injection, intranasal [56-57], oral [58], intradermal [59,60], transcutaneous, transdermal [61], etc.

> Vaccines prepared according to the invention may be used to treat both children and adults. Influenza vaccines are currently recommended for use in pediatric and adult immunisation, from the age of 6 months. Thus a human subject may be less than 1 year old, 1-5 years old, 5-15 years old, 15-55 years old, or at least 55 years old. Preferred subjects for receiving the vaccines are the elderly (e.g. >50 years old, ≥60 years old, and preferably ≥65 years), the young (e.g. ≤5 years old), hospitalised subjects, healthcare workers, armed service

and military personnel, pregnant women, the chronically ill, immunodeficient subjects, subjects who have taken an antiviral compound (e.g. an oseltamivir or zanamivir compound; see below) in the 7 days prior to receiving the vaccine, people with egg allergies and people travelling abroad. The vaccines are not suitable solely for these groups, however, and may be used more generally in a population. For pandemic strains, administration to all age groups is preferred.

Preferred compositions of the invention satisfy 1, 2 or 3 of the CPMP criteria for efficacy. In adults (18-60 years), these 10 criteria are: (1) \geq 70% seroprotection; (2) \geq 40% seroconversion; and/or (3) a GMT increase of \geq 2.5-fold. In elderly (>60 years), these criteria are: (1) \geq 60% seroprotection; (2) \geq 30% seroconversion; and/or (3) a GMT increase of \geq 2-fold. These criteria are based on open label studies with at least 50 15 patients.

Treatment can be by a single dose schedule or a multiple dose schedule. Multiple doses may be used in a primary immunisation schedule and/or in a booster immunisation schedule. In a multiple dose schedule the various doses may 20 be given by the same or different routes e.g. a parenteral prime and mucosal boost, a mucosal prime and parenteral boost, etc. Administration of more than one dose (typically two doses) is particularly useful in immunologically naïve patients e.g. for people who have never received an influenza vaccine before, 25 or for vaccinating against a new HA subtype (as in a pandemic outbreak). Multiple doses will typically be administered at least 1 week apart (e.g. about 2 weeks, about 3 weeks, about 4 weeks, about 6 weeks, about 8 weeks, about 10 weeks, about 12 weeks, about 16 weeks, etc.).

Vaccines produced by the invention may be administered to patients at substantially the same time as (e.g. during the same medical consultation or visit to a healthcare professional or vaccination centre) other vaccines e.g. at substantially the same time as a measles vaccine, a mumps vaccine, a substantially vaccine, a MMR vaccine, a varicella vaccine, a MMRV vaccine, a diphtheria vaccine, a tetanus vaccine, a pertussis vaccine, a DTP vaccine, a conjugated *H. influenzae* type b vaccine, an inactivated poliovirus vaccine, a hepatitis B virus vaccine, a meningococcal conjugate vaccine (such as a 40 tetravalent A-C-W135-Y vaccine), a respiratory syncytial virus vaccine, a pneumococcal conjugate vaccine, etc. Administration at substantially the same time as a pneumococcal vaccine and/or a meningococcal vaccine is particularly useful in elderly patients.

Similarly, vaccines of the invention may be administered to patients at substantially the same time as (e.g. during the same medical consultation or visit to a healthcare professional) an antiviral compound, and in particular an antiviral compound active against influenza virus (e.g. oseltamivir and/or zanamivir). These antivirals include neuraminidase inhibitors, such as a (3R,4R,5S)-4-acetylamino-5-amino-3(1-ethylpropoxy)-1-cyclohexene-1-carboxylic acid or 5-(acetylamino)-4-[(aminoiminomethyl)-amino]-2,6-anhydro-3,4,5-trideoxy-D-glycero-D-galactonon-2-enonic acid, including seters thereof (e.g. the ethyl esters) and salts thereof (e.g. the phosphate salts). A preferred antiviral is (3R,4R,5S)-4-acetylamino-5-amino-3(1-ethylpropoxy)-1-cyclohexene-1-carboxylic acid, ethyl ester, phosphate (1:1), also known as oseltamivir phosphate (TAMIFLUTM).

The term "comprising" encompasses "including" as well as "consisting" e.g. a composition "comprising" X may consist exclusively of X or may include something additional e.g.

The word "substantially" does not exclude "completely" e.g. a composition which is "substantially free" from Y may

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be completely free from Y. Where necessary, the word "substantially" may be omitted from the definition of the invention

The term "about" in relation to a numerical value x is optional and means, for example, x±10%.

Unless specifically stated, a process comprising a step of mixing two or more components does not require any specific order of mixing. Thus components can be mixed in any order. Where there are three components then two components can be combined with each other, and then the combination may be combined with the third component, etc.

The various steps of the methods may be carried out at the same or different times, in the same or different geographical locations, e.g. countries, and by the same or different people or entities.

Where animal (and particularly bovine) materials are used in the culture of cells, they should be obtained from sources that are free from transmissible spongiform encephalopathies (TSEs), and in particular free from bovine spongiform encephalopathy (BSE). Overall, it is preferred to culture cells in the total absence of animal-derived materials.

Where a compound is administered to the body as part of a composition then that compound may alternatively be replaced by a suitable prodrug.

References to a percentage sequence identity between two amino acid sequences means that, when aligned, that percentage of amino acids are the same in comparing the two sequences. This alignment and the percent homology or sequence identity can be determined using software programs known in the art, for example those described in section 7.7.18 of reference 62. A preferred alignment is determined by the Smith-Waterman homology search algorithm using an affine gap search with a gap open penalty of 12 and a gap extension penalty of 2, BLOSUM matrix of 62. The Smith-Waterman homology search algorithm is taught in reference 63

References to a percentage sequence identity between two nucleic acid sequences mean that, when aligned, that percentage of bases are the same in comparing the two sequences. This alignment and the percent homology or sequence identity can be determined using software programs known in the art, for example those described in section 7.7.18 of reference 62. A preferred alignment program is GCG Gap (Genetics Computer Group, Wisconsin, Suite Version 10.1), preferably using default parameters, which are as follows: open gap=3; extend gap=1.

BRIEF DESCRIPTION OF THE DRAWINGS

FIG. 1 illustrates virus titers (by Focus-Formation assay (FFA); (A) and HA titers (by Red Blood Cell Hemagglutination assay; (B) at different times post-infection of wt PR8 and PR8-X viruses grown in MDCK cells. The solid line in (A) and hatched columns in (B) represent results with wild-type PR8. The dotted line in (A) and empty columns in (B) represent results with wild-type PR8-X. The x-axis shows the hours post infection and the y-axis in (A) and (B) shows the virus titer (IU/ml) and HA titre, respectively.

FIG. 2 illustrates virus titers (by FFA; (A) and HA titers (by Red Blood Cell Hemagglutination assay; (B) at different times post-infection of reverse genetics derived PR8 and PR8-X viruses grown in MDCK cells. The solid line in (A) and hatched columns in (B) represent results with PR8. The dotted line in (A) and empty columns in (B) represent results with RG-derived PR8-X. The x-axis shows the hours post infection and the y-axis in (A) and (B) shows the virus titer (IU/ml) and HA titre, respectively.

FIG. 3 compares virus titers (by FFA; (A) and HA titers (by Red Blood Cell Hemagglutination assay; (B)—at different times post-infection in MDCK cells of reverse genetics-derived 6:2 reassortant viruses made with either PR8 or PR8-X backbone segments which contain the HA and NA segments from PR8-X. The solid line in (A) and hatched columns in (B) represent results with the PR8 backbone. The dotted line in (A) and empty columns in (B) represent results with the PR8-X backbone. The x-axis shows the hours post infection and the y-axis in (A) and (B) shows the virus titer (IU/ml) and

FIG. 4 compares virus titers by FFA (A) and HA titers (by Red Blood Cell Hemagglutination assay; (B) at different times post-infection in MDCK cells of reverse genetics-derived 6:2 reassortant viruses made with either wt PR8 or PR8-X backbone segments which contain the HA and NA segments from a pandemic H1 strain (strain 1). The solid line in (A) and hatched columns in (B) represent results with the wt PR8 backbone. The dotted line in (A) and empty columns in (B) represent results with the PR8-X backbone. The x-axis shows the hours post infection and the y-axis in (A) and (B) shows the virus titer (IU/ml) and HA titre, respectively.

HA titre, respectively.

FIG. 5 compares virus titers by a focus-formation assay (FFA) (A) and HA titers (B) at different times post-infection 25 in MDCK cells of reverse genetics-derived 6:2 reassortant viruses made with either PR8 or PR8-X backbone segments which contain the HA and NA segments from 105p30. The solid line in (A) and hatched columns in (B) represent results with the wt PR8 backbone. The dotted line in (A) and empty 30 columns in (B) represent results with the PR8-X backbone. The x-axis shows the hours post infection and the y-axis shows the virus titer (IU/ml).

FIG. 6 illustrates virus titers by a focus-formation assay (FFA) at different times post-infection of wild-type PR8-X 35 and 105p30 viruses (A) or reverse genetics-derived PR8-X and 105p30 viruses (B) grown in MDCK cells. In (A) and (B), the solid lines represent results with 105p30. The dotted lines represent results with PR8-X. The x-axis shows the hours post infection and the y-axis in (A) and (B) shows the virus titer 40 (IU/ml) and HA titre, respectively.

FIG. 7 shows the growth characteristics of reassortant viruses containing the backbone segments of the wt PR8 strain (line with triangles) or 105p30 strain (line with squares) and the HA and NA segments of a pandemic H1 influenza 45 strain (strain 2). The x-axis in (A) and (B) indicates the hours post infection. The y-axis in (A) shows the titre Log 10 in FFU per mL. The y-axis in (B) shows the titre log 10 in virus particles per mL.

FIG. **8** compares virus titers by a focus-formation assay 50 (FFA) at different times post-infection in MDCK cells of reverse genetics-derived 6:2 reassortant viruses made with either 105p30 or PR8-X backbone segments which contain the HA and NA segments from (A) a H1 strain (strain 1) or (B) a pandemic H1 strain (strain 2). The solid lines represent results with the 105p30 backbone. The dotted lines represent results with the PR8-X backbone. The x-axis shows the hours post infection and the y-axis shows the virus titer (IU/ml).

FIG. 9 compares virus titers by a focus-formation assay (FFA) at different times post-infection in MDCK cells of 60 reverse genetics-derived 6:2 reassortant viruses made with either the #17, #19, or PR8-X backbone in combination with the HA and NA segments from (A) a pandemic H1 strain (strain 3) or (B) a H3 (strain 1). In (A) and (B), the dotted lines with the circle markers represent results with the #17 backbone. The solid lines with diamond markers represent results with the #19 backbone. The dotted lines with square markers

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represent results with the PR8-X backbone. The x-axis shows the hours post infection and the y-axis shows the virus titer (IU/ml).

FIG. 10 compares virus titers by a focus-formation assay (FFA) at different times post-infection in MDCK cells of a panel of different reverse genetics-derived 6:2 reassortant viruses made with either the chimeric #19 or PR8-X backbone plus the HA and NA segments from the following strains: (A) a pandemic H1 strain (strain 2), (B) a pandemic H1 strain (strain 4)-In (A) and (B), the solid lines with the triangle markers represent results with the #19 backbone. The dotted lines with square markers represent results with the PR8-X backbone. The x-axis shows the hours post infection and the y-axis shows the virus titer (IU/ml).

FIG. 11 compares virus titers by a focus-formation assay (FFA) at different times post-infection in MDCK cells of a panel of different reverse genetics-derived 6:2 reassortant viruses made with either the chimeric #19 or PR8-X backbone plus the HA and NA segments from the following strains: (C) a H1 strain (strain 2), (D) a H1 strain (strain 3). In (C) and (D), the solid lines with the triangle markers represent results with the #19 backbone. The dotted lines with square markers represent results with the PR8-X backbone. The x-axis shows the hours post infection and the y-axis shows the virus titer (IU/ml).

FIG. 12 compares virus titers by a focus-formation assay (FFA) at different times post-infection in MDCK cells of a panel of different reverse genetics-derived 6:2 reassortant viruses made with either the chimeric #19 or PR8-X backbone plus the HA and NA segments from the following strain: a H3 strain (strain 2). In FIG. 12, the solid lines with the triangle markers represent results with the #19 backbone. The dotted lines with square markers represent results with the PR8-X backbone. The x-axis shows the hours post infection and the y-axis shows the virus titer (IU/ml).

FIG. 13 compares HA yields (by lectin-capture ELISA) at 60 hr post-infection in MDCK cells of different 6:2 reassortant viruses made with either the chimeric #19 (empty columns) or PR8-X backbone (solid columns) plus the HA and NA segments from the following strains: (A) a pandemic H1 strain (strain 2), (B) a pandemic H1 strain (strain 4). Corresponding 6:2 reassortant viruses made by classical reassortment ("classical") with the wt PR8 backbone were included as controls (hatched columns). The y-axis shows the HA content in µg per mL.

FIG. 14 compares HA yields (by lectin-capture ELISA) at 60 hr post-infection in MDCK cells of different 6:2 reassortant viruses made with either the chimeric #19 (empty columns) or PR8-X backbone (solid columns) plus the HA and NA segments from the following strains: (C) a H3 strain (strain 1), or (D) a H3 strain (strain 2). Corresponding 6:2 reassortant viruses made by classical reassortment ("classical") with the wt PR8 backbone were included as controls (hatched columns) The y-axis shows the HA content in µg per mL.

FIG. 15 shows the growth curves of reassortant influenza viruses. (A) shows growth curves of reassortant influenza viruses comprising backbones 17, 18, 19 and 20 (as shown in table 1; line with diamonds, squares, triangles and crosses, respectively), a control comprising the same HA and NA segments from a H3 influenza strain (strain 1) but all backbone segments from PR8-X (line with circles) and the equivalent wildtype strain (line with plus sign). The x axis indicates the hours post infection (hpi) and the y-axis shows IU/mL. (B) shows the growth curve of reassortant influenza viruses comprising backbones 17 and 19 (line with diamonds and triangles, respectively) and the HA segments from a H3 influ-

enza strain (strain 3), a control comprising the same HA and NA segments but all backbone segments from PR8-X (line with plus sign) and the equivalent wildtype strain (line with circles).

FIG. **16** shows the results of a FFA (A) and HA-ELISA (B) assay using reassortant influenza viruses comprising backbone 19 (open box), PR8-X backbone (hatched box) and the wildtype influenza virus (dotted box). (A) and (B) show the results with a H1 influenza strain (strain 2). The y axis in (A) indicates the virus titre in IU/mL and the y axis in (B) indicates HA in μ g/mL.

FIG. 17 shows the results of a FFA (C) and HA-ELISA (D) assay using reassortant influenza viruses comprising backbone 19 (open box), PR8-X backbone (hatched box) and the wildtype influenza virus (dotted box). (C) and (D) show the results with a H3 influenza virus strain. The y axis in (C) indicates the virus titre in IU/mL and the y axis in (D) indicates HA in µg/mL.

FIG. 18 is an alignment of the M1 viral segment of A/New $_{20}$ Caledonia/20/99 (SEQ ID NO: 33) and 105p30 (SEQ ID NO: 45).

MODES FOR CARRYING OUT THE INVENTION

Development of New Donor Strains

In order to provide high-growth donor strains, the donor strain A/Puerto Rico/8/34 is passaged in MDCK 33016 cells five times. Using this method, the inventors were able to obtain the strain PR8-X which shows improved growth characteristics compared with the original strain.

The 105p30 influenza donor strain was provided by isolating an A/New Caledonia/20/1999 influenza virus from a clinical isolate in MDCK 33016 cells and passaging the virus 30 times. The resulting strain has a M segment with lysine in the 35 position corresponding to amino acid 95 of SEQ ID NO: 33 when aligned to SEQ ID NO: 33.

Growth Characteristics of Wt PR8 and PR8-X Viruses

In order to compare the growth characteristics of PR8-X and wt PR8 donor strains, the viral titre of these virus strains 40 is measured in MDCK cells by focus-forming assays and hemagglutination assays.

Focus-Forming Assays (FFA)

For the FFA, uninfected MDCK cells are plated at a density of 1.8×10^4 cells/well in 96 well plates in 100 μ l of DMEM 45 with 10% FCS. The next day, medium is aspirated and cells are infected with viruses in a volume of 50 μ l (viruses diluted in DMEM+1% FCS). The cells are incubated at 37° C. until the next day.

At several time points after infection, the medium is aspi- 50 rated and the cells washed once with PBS. 50 µl of ice-cold 50%/50% acetone-methanol is added to each well followed by incubation at -20° C. for 30 minutes. The acetone mix is aspirated and the cells washed once with PBST (PBS+0.1% Tween). 50 µl of 2% BSA in PBS is added to each well 55 followed by incubation at room temperature (RT) for 30 minutes. 50 µl of a 1:6000 dilution of anti-NP is added in blocking buffer followed by incubation at RT for 1 hours. The antibody solution is aspirated and the cells washed three times with PBST. Secondary antibody (goat anti mouse) is added at 60 a dilution 1:2000 in 50 µl blocking buffer and the plate is incubated at RT for 1 hours. The antibody solution is aspirated and the cells washed three times with PBST. 50 µl of KPL True Blue is added to each well and incubated for 10 minutes. The reaction is stopped by aspirating the True-Blue and wash- 65 ing once with dH₂O. The water is aspirated and the cells are left to dry.

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The results (FIG. 1) show that the PR8-X strain can grow to higher titres in the same time frame compared to the wt PR8 strain from which it is derived.

Growth Characteristics of Reassortant Viruses Containing PR8-X or Wt PR8 Backbones

In order to test the suitability of the PR8-X strain as a donor strain for virus reassortment, reassortant viruses are produced by reverse genetics which contain the HA and NA proteins from a pandemic H1 strain and the other viral segments from either PR8-X or PR8. The viral titres of these reassortant viruses are determined by FFA and HA assays as described above. The results are shown in FIG. 4.

The results indicate that reassortant viruses which contain viral segments from PR8-X grow faster in MDCK cells compared to reassortant viruses containing viral segments from the PR8/34 strain.

Growth Characteristics of 105p30 Strain Compared with PR8-X

MDCK cells are infected with 105p30 and PR8-X at a moi of 10⁻³ and samples are taken at several time points after infection. The titre is determined by a FFA assay. The results show that 105p30 grows even faster in MDCK cells compared to PR8-X (FIG. 6).

Growth Characteristics of Reassortant Viruses Containing 105p30 or wt PR8 Backbones

In order to test the suitability of the 105p30 strain as a donor strain for virus reassortment, reverse genetics is used to produce reassortant viruses that contain the HA and NA segments from a pandemic H1 influenza strain and the backbone segments either from the 105p30 or the wt PR8 strain. MDCK cells are infected with the reassortant viruses at a moi of 10^{-3} and samples are taken 1 hour, 12 hours, 36 hours and 60 hours after infection. The titres are determined either by focusforming assays or by determining the virus particles by real-time detection PCR. The reassortant viruses that contain the backbone segments from the 105p30 strain grow faster than the viruses that are reassorted with the backbone segments of the wt PR8 strain. This shows that the 105p30 strain is a good donor strain for producing fast-growing reassortant viruses (FIG. 7).

Rescue of Influenza Viruses Using Backbone Segments from Two Donor Strains

The rescue efficiency of reassortant influenza viruses containing the HA and NA segments from a H3 influenza virus and backbone segments from the 105p30 and the PR8-X donor strains is tested in MDCK cells. The reassortant influenza viruses contain backbone segments of the 105p30 and the PR8-X donor strains, as indicated in the following table:

TABLE 1

_							
	Backbone #	FB1	PB2	PA	NP	M	NS
_	1	PR8-X	PR8-X	PR8-X	105p30	105p30	105p30
	2	PR8-X	PR8-X	105p30	PR8-X	105p30	105p30
	3	PR8-X	PR8-X	105p30	105p30	PR8-X	105p30
	4	PR8-X	PR8-X	105p30	105p30	105p30	PR8-X
	5	PR8-X	105p30	PR8-X	PR8-X	105p30	105p30
	6	PR8-X	105p30	PR8-X	105p30	PR8-X	105p30
	7	PR8-X	105p30	PR8-X	105p30	105p30	PR8-X
	8	PR8-X	105p30	105p30	PR8-X	PR8-X	105p30
	9	PR8-X	105p30	105p30	PR8-X	105p30	PR8-X
'	10	PR8-X	105p30	105p30	105p30	PR8-X	PR8-X
	11	105p30	PR8-X	PR8-X	PR8-X	105p30	105p30
	12	105p30	PR8-X	PR8-X	105p30	PR8-X	105p30
	13	105p30	PR8-X	PR8-X	105p30	105p30	PR8-X
	14	105p30	PR8-X	105p30	PR8-X	105p30	PR8-X
	15	105p30	PR8-X	105p30	PR8-X	PR8-X	105p30
	16	105p30	PR8-X	105p30	105p30	PR8-X	PR8-X
	17	105p30	105p30	PR8-X	PR8-X	PR8-X	105p30

TABLE 1-continued

Backbone #	FB1	PB2	PA	NP	M	NS
18 19 20	105p30	105p30	PR8-X	105p30	105p30 PR8-X PR8-X	PR8-X

Reassortant influenza viruses which contain a backbone according to number 3, 4, 10, 11, 14 and 16-20 are rescuable. Influenza viruses which contain backbones number 3, 4, 10, 11 or 16 achieve viral titres of less than 10² IU/mL. Influenza viruses containing backbone numbers 17 and 18 achieve viral titres between 102 and 106 IU/mL and influenza viruses having backbone numbers 19 and 20 even achieve titres of more than 106 IU/mL.

These data show that influenza viruses in which the PB 1 and PB2 segments come from the same influenza donor strain can show a higher rescue efficiency compared with influenza viruses in which these segments come from different influenza donor strains.

Growth Characteristics of Reassortant Influenza Viruses Containing Backbone Segments from Two Donor Strains

Reassortant influenza strains are created which contain backbone numbers 17, 18, 19 and 20 (as shown in table 1 above) and the HA and NA segments from a H3 influenza $\,^{25}$ strain (strain 1). As controls, the equivalent wildtype H3 influenza virus, and a reassortant influenza virus comprising the same HA and NA segments and all backbone segments from PR8-X are used.

Furthermore, reassortant influenza strains are produced 30 which contain backbone numbers 17 and 19 and the HA and NA segments from either a second H3 influenza (strain 1) virus or a pandemic H1 influenza virus (strain 3). As controls for the H3 strain, the equivalent wildtype H3 (strain 2) influenza virus, and a reassortant influenza virus comprising the 35 same HA and NA segments and all backbone segments from PR8-X is used. For the pandemic H1 influenza virus a reassortant influenza virus comprising the same HA and NA segments and all backbone segments from PR8-X is used.

The reassortant influenza viruses and the control viruses 40 are grown in MDCK cells and the viral titre is measured by FFA at different time points. For the reassortant H3 viruses (strain 1) containing backbones 17, 19 and 20, and the H3 influenza viruses (strain 3) containing backbones 17 and 19, the influenza viruses containing backbone segments from two 45 donor strains grow to higher titres compared with the wild-

type virus and the reassortant virus which contains backbone segments from only a single donor strain (see FIG. 13, FIG. 14 and FIG. 15(A)).

For the pandemic H1 influenza virus, the reassortant influenza strains containing backbones 17 and 19 grow to higher titres compared with the control which contained all backbone segments from PR8-X (see FIG. 9).

The data show that reassortant influenza viruses which contain backbone segments from two different donor strains can show improved growth rates compared with reassortant influenza viruses which contain backbone segments from only a single donor strain.

The experiments were also repeated using reassortant influenza viruses which contain backbone 19 or the backbone segments from PR8-X in combination with the HA and NA segments from four different H1 strains or a H3 strain. The results are shown in FIG. 10, FIG. 11, and FIG. 12.

Reassortant Influenza Viruses with Backbone Segments from Two Different Donor Strains Give Higher Yields

To test whether reassortant influenza viruses containing backbone segments from two different influenza donor strains can also provide higher yields, the HA yield of the reassortant strains is tested by HA-ELISA. To this end, the same reassortant influenza viruses as described above containing backbone #19 and the HA/NA segments of the H3 (strain 2) and H1 influenza strains are used. As controls, the equivalent wildtype influenza viruses and reassortant influenza viruses comprising the same HA and NA segments and all backbone segments from PR8-X are used. In addition, the viral titres are confirmed with a FFA assay.

The results confirm that the reassortant influenza strains which contain backbone segments from two different donor strains can grow to higher yields compared with influenza viruses which contained all backbones from PR8-X (see FIG. 16 (A) and FIG. 17 (C)). Furthermore, reassortant influenza viruses comprising backbone segments from two donor strains also give higher HA yields (see FIG. 16 (B) and FIG. 17 (D)).

These data show that reassortant influenza viruses which contain backbone segments from two donor strains give higher yields compared with reassortant influenza viruses which contain backbone segments from only a single donor strains.

It will be understood that the invention has been described by way of example only and modifications may be made whilst remaining within the scope and spirit of the invention.

SEQUENCES

DEPOSIT INFORMATION

A deposit of the microorganism MDCK 33016 (DSM ACC2219) was deposited on Jun. 7, 1995 according to the Budapest Treaty

in the International Depository
Authority DSM-Deutsche Sammlung Von Mikroorganismem und Zellkulturen GmbH, Mascheroder Weg 1b, D-38124 Braunschweig.

SEQUENCE: 1 (PA, A/New Caledonia/20/1999)

GATTCGAAATGGAAGATTTTGTGCGACAATGCTTCAATCCGATGATTGTCGAGCTTGCGGAAAAGGCAATGAAAG

ATTCAGATTTCATCATCAATGAGCAAGGCGAATCAATAATAGTAGAGCCTGAGGACCCAAATGCACTTTTAA

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-continued

SEQUENCES

 $\verb|TCTCATTCACTGGCGAAGAAATGGCCACAAAGGCCGATTACACTCTCGATGAAGAAAGCAGGGCTAGGATTAAAA|$ $\tt CCAGACTATTCACCATAAGACAAGAAATGGCAAGCAGAGGTCTTTGGGACTCCTTTCGTCAGTCCGAAAGAGGGCG$ AAGAAACAATTGAAGAAAGATTTGAAATCACAGGGACAATGCGCAGGCTCGCTGACCAAAGCCTTCCGCCGAACT ${\tt TCTCCTGCATTGAGAATTTTAGAGCCTATGTGGATGGATTTGAACCGAACGGCTACATTGAGGGCAAGCTTTCTC}$ AAATGTCCAAAGAAGTAAATGCTAGAATTGAGCCTTTTTTGAAAACAACACCACGACCAATTAGACTTCCGGATG GGCCTCCTTGTTTTCAGCGGTCAAAATTCCTGCTGATGGATTCTTTAAAATTAAGCATTGAGGATCCAAATCATG TCAAGCCACACGGGAAGGGAATAAATCCGAATTATCTGCTGTCATGGAAGCAGGTATTGGAAGAGCTGCAGGACA TTGAGAGTGAGGAGAAGATTCCAAGAACAAAAAACATGAAAAAACGAGTCAGCTAAAGTGGGCACTTGGTGAGA $\tt CTGAATTAAGGTCATTTTCAAGTTGGATCCAGAATGAGTTCAACAAGGCATGCGAGCTGACCGATTCAATCTGGA$ TAGAGCTCGATGAGAATTGGAGAAGATGTGGCCCCCGATTGAACACATTGCAAGCATGAGAAGAAATTACTTCACAG CTGAGGTGTCCCATTGCAGAGCCACAGAATATATAATGAAGGGGGTATACATTAATACTGCTTTGCTTAATGCAT CCTGTGCAGCAATGGATGATTTCCAACTAATTCCCATGATAAGCAAATGTAGAACTAAAGAGGGAAGGAGAAAGA $\tt TGGAGTTTTCCCTCACTGACCCAAGACTTGAGCCACAAATGGGAGAAGTACTGTTCTTGAGATAGGAGATACTGAGATAGAGATAGGAGATACTGAGATAGATAGAGATAGAGATAGATAGAGATAGAGATAGAGATAGATAGAGATAGATAGATAGAGATAGAGATAG$ CTGAGTCCTCCGTCAAGGAGAAAGACATGACAAAAGAGTTTTTTGAGAATAGATCAGAAACATGGCCCATTGGAG AGTCACCAAAAGGAGTGGAAGAAGGTTCCATTGGGAAAGTATGCAGGACACTATTGGCTAAGTCAGTATTCAATA GTCTGTATGCATCTCCACAATTAGAAGGATTTTCAGCTGAGTCAAGAAAGTTGCTCCTCATTGTTCAGGCTCTTA GGGACAATCTGGAACCTGGGACCTTTGATCTTGGGGGGGCTATATGAAGCAATTGAGGAGTGCCTGATTAATGATC ATTTACTATCCATACTGTCCAAAAAA

SEQUENCE: 2 (PB1, A/New Caledonia/20/1999)

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SEQUENCES

ACTAGGTAAGGGGTACATGTTTGAAAGCAAGAGTATGAAACTGAGAACTCAAATACCTGCAGAGATGCTAGCCAA CATAGATTTGAAATATTTCAATGATTCAACTAAAAAGAAAATTGAAAAAATCCGGCCATTATTAATAGATGGAAC $\tt TGTGAATGCACCCAACTATGCAGGAATTCAAGCTGGAGTTGACAGGTTTTATCGAACCTGTAAGCTGCTCGGAATTGTGAATGCACCAACTATGCAGGAATTCAAGCTGGAGTTGACAGGTTTTATCGAACCTGTAAGCTGCTCGGAATTCAAGCTGCAGGAATTCAAGCTGCAGGAATTGAACAGGTTTTATCGAACCTGTAAGCTGCTCGGAATTCAAGCTGCAGGATTTGACAGGTTTTATCGAACCTGTAAGCTGCTCGGAATTCAAGCTGCAGGTTTGACAGGTTTTATCGAACCTGTAAGCTGCTCGGAATTCAAGCTGCAGAATTCAAGCTGCAGAATTCAAGCTGCAGAATTCAAGCTGCAGGTTTTATCGAACCTGTAAGCTGCTCGGAATTCAAGCTGCAGAATTCAAGCTGCAGAATTCAAGCTGCAGAACCTGTAAGCTGCTCGGAATTCAAGCTGCAGAATTCAAGAAATTCAAGAATTCAAATTCAAATTCAAATTCAAGAATTCA$ TAATATGAGCAAAAAGAAGTCTTACATAAACAGAACAGGTACCTTTGAGTTCACGAGCTTTTTCTATCGTTATGG GTTTGTTGCCAATTTCAGCATGGAGCTTCCTAGTTTTGGGGTGTCTTGGGGTCAATGAATCTGCAGACATGAGTAT TGGAGTCACTGTCATCAAAAACAATATGATAAACAATGACCTTGGCCCAGCAACTGCTCAAATGGCCCTTCAGTT ATTTATAAAAGATTACAGGTACACGTATCGATGCCACAGAGGTGACACAAATACAAACCCGGAGATCATTTGA ${\tt GATAAAGAAACTATGGGACCAAACCCGCTCCAAAGCTGGGCTGTTGGTCTCTGATGGAGGCCCCAATTTATATAA}$ CAACCCATTGAACCCGTTTGTCAGTCATAAAGAGATTGAATCAGTGAACAATGCAGTGATGATGCCGGCACATGG ${\tt TCCAGCCAAAAATATGGAGTATGACGCTGTTGCAACAACACCTCCTGGGTTCCCAAAAGGAATCGATCCATTTT}$ TGCACGGATTGATTTCGAATCTGGAAGGATAAAAAAAGGGGAATTCGCTGAGATCATGAAGACCTGTTCCACCAT ${\tt TGAAGACCTCAGACGGCAAAAATAGGGAATTTGGCTTGTCCTTCATGAAAA}$

SEOUENCE: 3 (PB2, A/New Caledonia/20/1999) GTAGACCACATGGCCATAATCAAGAAATACACATCAGGAAGACAGGAGAAAAACCCATCACTTAGAATGAAATGG TTAAAACATGGAACCTTTGGCCCTGTACACTTTAGAAACCAAGTCAAAATACGCCGAAGAGTCGACATAAATCCT ${\tt GGTCATGCAGACCTCAGCGCCAAGGAGGCACAGGATGTAATTATGGAAGTTGTTTTCCCTAATGAAGTGGGAGCC}$ ${\tt AGAATACTAACATCAGAATCGCAATTAACGATAACCAAGGAGAAAAAAGAAGAACTCCAGAATTGCAAAATTTCC}$ $\tt CCTTTGATGGTTGCATACATGTTAGAGAGGGAACTTGTCCGCAAAACGAGATTTCTCCCGGTTGCTGGTAGAACA$ AGCAGTGTGTACATTGAAGTTTTGCATTTAACACAGGGGACATGCTGGGAGCAGATGTACACTCCAGGTGGGGAG GTGAGGAATGATGATGATCAAAGCCTAATTATTGCTGCTAGGAACATAGTGAGAAGAGCTGCAGTATCAGCA GATCCACTAGCATCTTTATTAGAAATGTGCCATAGCACACAGATTGGTGGACAAGGATGGTGGATATTCTCAGG GTAGCAATGGTATTCTCACAAGAAGATTGCATGGTAAAAGCAGTTAGAGGTGATCTGAATTTCGTTAATAGAGCG AATCAGCGGTTGAATCCCATGCATCAACTTTTGAGACATTTTCAGAAGGATGCTAAAGTACTTTTCTTAAATTGG GGAATTGAACCTATCGACAATGTGATGGGAATGATTGGGATATTACCTGATATGACTCCAAGTACCGAGATGTCA ATGAGAGGAGTGAGAGTCAGCAAAATGGGTGTAGATGAATACTCCAATGCTGAAAGGGTAGTGGTGAGCATTGAC CGTTTTTTGAGAGTCCGGGACCAAAGAGGAAATGTACTACTGTCTCCAGAGGAAGTCAGTGAAACACAGGGAACA

SEQUENCES

SEQUENCE: 4 (NP, A/New Caledonia/20/1999) GGGAACGCCAGAATGCAACTGAAATCAGAGCATCCGTCGGAAGAATGATTGGTGGAATTGGGCGATTCTACATCC AAATGTGCACCGAGCTTAAACTCAATGATTATGAGGGACGACTGATCCAGAACAGCTTGACAATAGAGAGAATGG $\tt CTGGAGGACCCATATACAAGAGAGTAGATGGAAAGTGGGTGAGGGAACTCGTCCTTTATGACAAAGAAGAAATAA$ GGCGGATTTGGCGCCAAGCCAACAATGGTGATGATGCAACGGCTGGTTTGACTCACATTATGATCTGGCATTCTA ATTTGAATGATACAACTTACCAGAGGACAAGAGCTCTTGTCCGCACCGGAATGGATCCCAGGATGTGCTCTTTGA AGTTAATCAGGATGATCAAACGTGGGATCAATGACCGAAACTTCTGGAGGGGTGAGAATGGAAGAAAAACAAGGA TGAGAGAAAGCCGGAACCCAGGAAATGCTGAGATCGAAGATCTCACTTTTCTGGCACGGTCTGCACTCATATTAA GAGGGTCAGTTGCTCACAAGTCTTGCCTGCCTGTCTGTGTATGGACCAGCCGTAGCCAGTGGGTACGACTTCG AAAAAGAGGGATACTCTTTGGTAGGGGTAGACCCTTTTAAACTGCTTCAAACCAGTCAGGTATACAGCCTAATCAGACCAAACGAGAATCCCGCACACAAGAGTCAGTTGGTGTGGATGGCATGCAATTCTGCTGCATTTGAAGATCTAA GAGTGTCAAGCTTCATCAGAGGGACAAGAGTACTTCCAAGGGGGAAGCTCTCCACTAGAGGAGTACAAATTGCTT CAAATGAAAACATGGATGCTATTGTATCAAGTACTCTTGAACTGAGAAGCAGATACTGGGCCATAAGAACCAGAA GTGGAGGGAACACTAATCAACAAAGGGCCTCTGCGGGCCAAATCAGCACACACCTACGTTTTCTGTGCAGAGAA CAGAAATCATAAAGATGATGGAAAGTGCAAGACCAGAAGAAGTGTCCTTCCAGGGGCGGGAGTCTTTGAGCTCT ATGCAGAGGAGTACGACAATTAATGAA

SEQUENCE: 5 (M, A/New Caledonia/20/1999)
GATGAGTCTTCTAACCGAGGTCGAAACGTACGTTCTCTCTATCGTCCCGTCAGGCCCCCTCAAAGCCGAGATCGC
ACAGAGACTTGAAAAATGTCTTTGCTGGAAAGAATACCGATCTTGAGGCTCCATGGAATGGCTAAAGACAAGACC
AATCCTGTCACCTCTGACTAAGGGGATTTTAGGATTTGTGTTCACGCTCACCGTGCCCAGTGAGCGAGGACTGCA
GCGTAGACGCTTTGTCCAAAATGCCCTTAATGGGAATGGGGATCCAAATAATATGGACAGAGCAGTTAAACTGTA
TCGAAAGCTTAAGAGGGAGATAACATTCCATGGGGCCAAAGAAATAACACCAGCTTATTCTGCTGGTGCACTTGC
CAGTTGTATGGGACTCATATACAACAGGATGGGGGCTGTGACCACCGAATCAGCATTTGGCCTTATATGCGCAAC

SEQUENCES

SEQUENCE: 6 (NS, A/New Caledonia/20/1999)
ATGGATTCCCACACTGTGCAAGCTTTCAGGTAGATTGCTTCCTTTGGCATGTCCGCAAACAAGTTGCAGACCAA
GATCTAGGCGATGCCCCATTCCTTGATCGGCTTCGCCGAGATCAGAAGTTCCTAAAGGGAAGAGCAGCACTCTC
GGTCTGAACATCGAAACAGCCACTTGTGTTGGAAAGCAAATAGTAGAGAGGATTCTGAAAGAAGAAATCCGATGAG
GCATTTAAAATGACCATGGCCTCCGCACTTGCTTCGCGGTACCTAACTGACATGACTATTGAAGAAAATGTCAAGG
GACTGGTTCATGCTCATGCCCAAGCAGAAAGTGGCTGGCCCTCTTTGTGTCAGAATGGACCAGGCGATAATGGAT
AAGAACATCATACTGAAAGCGAAATTTCAGTGTGATTTTTGACCGGTTGGAGAATCTGACATTACTAAGGGCTTTC
ACCGAAGAGGGGAGCAATTGTTGGCGAAATTTCACCATTGCCTTCTCTCCAGGACATACTAATGAGGATGTCAAA
AATGCAATTGGGGTCCTCATCGGGGGACTTGAATGGAATGATAACACAGTTCGAGTCTCTGAAACTCTACAGAGA
TTCGCTTGGAGAAGCAGTAATGAGACTGGGGGACCTCCATTCACTCCAACACAGAAACCGGAAAATGGCGGGAACA
ATTAGGTCAGAGAGTTTGAAGAAATAAGATGGCTGATTGAAGAAGTGAGGCATAAATTGAAGACGACAGAAATAG
TTTTGAGCCAAATAACATTTATGCAAGCATTACAGCTATTGTTTGAAGTGGAACAAGAGATTAGAACGTTTTCGTT

SEQUENCE: 7 (HA, A/New Caledonia/20/1999)

TCAGCTTATTTAATGATAA

ACCATGCCAACAACTCAACCGACACTGTTGACACAGTACTTGAGAAGAATGTGACAGTGACACACTCTGTCAACC CACCAAATCCTGAGAATGGAACATGTTACCCAGGGTATTTCGCCGACTATGAGGAACTGAGGGAGCAATTGAGTT ${\tt CAGCATCATGCTCCCATAATGGGAAAAGCAGTTTTTACAGAAATTTGCTATGGCTGACGGGGAAGAATGGTTTGT}$ $\tt CTAACATAGGGAACCAAAGGGCCCTCTATCATACAGAAAATGCTTATGTCTCTGTAGTGTCTTCACATTATAGCA$ TGCTGGAACCTGGGGATACAATAATATTTGAGGCAAATGGAAATCTAATAGCGCCATGGTATGCTTTTGCACTGA GTAGAGGCTTTGGATCAGGAATCATCACCTCAAATGCACCAATGGATGAATGTGATGCGAAGTGTCAAACACCTC AGGGAGCTATAAACAGCAGTCTTCCTTTCCAGAATGTACACCCAGTCACAATAGGAGAGTGTCCAAAGTATGTCA TTGAGAAAATGAACACTCAATTCACAGCTGTGGGCAAAGAATTCAACAAATTGGAAAGAAGGATGGAAAACTTAA ATAAAAAAGTTGATGATGGTTTCTAGACATTTGGACATATAATGCAGAATTGTTGGTTCTACTGGAAAATGAAA

SEQUENCES

SEQUENCE: 8 (NA, A/New Caledonia/20/1999) ${\tt AATAGGAAATATTATTTCAATATGGGCTAGTCACTCAATCCAAACTGGAAGTCAAAACCACACTGGAGTATGCAA}$ CCAAAGAATCATCACATATGAAAACAGCACCTGGGTGAATCACACATATGTTAATATTAACAACACTAATGTTGT ${\tt TCACTTGGAATGCAGAACCTTTTTTCTGACCCAAGGTGCTCTATTAAATGACAAACATTCAAATGGGACCGTTAA}$ ATCAGTTGCATGGTCAGCAAGCGCATGCCATGATGGCATGGGCTGGTTAACAATCGGAATTTCTGGTCCAGACAA TGGAGCTGTGGCTGTACTAAAATACAACGGCATAATAACTGAAACCATAAAAAGTTGGAAAAAGCGAATATTAAG AACACAGAGTCTGAATGTGTCTGTGTGAACGGGTCATGTTTCACCATAATGACCGATGGCCCGAGTAATGGGGC CGCCTCGTACAAAATCTTCAAGATCGAAAAGGGGAAGGTTACTAAATCAATAGAGTTGAATGCACCCAATTTTCA ATACAAATATGGTAATGGTGTTTGGATAGGAAGGACTAAAAGTAACAGACTTAGAAAGGGGTTTGAGATGATTTG GGATCCTAATGGATGGACAGATACCGACAGTGATTTCTCAGTGAAACAGGATGTTGTGGCAATAACTGATTGGTC GTTAGTCAGAGGACTGCCTAGAGAAAATACAACAATCTGGACTAGTGGGAGCAGCATTTCTTTTTTGTGGCGTAAA ${\tt TAGTGATACTGCAAACTGGTCTTGGCCAGACGGTGCTGAGTTGCCGTTCACCATTGACAAGTAG}$

SEQUENCES

AAGGAACCCAATGTTGTTAAACCACACGAAAAGGGAATAAATCCAAATTATCTTCTGTCATGGAAGCAAGTACTG GCAGAACTGCAGGACATTGAGAATGAGGAGAAAATTCCAAAGACTAAAAATATGAAGAAAAACAAGTCAGCTAAAG $\tt TGGGCACTTGGTGAGAAACATGGCACCAGAAAAGGTAGACTTTGACGACTGTAAAGATGTAGGTGATTTGAAGCAA$ TATGATAGTGATGAACCAGAATTGAGGTCGCTTGCAAGTTGGATTCAGAATGAGTTTAACAAGGCATGCGAACTG A CAGATTCAAGCTGGATAGAGCTCGATGAGATTGGAGAAGATGTGGCTCCAATTGAACACATTGCAAGCATGAGAGAGGGAAGGCGAAAGACCAACTTGTATGGTTTCATCATAAAAGGAAGATCCCACTTAAGGAATGACACCGACGTG GTAAACTTTGTGAGCATGGAGTTTTCTCTCACTGACCCAAGACTTGAACCACATAAATGGGAGAAGTACTGTGTT $\tt CTTGAGATAGGAGATATGCTTATAAGAAGTGCCATAGGCCAGGTTTCAAGGCCCATGTTCTTGTATGTGAGAACA$ ${\tt GAGAGTATGATTGAAGCTGAGTCCTCTGTCAAAGAGAAAGACATGACCAAAGAGTTCTTTGAGAACAAATCAGAA}$ ${\tt ACATGGCCCATTGGAGAGTCCCCCAAAGGAGTGGAGGAAAGTTCCATTGGGAAGGTCTGCAGGACTTTATTAGCA}$ ${\tt AAGTCGGTATTCAACAGCTTGTATGCATCTCCACAACTAGAAGGATTTTCAGCTGAATCAAGAAAACTGCTTCTT}$ ATCGTTCAGGCTCTTAGGGACAACCTTGAACCTGGGACCTTTGATCTTGGGGGGGCTATATGAAGCAATTGAGGAG $\tt TTGTGGCAGTGCTACTATTTGCTATCCATACTGTCCAAAAAAGTACCTTGTTTCTACT$

SEQUENCE: 10 (PB1, PR8-X)

AGCGAAAGCAGCAAACCATTTGAATGGATGTCAATCCGACCTTACTTTTCTTAAAAGTGCCAACACAAAATGCT ATAAGCACAACTTTCCCTTATACTGGAGACCCTCCTTACAGCCATGGGACAGGAACAGGATACACCATGGATACT GCTTTCCTTGAGGAATCCCATCCTGGTATTTTTGAAAACTCGTGTATTGAAACGATGGAGGTTGTTCAGCAAACA $\tt TTGGCCAACACAATAGAAGTGTTCAGATCAAATGGCCTCACGGCCAATGAGTCTGGAAGGCTCATAGACTTCCTT$ GACAATATGACTAAGAAAATGATAACACAGAGAACAATGGGTAAAAAGAAGCAGAGATTGAACAAAAGGAGTTAT $\tt CTAATTAGAGCATTGACCCTGAACACAATGACCAAAGATGCTGAGAGAGGGGAAGCTAAAACGGAGAGCAATTGCA$ ${\tt ACCCCAGGGATGCAAATAAGGGGGTTTGTATACTTTGTTGAGACACTGGCAAGGAGTATATGTGAGAAACTTGAA}$ CAATCAGGGTTGCCAGTTGGAGGCAATGAGAAGAAGCAAAGTTGGCAAATGTTGTAAGGAAGATGATGACCAAT ${\tt TCTCAGGACACCGAACTTTCTTCACCATCACTGGAGATAACACCAAATGGAACGAAAATCAGAATCCTCGGATG}$ $\tt TTTTTGGCCATGATCACATATATGACCAGAAATCAGCCCGAATGGTTCAGAAATGTTCTAAGTATTGCTCCAATA$ ATGTTCTCAAACAAAATGGCGAGACTGGGAAAAGGGTATATGTTTGAGAGCAAGAGTATGAAACTTAGAACTCAA $\tt ATACCTGCAGAAATGCTAGCAAGCATCGATTTGAAATATTTCAATGATTCAACAAGAAGAAGAAGATTGAAAAAATC$ ${\tt TCCTCTGACGATTTTGCTCTGATTGTGAATGCACCCAATCATGAAGGGATTCAAGCCGGAGTCGACAGGTTTTAT}$ CGAACCTGTAAGCTACTTGGAATCAATATGAGCAAGAAAAAGTCTTACATAAACAGAACAGGTACATTTGAATTC ${\tt ACAAGTTTTTCTATCGTTATGGGTTTGTTGCCAATTTCAGCATGGAGCTTCCCAGTTTTGGGGTTCTGGGATCTGGGATCTGGGATCTGGGATCTGGGATCTGGGATCTGGGATCTGGGATCTGGGGATCTGGAGATCTGGAGATTGGAGATTGGAGATCTGGAGATCTGGAGATCTGGAGATCTGGAGATGGAGATGGAGATGGAGATGGAGATGGAGATCTGGAGATGGAGATGGAGATGGAGATGGAGATGAGATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAGAATGAA$

SEQUENCES

SEQUENCE: 11 (PB2, PR8-X)

AGCGAAAGCAGGTCAATTATATTCAATATGGAAAGAATAAAAGAACTAAGAAATCTAATGTCGCAGTCTCGCACC AACCCAGCACTTAGGATGAAATGGATGATGGCAATGAAATATCCAATTACAGCAGACAAGAGGATAACGGAAATG $\tt TCACCTCTGGCTGTGACATGGTGGAATAGGAATGGACCAATAACAAATACAGTTCATTATCCAAAAATCTACAAAA$ ACTTATTTTGAAAGAGTAGAAAGGCTAAAGGCATGGAAGCTTTTGGCCCTGTCCATTTTAGAAACCCAGTCAAAATACGTCGGAGAGTTGACATAAATCCTGGTCATGCAGATCTCAGTGCCAAGGAGGCACAGGATGTAATCATGGAAGTT CAGATGTATACTCCAGGAGGGGAAGTGAGGAATGATGATGATCAAAGCTTGATTATTGCTGCTAGGAACATA GAAGAGGTGCTTACGGGAAATCTTCAAACATTGAAGATAAGAGTGCATGAGGGATATGAAGAGTTCACAATGGTT CAGTCGATTGCCGAAGCAATAATTGTGGCCATGGTATTTTCACAAGAGGATTGTATGATAAAAGCAGTCAGAGGT GATCTGAATTTCGTCAATAGGGCGAATCAGCGATTGAATCCTATGCATCAACTTTTAAGACATTTTCAGAAGGAT $\tt GCGAGAGTGCTTTTTCAAAATTGGGGAGTTGAACCTATCGACAATGTGATGGGAATGATTGGGATATTGCCCGAC$ GAGAGGGTAGTGGTGAGCATTGACCGTTTTTTGAGAATCCGGGACCAACGAGGAAATGTACTACTGTCTCCCGAG GAGGTCAGTGAAACACAGGGAACAGAGAAACTGACAATAACTTACTCATCGTCAATGATGTGGGAGATTAATGGT $\tt CCTGAATCAGTATTGGTCAATACCTATCAATGGATCATCAGAAACTGGGAAACTGTTAAAATTCAGTGGTCCCAG$ TACAGTGGGTTTGTAAGAACTCTGTTCCAACAAATGAGGGATGTGCTTGGGACATTTGATACCGCACAGATAATA GGATCAGGAATGAGAATACTTGTAAGGGGCAATTCTCCTGTATTCAACTATAACAAGGCCACGAAGAGACTCACA

SEQUENCES

GTTCTCGGAAAGGATGCTGGCACTTTAACTGAAGACCCAGATGAAGGCACAGCTGGAGTGGAGTCCGCTGTTCTG
AGGGGATTCCTCATTCTGGGCAAAGAAGACAAGAGATATGGGCCAGCACTAAGCATCAATGAACTGAGCAACCTT
GCGAAAGGAGAGAGGCTAATGTGCTAATTGGGCAAGGAGACGTGGTGTTGGTAATGAAACGGAAACGGGACTCT
AGCATACTTACTGACAGCCAGACAGCGACCAAAAGAATTCGGATGGCCATCAATTAGTGTCGAATAGTTTAAAAA
CGACCTTGTTTCTACT

SEQUENCE: 12 (NP, PR8-X)

GAACAGATGGAGACTGATGGAGAACGCCAGAATGCCACTGAAATCAGAGCATCCGTCGGAAAAATGATTGGTGGA ATTGGACGATTCTACATCCAAATGTGCACCGAACTCAAACTCAGTGATTATGAGGGACGGTTGATCCAAAACAGC $\tt GGAAAAGATCCTAAGAAAACTGGAGGACCTATATACAGGAGAGGTAAACGGAAAGTGGATGAGAGAACTCATCCTT$ ATGATGATCTGGCATTCCAATTTGAATGATGCAACTTATCAGAGGACAAGAGCTCTTGTTCGCACCGGAATGGAT CCCAGGATGTGCTCTCTGATGCAAGGTTCAACTCTCCCTAGGAGGTCTGGAGCCGCAGGTGCTGCAGTCAAAGGA GTTGGAACAATGGTGATGGAATTGGTCAGAATGATCAAACGTGGGATCAATGATCGGAACTTCTGGAGGGGTGAG AATGGACGAAAAACAAGAATTGCTTATGAAAGAATGTGCAACATTCTCAAAGGGAAATTTCAAACTGCTGCACAA A A A G C A A T G A T G G A T C A A G T G A G A G A G A G C C G G G A C C C A G G A T C T C G A G A T C T C A C T T T T C T A G C A ${\tt CAAGTGTACAGCCTAATCAGACCAAATGAGAATCCAGCACACAAGAGTCAACTGGTGTGGATGGCATGCCATTCT}$ GCCGCATTTGAAGATCTAAGAGTATTAAGCTTCATCAAAGGGACGAAGGTGCTCCCAAGAGGGAAGCTTTCCACT AGAGGAGTTCAAATTGCTTCCAATGAAAATATGGAGACTATGGAATCAAGTACACTTGAACTGAGAAGCAGGTAC ${\tt TGGGCCATAAGGACCAGAAGTGGAGGAAACACCAATCAACAGAGGGCATCTGCGGGCCAAATCAGCATACAACCT}$ AGAACATCTGACATGAGGACCGAAATCATAAGGATGATGGAAAGTGCAAGACCAGAAGATGTGTCTTTCCAGGGG

SEQUENCE: 13 (M, PR8-X)

TCTTATTTCTTCGGAGACAATGCAGAGGAGTACGACAATTAAAGAAAAATACCCTTGTTTCTACT

SEQUENCES

SEQUENCE: 14 (NS, PR8-X)

SEQUENCE: 15 (HA, PR8-X)

 $\tt AGCAAAAGCAGGGGAAAATAAAAACAACCAAAATGAAGGCAAACCTACTGGTCCTGTTATGTGCACTTGCAGCTG$ ${\tt CAGATGCAGACACTATGTATAGGCTACCATACGAACAATTCAACCGACACTGTTGACACAGTACTCGAGAAGA}$ $\tt ATGTGACAGTGACACTCTGTTAACCTGCTCGAAGACAGCCACAACGGAAAACTATGTAGATTAAAAGGAATAGAATAGAAATAGAATAGAATAGAATAGA$ CTGGGAGGATGAACTATTACTGGACCTTGCTAAAACCCGGAGACACAATAATATTTGAGGCAAATGGAAATCTAA AGTGTAACAGAAGTGTCAAACACCCCTGGGAGGCTATAAACAGCAGTCTCCCTTACCAGAATATACACCCAGTCAGGATTACAAACAAGGTGAACACTGTTATCGAGAAAATGAACATTCAATTCACAGCTGTGGGTAAAGAATTCAACA TAAAAAGCCAATTAAAGAATAATGCCAAAGAAATCGGAAATGGATGTTTTGAGTTCTACCACAAGTGTGACAATG AATGCATGGAAAGTGTAAGAAATGGGACTTATGATTATCCCAAATATTCAGAAGAGTCAAAGTTGAACAGGGAAA

SEQUENCES

 $\label{thm:constraint} \mbox{Aggta} \mbox{Aggta} \mbox{Aggta} \mbox{Carttaca} \mbox{Carttaca} \mbox{Carttaca} \mbox{Carttaca} \mbox{Carttaca} \mbox{Carttaca} \mbox{Carttaca} \mbox{Carttaca} \mbox{Cartaca} \mbox{Car$

SEQUENCE: 16 (NA, PR8-X)

 ${\tt ACCATACTGGAATATGCAACCAAAACATCATTACCTATAAAAATAGCACCTGGGTAAAGGACCACACTTCAGTGA}$ GTTCCAAAGGAGACGTTTTTGTCATAAGAGAGCCCTTTATTTCATGTTCTCACTTGGAATGCAGGACCTTTTTTC TGACCCAAGGTGCCTTACTGAATGACAAGCATTCAAGTGGGACTGTTAAGGACAGAAGCCCTTATAGGGCCTTAA $\tt GTCATGATGGCATGGCTGACAACAATCGGAATTTCAGGTCCAGATAATGGAGCAGTGGCTGTATTAAAATACA$ TAAATGGTTCATGTTTTACTATAATGACTGATGGCCCGAGTGATGGGCTCGCCTCGTACAAAATTTTCAAGATCG AAAAGGGGAAGGTTACTAAATCAATAGAGTTGAATGCACCTAATTCTCACTATGAGGAATGTTCCTGTTACCCTG ATACCGACAAAGTGATGTGTGTGCAGAGACAATTGGCATGGTTCGAACCGGCCATGGGTGTCTTTCGATCAAA ACCTGGATTATCAAATAGGATACATCTGCAGTGGGGTTTTCGGTGACAACCCGCGTCCCGAAGATGGAACAGGCA GCTGTGGTCCAGTGTATGTTGATGGAGCAAACGGAGTAAAGGGATTTTCATATAGGTATGGTAATGGTGTTTTGGA A TAGTA AGTT CTCTGTGAGGCAAGA TGTTGTGGCAA TGACTGATTGGTCAGGGTATAGCGGA AGTTTCGTTCAAC $\verb| ATCCTGAGCTGACAGGGCTAGACTGTATGAGGCCGTGCTTCTGGGGTTGAATTAATCAGGGGACGACCTAAAGAAAA$ AAACAATCTGGACTAGTGCGAGCAGCATTTCTTTTTGTGGCGTGAATAGTGATACTGTAGATTGGTCTTGGCCAG ACGGTGCTGAGTTGCCATTCAGCATTGACAAGTAGTCTGTTCAAAAAACTCCTTGTTTCTACT

SEQUENCE: 17 (PA, 105p30)

 ${\tt AGTATTTGCAACACCACAGGAGCTGAGAAACCAAAGTTTCTGCCAGATCTGTATGATTACAAAGAGAATAGGTTC}$ ATCGAAATTGGAGTGACAAGGAGAAGTTCACATATACTATCTGGAAAAGGCCAACAAAATTAAATCTGAGAAG ACACATATTCACATTTTCTCATTTACTGGCGAAGAATGGCCACAAAGGCCGATTACACTCTCGATGAAGAAAGC $\tt AGGGCTAGAATTAAAACCAGACTATTCACCATAAGGCAAGAAATGGCAAGCAGAGGTCTTTGGGACTCCTTTCGT$ CAGTCCGAAAGAGGCGAAGAGACAATTGAAGAAAGGTTTGAAATCACAGGGACAATGCGCAGGCTCGCTGATCAA ATTAGACTTCCGAATGGGCCTCCTTGTTTTCAGCGGTCAAAATTCCTGCTGATGGATTCTTTAAAATTAAGCATT GAGGATCCAAATCATGAAGGGGAGGGAATACCACTATATGATGCAATCAAGTGTATGAGAACATTCTTTGGATGG AAAGAACCCACTGTTGTCAAGCCACACGAGAAGGGAATAAATCCGAATTATCTGCTGTCGTGGAAGCAGGTGTTG GAAGACTGCAGGACATTGAGAGTGAGGAGAAGATTCCAAGAACAAAAAACATGAAAAAAACGAGTCAGTTAAAG

SEOUENCE: 18 (PB1, 105p30)

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SEQUENCES

 $\tilde{\mathsf{AGCGAAAGCAGCAAACCATTTGAATGGATGTCAATCCGACATTACTTTTCTTAAAAGTGCCAGCACAAAATGCT}$

A TA A GC A CA A CTTTTCCTTA TA CTGGTGA C CCTCCTTA CA GCCA TGGA A CA GGA A CA GGA TA CA CCA TGGA TA CA GCTTTCCTTGAAGAATCCCATCCTGGTATTTTTGAAAACTCTTGTATTGAAACAATGGAGGTTGTTCAGCAAACA AGGGTGGACAAACTGACACAAGGCAGACAAACCTATGACTGGACTCTAAATAGGAACCAGCCTGCTGCCACAGCA $\tt TTGGCAAACACCATAGAAGTATTCAGATCAAATGGCCTCATAGCAAATGAATCTGGAAGGCTAATAGACTTCCTT$ GACAATGTAACTAAAAAAATGGTGACCCAAAGAACAATAGGAAAAAAGAAACATAAATTAGACAAAAGAAGTTAC ${\tt CAATCAGGGTTGCCAGTTGGAGGAAATGAGAAAGCAAAGTTAGCAAATGTTGTAAGGAAGATGATGACCAAC}$ TCCCAGGACACTGAAATTTCTTTTACCATCACTGGAGATAACACAAAATGGAACGAAAATCAAAACCCTAGAATG TTCTTGGCCATGATCACATATATAACCAAAGATCAGCCTGAATGGTTCAGAAATATTCTAAGTATTGCTCCAATA ATGTTTTCAAACAAAATGGCGAGACTAGGTAGGGGGGTATATGTTTGAAAGCAAGAGTATGAAACTGAGAACCCAA $\tt ATACCTGCAGAGATGCTAGCCAACATAGATTTGAAATATTTCAATGATTCAACTAAAAAGAAAATTGAAAAAATT$ CGACCATTATTAATAGATGGAACTGCATCATTGAGTCCTGGAATGATGGTGGCCATGTTCAATATGTTAAGCACC TCGTCTGATGATTTTGCTTTGATTGTGAATGCACCCAATTATGCAGGAATTCAAGCTGGAGTTGACAGGTTTTAT ${\tt ACGAGCTTTTTCTATCGTTATGGGTTTGTTGCCAATTTCAGCATGGAGCTTCCTAGTTTTTGGGGTGTCTGGGGTC$

AATGAATCTGCAGACATGAGTATTGGAGTCACTGTCATCAAAAACAATATGATAAACAATGACCTTGGCCCAGCA
ACTGCTCAAATGGCCCTTCAGTTATTTATAAAAGATTACAGGTACACTTATCGATGCCACAGAGGTGACACACAA
ATACAAACCCGGAGATCATTTGAAATAAAGAAACTATGGGACCAAACCCGCTCCAAAGCTGGGCTGTTGGTCTCT

SEQUENCES

GATGGAGGCCCCAATTTATATAACATTAGGAATCTACATATTCCTGAAGTCTGCTTGAAATGGGAGTTGATGGAT
GAGGATTACCAGGGGCGTTTATGCAACCCATTGAACCCGTTTGTCAGCCATAAAGAGATTGAATCAGTGAACAAT
GCAGTGATAATGCCGGCCACAAGAATATGGAGTATGACGCTGTTGCAACAACACCACTCTTGGGTC
CCCAAAAGAAATCGATCCATTTTAAACACGAGCCAAAGAGGGATACTTGAAGATGAGCAAATGTACCAAAGGTGC
TGCAATTTATTTGAAAAATTCTTCCCCAAGTAGCTCATACAGAAGACCAGTTGGAATATCCAGTATGGTAGAGGCT
ATGGTTTCAAGAGCCCGAATTGATGCACGGATTGATTTCGAATCTGGAAGGATAAAGAAGAAGAAGAATTCGCTGAG
ATCATGAAGACCTGTTCCACCATTGAAGACCTCAGACGGCAAAAAATAGGGAATTTGGCTTGTCCTTCATGAAAAA
ATGCCTTGTTTCTACT

SEQUENCE: 19 (PB2, 105p30)

 $\tt AGCGAAAGCAGGTCAATTATATTCAATATGGAAAGAATAAAAGAGCTAAGGAATCTGATGTCACAATCTCGCACT$ CGCGAGATACTTACCAAAACTACTGTAGACCACATGGCCATAATAAAGAAATACACATCAGGAAGACAGGAGAAA ${\tt TCACCCCTAGCTGTGACATGGTGGAACAGAAATGGACCAGTGGCAAACACTATCCACTATCCAAAAAATCTACAAA$ ACTTACTTTGAAAAGGTTGAAAGGTTAAAACATGGAACCTTTGGCCCTGTACACTTTAGAAACCAAGTCAAAATA GTTTTCCCTA ATGA AGTGGGAGCCAGA ATA CTA A CATCAGAATCGCA ATTA ACGATA A CTA AGGAGAAA A A A GAG GAACTCCAGAATTCCCCTTTCCCCTTTGATGCTTCCCTTTCATCATCTCCTCAGACACTCTCCCCAAACCACAC TTTCTCCCGGTTGCAGGTGGAACAAGCAGTGTGTACATTGAAGTTTTGCATTTAACACAGGGGACATGCTGGGAG $\tt CAGATGTACACTCCAGGTGGGGAGGTGAGGAATGATGATGATGATCAAAGCCTAATTATTGCTGCTAGGAACATA$ GTGAGAAGAGCTGCAGTATCAGCAGATCCACTAGCATCTTTATTAGAAATGTGCCATAGCACACAGATTGGTGGA ACAAGGATGGTGGATATTCTCAGGCAAAATCCAACAGAAGAACAAGCTGTGGACATATGCAAAGCAGCAATGGGG GAAGAAGTGCTAACGGGCAATCTGCAAACATTGAAGCTAACTGTGCATGAGGGATATGAAGAATTCACAATAGTT GATCTGAATTTCGTTAATAGAGCGAATCAGCGGTTGAATCCCATGCATCAACTTTTGAGACATTTTCAGAAGGAT GCTAAAGTACTTTTCCTAAATTGGGGAATTGAACATATTGACAATGTGATGGGAATGATTGGGATATTACCTGAT GAAAGGGTAGTGGTAAGCATTGACCGTTTTTTGAGGGTCCGGGACCAAAGAGGAAATGTATTACTGTCTCCAGAG ${\tt GAAGTCAGTGAAACACAAGGAACAGGAAAACTGACAATAACTTACTCTTCATCATTGATGTGGGAGATTAATGGC}$ $\tt CCTGAGTCAGTGTTGATCAATACCTACCAATGGATCATCAGAAACTGGGAGACTGTTAAAATTCAGTGGTCTCAG$ AACCCTACAATGCTATACAATAAAATGGAATTTGAGCCATTTCAATCTCTAGTCCCCAAGGCCATTAGAGGCCAA TACAGTGGGTTTGTTAGAACTCTATTTCAACAAATGAGGGATGTGCTCGGGACCTTTGACACAACTCAGATAATA GGATCAGGAATGAGAATACTTGTAAGGGGTAATTCTCCAGTATTCAACTACAACAAGACCACTAAGAGACTCACA GCGAAAGGGGAGAAAGCTAATGTGCTAATTGGGCAAGGGGATGTAGTGTTGGTAATGAAACGAAAACGGGACTCT

SEQUENCES

SEQUENCE: 20 (NP, 105p30)

 ${\tt AGCAAAAGCAGGGTAGATAATCACTCACTGAGTGACATCAAAGTCATGGCGTCCCAAGGCACCAAACGGTCTTAC}$ GAACAGATGGAGACTGATGGGGAACGCCAGAATGCAACTGAAATCAGAGCATCCGTCGGAAGAATGATTGGGGGA ATTGGGCGATTCTACATCCAAATGTGCACCGAGCTTAAGCTCAATGATTATGAGGGACGACTGATCCAGAACAGC GGGAAAGATCCTAAGAAAACTGGAGGACCCATATACAAGAGAGTAGATGGAAAGTGGGTGAGGGAACTCGTCCTT TATGACAAAGAAGAATAAGGCGGATTTGGCGCCAAGCCAACAATGGTGATGATGCAACAGCTGGTTTGACTCAC $\tt CCCAGGATGTGCTCTTTGATGCAAGGTTCAACTCTCCCTAGAAGATCTGGAGCAGCAGGCGCTGCAGTCAAAGGA$ AATGGGAGAAAAACAAGGATTGCTTATGAGAGAATGTGCAACATTCTCAAAGGAAAATTTCAAACAGCTGCACAA GCCAGTGGGTATGACTTCGAAAAAGAGGGATACTCTTTGGTGGGAGTAGACCCTTTCAAACTGCTTCAAACCAGT AGAGGAGTACAAATTGCTTCAAATGAAAACATGGATGCTATTGTCTCAAGTACTCTTGAACTGAGAAGCAGATAC $\tt TGGGCCATAAGAACCAGAAGTGGAGGGAACACCAATCAACAAAGGGCCTCTGCGGGCCAAATCAGCACAACCT$ ACGTTTTCTGTGCAGAGAAACCTCCCATTTGACAAAACAACCATCATGGCAGCATTCACTGGGAATACAGAGGGA AGAACATCAGACATGCGGGCAGAAATCATAAAGATGATGGAAAGTGCAAGACCAGAAGAAGTGTCCTTCCAGGGA CGGGGAGTCTTTGAGCTCTCGGACGAAAGGGCAACGAACCCGATCGTGCCCTCCTTTGACATGAGTAATGAAGGA TCTTATTTCTTCGGAGACAATGCAGAGGAGTACGACAATTAATGAAAAATACCCTTGTTTCTACT

SEQUENCE: 21 (M, 105p30)

SEQUENCES

SEQUENCE: 23 (HA, 105p30)

 $\tt AGCGAAAGCAGGGGAAAATAAAAGCAACCAAAATGAAAGTAAAACTACTGGTTCTGTTATGTACATTTACAGCTA$ CATATGCAGACACAATATGTATAGGCTACCATGCCAACACTCAACCGACACTGTTGACACAGTACTTGAGAAGA ATGTAACAGTGACACACTCTGTCAACCTACTTGAGGACAGTCACAATGGAAAACTATGTCTACTAAAAGGAATAG AGGAATCATGGTCCTACATTGTAGAAACACCAAATCCTGAGAATGGAACATGTTACCCAGGGTATTTCGCCGACT GGCCCAACCACACCGTAACCGGAGTATCAGCATCATGCTCCCATAACGGGAAAAGCAGTTTTTACAGAAATTTGC TCTCTGTAGTGTCTTCACATTATAGCAGAAGATTCACCCCAGAAATAGCCAAAAGACCCAAGGTGAGAGACCAGG AAGGAAGAATCAACTACTGCTGGACTCTGCTGGAACCCGGGGATACAATAATATTTGAGGCAAATGGAAATCTAA CAATAGGAGAGTGTCCAAAGTATGTCAGGAGTGCAAAATTAAGGATGGTTACAGGACTAAGGAACATCCCATCCA GGATTACAAACAAGGTGAATTCTGTAATTGAGAAAATGAACACTCAATTCACAGCTGTGGGCAAAGAATTCAACA AATTGGAAAGAAGGATGGAAAACTTAAATAAAAAAGTTGATGATGGTTTCTAGACATTTGGACCTATAATGCAG TAAAAAGCCAATTAAAGAATAATGCCAAAGAAATAGGAAACGGGTGTTTTGAATTCTATCACAAGTGTAACGATG AATGCATGGAGAGTGTGAAAAATGGAACTTATGACTATCCAAAATATTCCGAAGAATCAAAGTTAAACAGAGAGA ${\tt AAATTGATGGAGTGAAATTGGAATCAATGGGAGTCTATCAGATTCTGGCGATCTACTCAACAGTCGCCAGTTCCCAGTTCCCAGTTCAGTTC$ TCTAAGACCAGAATTTCAGAAATATAAGGAAAAACACCCTTGTTTCTACT

SEQUENCES

SEQUENCE: 24 (NA, 105p30) TAATTAGTCTAATGTTGCAAATAGGAAATATTATTTCAATATGGGCTAGTCACTCAATCCAAACTGGAAGTCAAA ${\tt ACCACACTGGAATATGCAACCAAAAAATCATCACATATGAAAAACAGCACCTGGGTGAATCACACATATGTTAATA}$ ${\tt TTAACAACATGTTGTTGCTGGAAAGGACAAAACTTCAGTGACACTGGCCGGCAATTCATCTCTTTGTCCTA}$ CATACAATTCAAAGTTTGAATCAGTTGCATGGTCAGCAAGCGCATGCCATGATGGCAAGGGCTGGTTAACAATCG GAATTTCTGGTCCAGACAATGGAGCTGTGGCTGTACTAAAATACAACGGAATAATAACTGAAACCATAAAAAGTT GGGAAAAGCGAATATTGAGAACACAAGAGTCTGAATGTGTTTGTGTGAACGGGTCATGTTTCACCATAATGACCG TGAATGCACCCAATTTTCATTATGAGGAATGTTCCTGTTACCCAGACACTGGCACAGTGATGTTGTTATGCAGGG A CAACTGGCATGGTTCAAATCGACCTTGGGTATCTTTTAATCAAAACTTGGATTATCAAATAGGATACATCTGCAAGGGGTTTGAGATGATTTGGGATCCTAATGGATGGACAGATACCGACAGTGATTTCTCAGTGAAACAGGATGTTG TGGCAATAACTGATTGGTCAGGGTACAGCGGAAGTTTCGTCCAACATCCTGAGTTAACAGGATTGGACTGTATAA GACCTTGCTTCTGGGTTGAGTTAGTCAGAGGACTGCCTAGAGAAAATACAACAATCTGGACTAGTGGGAGCAGCA TTTCTTTTTGTGGCGTTGATAGTGATACTGCAAATTGGTCTTGGCCAGACGGTGCTGAGTTGCCGTTCACCATTG ACAAGTAGCTCGTTGAAAAAAACTCCTTGTTTCTACT

SEQUENCE: 25 (HA, A/Chile/1/1983)
MKAKLLVLLCALSATDADTICIGYHANNSTDTVDTVLEKNVTVTHSVNLLEDNHNGKLCKLKGIAPLQLGKCSIA
GWILGNPECESLFSKKSWSYIAETPNSENGTCYPGYFADYEELREQLSSVSSFERFEIFPKESSWPKHNVTKGVT
AACSHKGKSSFYRNLLWLTEKNGSYPNLSKSYVNNKEKEVLVLWGVHHPSNIEDQKTIYRKENAYVSVVSSHYNR
RFTPEIAKRPKVRNQEGRINYYWTLLEPGDTIIFEANGNLIAPWYAFALSRGFGSGIITSNASMDECDAKCQTPQ
GAINSSLPFQNVHPVTIGECPKYVRSTKLRMVTGLRNIPSIQSRGLFGAIAGFIEGGWTGMIDGWYGYHHQNEQG
SGYAADQKSTQNAINGITNKVNSIIEKMNTQFTAVGKEENKLEKRMENLNKKVDDGFLDIWTYNAELLVLLENER
TLDFHDSNVKNLYEKVKSQLKNNAKEIGNGCFEFYHKCNNECMESVKNGTYDYPKYSEESKLNREKIDGVKLESM
GVYQILAIYSTVASSLVLLVSLGAISFWMCSNGSLQCRICI

SEQUENCE: 26 (NA, A/Chile/1/1983)
MNPNQKIITIGSICMTIGIISLILQIGNIISIWVSHSIQTGSQNHTGICNQRIITYENSTWVNQTYVNINNTNVV
AGKDTTSVTLAGNSSLCPIRGWAIYSKDNSIRIGSKGDVFVIREPFISCSHLECRTFFLTQGALLNDKHSNGTVK
DRSPYRALMSCPIGEAPSPYNSRFESVAWSASACHDGMGWLTIGISGPDDGAVAVLKYNGIITETIKSWRKRILR
TQESECVCVNGSCFTIMTDGPSNGPASYRIFKIEKGKITKSIELDAPNSHYEECSCYPDTGTVMCVCRDNWHGSN
RPWVSFNQNLDYQIGYICSGVFGDNPRPKDGKGSCDPVTVDGADGVKGFSYRYGNGVWIGRTKSNSSRKGFEMIW
DPNGWTDTDSNFLVKQDVVAMTDWSGYSGSFVQHPELTGLDCMRPCFWVELVRGRPREGTTVWTSGSSISFCGVN
SDTANWSWPDGAELPFTIDK

SEQUENCE: 27 (NA, A/California/04/09)
MNPNQKIITIGSVCMTIGMANLILQIGNIISIWISHSIQLGNQNQIETCNQSVITYENNTWVNQTYVNISNTNFA
AGQSVVSVKLAGNSSLCPVSGWAIYSKDNSVRIGSKGDVFVIREPFISCSPLECRTFFLTQGALLNDKHSNGTIK

SEQUENCES

DRSPYRTLMSCPIGEVPSPYNSRFESVAWSASACHDGINWLTIGISGPDNGAVAVLKYNGIITDTIKSWRNNILR
TQESECACVNGSCFTVMTDGPSNGQASYKIFRIEKGKIVKSVEMNAPNYHYEECSCYPDSSEITCVCRDNWHGSN
RPWVSFNQNLEYQIGYICSGIFGDNPRPNDKTGSCGPVSSNGANGVKGFSFKYGNGVWIGRTKSISSRNGFEMIW
DPNGWTGTDNNFSIKQDIVGINEWSGYSGSFVQHPELTGLDCIRPCFWVELIRGRPKENTIWTSGSSISFCGVNS
DTVGWSWPDGAELPFTIDK

SEQUENCE: 28 (encodes the same amino acid sequence as SEQUENCE: 3) GATCATATGGCGATTATTAAAAAATATACCAGCGGCCGCCAGGAAAAAAACCCGAGCCTGCGCATGAAATGGATG $\tt ATGGCGATGAAATATCCGATTACCGCGGATAAACGCATTACCGAAATGATTCCGGAACGCAACGAACAGGGCCAG$ $\tt CGCAACGGCCCGGTGGCGAGCACCATTCATTATCCGAAAATTTATAAAACCTATTTTGAAAAAGTGGAACGCCTG$ AGCGTGTATATTGAAGTGCTGCATCTGACCCAGGGCACCTGCTGGGAACAGATGTATACCCCGGGCGGCGAAGTG $\tt AACCCGACCGAAGAACAGCCGGTGGATATTTGCAAAGCCGCGATTGGCGCTTTAGCAGCAGCTTTAGCTTT$ GGCGGCTTTACCTTTAAACGCACCAGCGGCAGCAGCGTGAAACGCGAAGAAGTGCTGACCGGCAACCTGCAG AAAGCGACCCGCCGCCTGATTCAGCTGATTGTGAGCGGCCGCGATGAACAGAGCATTGTGGAAGCGATTGTGGTG GCGATGGTGTTTAGCCAGGAAGATTGCATGGTGAAAGCGGTGCGCGGCGATCTGAACTTTGTGAACCGCGCGAAC $\tt CAGCGCCTGAACCCGATGCATCAGCTGCTGCGCCATTTTCAGAAAGATGCGAAAGTGCTGTTTCTGAACTGGGGC$ $\tt CGCGGCGTGCGCGTGAGCAAAATGGGCGTGGATGAATATAGCAACGCGGAACGCGTGGTGAGCATTGATCGC$ $\tt CAGTGGATTATTCGCAACTGGGAAACCGTGAAAATTCAGTGGAGCCAGAACCGACCATGCTGTATAACAAAATG$ GAATTTGAACCGTTTCAGAGCCTGGTGCCGAAAGCGATTCGCGGCCAGTATAGCGGCTTTGTGCGCACCCTGTTT CAGCAGATGCGCGATGTGCTGGGCACCTTTGATACCACCCAGATTATTAAACTGCTGCCGTTTGCGGCGGCGCCC GGCAACAGCCCGGTGTTTAACTATAACAAAACCACCAAACGCCTGACCGTGCTGGGCAAAGATGCGGGCACCCTG ${\tt ACCGAAGATCCGGATGAAGGCACCGCGGGCGTGGAAAGCGCGGTGCTGCGCGGCTTTCTGATTCTGGGCAAAGAA}$ GATCGCCGCTATGGCCCGGCGCTGAGCATTAACGAACTGAGCAACCTGGCGAAAGGCGAAAAAGCGAACGTGCTG ATTGGCCAGGGCGATGTGGTGGTGATGAAACGCAAACGCGATAGCAGCATTCTGACCGATAGCCAGACCGCG ACCAAACGCATTCGCATGGCGATTAAC

SEQUENCES

SEQUENCE: 29 (PA, A/New Caledonia/20/1999)
medfvrqcfnpmivelaekamkeygedpkietnkfaaicthlevcfmysdfhfidergesiivesgdpnallkhr
feiiegrdrimawtvvnsicnttgvekpkflpdlydykenrfieigvtrrevhiyylekankiksekthihifsf
tgeematkadytldeesrariktrlftirqemasrslwdsfrqsergeetieekfeitgtmrkladqslppnfps
lenfrayvdgfepngciegklsqmskevnakiepflrttprplrlpdgplchqrskfllmdalklsiedpshege
giplydaikcmktffgwkepnivkphekginpnylmawkqvlaelqdieneekiprtknmkrtsqlkwalgenma
pekvdfddckdvgdlkqydsdepeprslaswvqnefnkaceltdsswieldeigedvapiehiasmrrnyftaev
shcrateyimkgvyintallnascaamddfqlipmiskcrtkegrrktnlygfiikgrshlrndtdvvnfvsmef
sltdprlephkwekycvleigdmllrtaigqvsrpmflyvrtngtskikmkwgmemrrcllqslqqiesmieaes
svkekdmtkeffenksetwpigesprgveegsigkvcrtllaksvfnslyaspqlegfsaesrklllivqalrdn
lepgtfdlgglyeaieeclindpwvllnaswfnsflthalk

SEQUENCE: 30 (PB1, A/New Caledonia/20/1999)
mdvnptllflkvpaqnaisttfpytgdppyshgtgtgytmdtvnrthqysergrwtkntetgapqlnpidgplpk
dnepsgyaqtdcvleamafleeshpgifenscietmevvqqtrvdkltqgrqtydwtlnrnqpaatalantievf
rsnglianesgrlidflkdvmesmdrdevevtthfqrkrrvrdnvtkkmvtqrtigkkkhkldkrsyliraltln
tmtkdaergklkrraiatpgmqirgfvyfvetlarsicekleqsglpvggnekkaklanvvrkmmtnsqdteisf
titgdntkwnenqnprmflamityitknqpewfrnilsiapimfsnkmarlgkgymfesksmklrtqipaemlan
idlkyfndstkrkiekirpllidgtaslspgmmmgmfnmlstvlgvsilnlgqkrytkttywwdglqssddfali
vnapnyagiqagvdrfyrtckllginmskkksyinrtgtfeftsffyrygfvanfsmelpsfgvsgvnesadmsi
gvtviknnminndlgpataqmalqlfikdyrytyrchrgdtqiqtrrsfeikklwdqtrskagllvsdggpnlyn
irnlhipevclkwelmdedyggrlcnpsnpfvshkeiesvnnavmmpahgpaknmeydavatthswvpkrnrsil
ntsqrgiledeqmyqrccnlfekffpsssyrrpvgissmveamvsraridaridfesgrikkeefaeimktcsti
edlrrqk

SEQUENCE: 31 (PB2, A/New Caledonia/20/1999)
merikelrnlmsqsrtreiltkttvdhmaiikkytsgrqeknpslrmkwmmamkypitadkritemiperneqgq
tlwskvndagsdrvmisplavtwwnrngpvastihypkiyktyfekverlkhgtfgpvhfrnqvkirrrvdinpg
hadlsakeaqdvimevvfpnevgariltsesqltitkekkeelqnckisplmvaymlerelvrktrflpvaggts
svyievlhltqgtcweqmytpggevrnddvdqsliiaarnivrraavsadplasllemchstqiggtrmvdilrq
npteeqavdickaamglrisssfsfggftfkrtsgssvkreeevltgnlqtlkltvhegyeeftmvgkratailr
katrrliqlivsgrdeqsiveaivvamvfsqedcmvkavrgdlnfvnranqrlnpmhqllrhfqkdakvlflnwg
iepidnvmgmigilpdmtpstemsmrgvrvskmgvdeysnaervvvsidrflrvrdqrgnvllspeevsetqgte
kltitysssmmweingpesvlintyqwiirnwetvkiqwsqnptmlynkmefepfqslvpkairgqysgfvrtlf
qqmrdvlgtfdttqiikllpfaaappkqsrmqfssltvnvrgsgmrilvrgnspvfnynkttkrltvlgkdagtl
tedpdegtagvesavlrgflilgkedrrygpalsinelsnlakgekanvligqgdvvlvmkrkrdssiltdsqta
tkrirmain

SEQUENCE: 32 (NP, A/New Caledonia/20/1999)
masqqtkrsyeqmetdgerqnateirasvgrmiggigrfyiqmctelklndyegrliqnsltiermvlsafderr
nkyleehpsagkdpkktggpiykrvdgkwvrelvlydkeeirriwrqanngddataglthimiwhsnlndttyqr
tralvrtgmdprmcslmqgstlprrsgaagaavkgvgtmvlelirmikrgindrnfwrgengrktriayermcni
lkgkfqtaaqkammdqvresrnpgnaeiedltflarsalilrgsvahksclpacvygpavasgydfekegyslvg
vdpfkllqtsqvyslirpnenpahksqlvwmacnsaafedlrvssfirgtrvlprgklstrgvqiasnenmdaiv

SEQUENCES

 $sstlelrs rywair trsggntnqqras agqistqptfsvqrnlpfdkttima aftgntegrtsdmraeiikmmes \\ arpeevsfqgrgvfelsder atnpivpsfdmsnegsyffgdnaeeydn$

SEQUENCE: 33 (M1, A/New Caledonia/20/1999)
mslltevetyvlsivpsgplkaeiaqrlenvfagkntdlealmewlktrpilspltkgilgfvftltvpserglq
rrrfvqnalngngdpnnmdravklyrklkreitfhgakeialsysagalascmgliynrmgavttesafglicat
ceqiadsqhkshrqmvtttnplirhenrmvlasttakameqmagsseqaaeamevasqarqmvqamraigthpss
stglkndllenlqayqkrmgvqmqrfk

SEQUENCE: 34 (NA, A/New Caledonia/20/1999)
mnpnqkiitigsisiaigiislmlqigniisiwashsiqtgsqnhtgvcnqriityenstwvnhtyvninntnvv
agkdktsvtlagnsslcsisgwaiytkdnsirigskgdvfvirepfiscshlecrtffltqgallndkhsngtvk
drspyralmscplgeapspynskfesvawsasachdgmgwltigisgpdngavavlkyngiitetikswkkrilr
tqesecvcvngscftimtdgpsngaasykifkiekgkvtksielnapnfhyeecscypdtgtvmcvcrdnwhgsn
rpwvsfnqnldyqigyicsgvfgdnprpkdgegscnpvtvdgadgvkgfsykygngvwigrtksnrlrkgfemiw
dpngwtdtdsdfsvkqdvvaitdwsgysgsfvqhpeltgldcirpcfwvelvrglprenttiwtsgssisfcgvn
sdtanwswpdgaelpftidk

SEQUENCE: 35 (PA, A/Wisconsin/67/2005) medfvrqcfnpmivelaekamkeygedlkietnkfaaicthlevcfmysdfhfineqgesivvelddpnallkhr feiiegrdrtmawtvvnsicnttgagkpkflpdlydykenrfieigvtrrevhiyylekankiksenthihifsf tgeematkadytldeesrariktrlftirqemanrglwdsfrqsergeetieekfeitgtmrrladqslppnfsc lenfrayvdgfepngciegklsqmskevnaqiepflkttprpiklpngppcyqrskfllmdalklsiedpshege giplydaikcmktffgwkepyivkphekginsnyllswkqvlselqdieneekiprtknmkktsqlkwalgenma pekvdfencrdisdlkqydsdepelrslsswiqnefnkaceltdsvwieldeigedvapiehiasmrrnyftaev shcrateyimkgvyintallnascaamddfqlipmiskcrtkegrrktnlygfiikgrshlrndtdvvnfvsmef sltdprlephkwekycvleigdmllrsaigqisrpmflyvrtngtskvkmkymmemrrcllqslqqiesmieaes svkekdmtkeffenkseawpigespkgveegsigkvcrtllaksvfnslyaspqlegfsaesrklllvvqalrdn lepgtfdlgglyeaieeclindpwvllnaswfnsflthalk

SEQUENCE: 36 (PB1, A/Wisconsin/67/2005) mdvnptllflkvpaqnaisttfpytgdppyshgtgtgytmdtvnrthqysekgkwttntetgapqlnpidgplpe dnepsgyaqtdcvleamafleeshpgifenscletmeavqqtrvdrltqgrqtydwtlnrnqpaatalantievf rsngltanesgrlidflkdvmesmdkeemeitthfqrkrrvrdnmtkkmvtqrtigkkkqrvnkrgyliraltln tmtkdaergklkrraiatpgmqirgfvyfvetlarsicekleqsglpvggnekkaklanvvrkmmtnsqdtelsf titgdntkwnenqnprmflamityitknqpewfrnilsiapimfsnkmarlgkgymfeskrmklrtqipaemlas idlkyfnestrkkiekirpllidgtaslspgmmmgmfnmlstvlgvsilnlgqkkytkttywwdglqssddfali vnapnhegiqagvnrfyrtcklvginmskkksyinktgtfeftsffyrygfvanfsmelpsfgvsginesadmsi gvtviknnminndlgpataqmalqlfikdyrytyrchrgdtqiqtrrsfelkklwdqtqsragllvsdggpnlyn irnlhipevclkwelmdenyrgrlcnplnpfvshkeiesvnnavvmpahgpaksmeydavatthswipkrnrsil ntsqrgiledeqmyqkccnlfekffpsssyrrpigissmveamvsraridaridfesgrikkeefseimkicsti eelrrqr

SEQUENCE: 37 (PB2, A/Wisconsin/67/2005)
merikelrnlmsqsrtreiltkttvdhmaiikkytsgrqeknpslrmkwmmamkypitadkritemvperneqgq
tlwskmsdagsdrvmvsplavtwwnrngpvtstvhypkvyktyfdkverlkhgtfgpvhfrnqvkirrrvdinpg

SEQUENCES

hadlsakeaqdvimevvfpnevgariltsesqltitkekkeelrdckisplmvaymlerelvrktrflpvaggts siyievlhltqgtcweqmytpggevrnddvdqsliiaarnivrraavsadplasllemchstqiggtrmvdilrq npteeqavdickaamglrisssfsfggftfkrtsgssvkkeeevltgnlqtlkirvhegyeeftmvgkratailr katrrlvqlivsgrdeqsiaeaiivamvfsqedcmikavrgdlnfvnranqrlnpmhqllrhfqkdakvlfqnwg iehidsvmgmvgvlpdmtpstemsmrgirvskmgvdeysstervvvsidrflrvrdqrgnvllspeevsetqgte rltitysssmmweingpesvlvntyqwiirnweavkiqwsqnpamlynkmefepfqslvpkairsqysgfvrtlf qqmrdvlgtfdttqiikllpfaaappkqsrmqfssltvnvrgsgmrilvrgnspvfnynkttkrltilgkdagtl iedpdestsgvesavlrgfliigkedrrygpalsinelsnlakgekanvligqgdvvlvmkrkrdssiltdsqta tkrirmain

SEQUENCE: 38 (NP, A/Wisconsin/67/2005) masggtkrsyeqmetdgdrqnateirasvgkmidgigrfyiqmctelklsdyegrliqnsltiekmvlsafderr nkyleehpsagkdpkktggpiyrrvdgkwmrelvlydkeeirriwrqanngedataglthimiwhsnlndatyqr tralvrtgmdprmcslmqgstlprrsgaagaavkgigtmvmelirmvkrgindrnfwrgengrktrsayermcni lkgkfqtaaqramvdqvresrnpgnaeiedliflarsalilrgsvahksclpacvygpavssgynfekegyslvg idpfkllqnsqvyslirpnenpahksqlvwmachsaafedlrllsfirgtkvsprgklstrgvqiasnenmdnmg sgtlelrsgywairtrsggntnqqrasagqtsvqptfsvqrnlpfekstimaaftgntegrtsdmraeiirmmeg akpeevsfrggyfelsdekatnpivpsfdmsnegsyffgdnaeeydn

 $\label{eq:sequence:39} SEQUENCE: 39 \ (M1, A/Wisconsin/67/2005) \\ mslltevetyvlsivpsgplkaeiaqrledvfagkntdlealmewlktrpilspltkgilgfvftltvpserglq \\ rrrfvqnalngngdpnnmdkavklyrklkreitfhgakeialsysagalascmgliynrmgavttevafglvcat \\ ceqiadsqhrshrqmvattnplirhenrmvlasttakameqmagsseqaaeameiasqarqmvqamraigthpss \\ stglrddllenlqtyqkrmgvqmqrfk$

 $\label{eq:sequence: 40 (M2, A/Wisconsin/67/2005)} \\ \text{mslltevetpirnewgcrcndssdplvvaaniigilhlilwildrlffkcvyrlfkhglkrgpstegvpesmree} \\ \text{yrkeqqnavdaddshfvsiele}$

SEQUENCE: 42 (HA, A/Wisconsin/67/2005)
mktiialsyilclvfaqklpgndnstatlclghhavpngtivktitndqievtnatelvqssstggicdsphqil
dgenctlidallgdpqcdgfqnkkwdlfverskaysncypydvpdyaslrslvassgtlefndesfnwtgvtqng

SEQUENCES

 ${\tt tsssckrrsnnsffsrlnwlthlkfkypalnvtmpnnekfdklyiwgvhhpvtdndqiflyaqasgritvstkrs}$ qqtvipnigsrprirnipsrisiywtivkpgdillinstgnliaprgyfkirsgkssimrsdapigkcnsecitp ng sipndk pfqnvnrity gac pryvkqntlklat gmrnvpekqtrgif gai ag fiengwegmvdgwygfrhqnse ${\tt gigq}$ aadlk ${\tt stq}$ aain ${\tt qing}$ kln ${\tt rlig}$ ktnekfh ${\tt qie}$ kef ${\tt sevegriq}$ dlek ${\tt yve}$ dtkidl ${\tt wsynaellvalenq}$ $\verb|htidltdsemnkl| fertkkqlrenaedmgngcfkiyhkcdnacigsirngtydhdvyrdealnnrfqikgvelks|$ gykdwilwisfaiscfllcvallgfimwacqkgnircnici

SEQUENCE: 43 (NA, A/Wisconsin/67/2005) $\verb|mnpnqk| iitigsvs| tisticff mqiailittvt| thfkqyefnsppnnqvm| ceptilernite ivy| tnttiek$ eicpklaeyrnwskpqcnitgfapfskdnsirlsaggdiwvtrepyvscdpdkcyqfalgqgttlnnvhsndtvh drtpyrtllmnelgvpfhlgtkqvciawsssschdgkawlhvcvtgddknatasfiyngrlvdsivswskeilrt $\tt qesecvcingtctvvmtdgsasgkadtkilfieegkivhtstlsgsaqhveecscyprylgvrcvcrdnwkgsnr$ pivdinikdysivssyvcsglvgdtprkndssssshcldpnneegghgvkgwafddgndvwmgrtiseklrsgye tfkviegwsnpnsklqinrqvivdrgnrsgysgifsvegkscinrcfyvelirgrkeetevlwtsnsivvfcgts

gtygtgswpdgadinlmpi

SEQUENCE: 44 (PA. 105p30)

 $\verb|medfvrqcfnpmivelaekam| keygedpkietnkfaaicthlevcfmysdfhfidergesiivesgdpnallkhr|$ feiiegrdrimawtvinsicnttgvekpkflpdlydykenrfieigvtrrevhiyylekankiksekthihifsf tgeematkadytldeesrariktrlftirqemaskslwdsfrqsergeetieekfeitgtmrkladqslppnfps lenfrayvdg fepngciegkl sqmskevnakiepflrttprplrlpdgplchqrskfllmdalklsiedpshege ${ t giply} { t daikc}$ mkt ${ t ff} { t gwkepnivk} { t phekginpnylmawkqvlaelq} { t dieneekiprtknmkrt} { t sqlkwalgenma}$ pekvdfddckdvgdlkqydsdepeprslaswvqnefnkaceltdsswieldeigedvapiehias mrrnyftaevshcrateyimk gvyintallnas caamddfqlipmisk crtkegrrktnlyg fiikgrshlrndt dvvn fvsmefund skalled find the statement of the stat ${\tt sltdprlephkwekycvleigdmllrtaigqvsrpmflyvrtngtskikmkwgmemrrcllqslqqiesmieaes}$ ${\tt svkekdmtkeffenksetwpigesprgveegsigkvcrtllaksvfnslyaspqlegfsaesrklllivqalrdn}$ lepgtfdlgglyeaieeclindpwvllnaswfnsflthalk

SEQUENCE: 45 (M1, 105p30)

 $\verb|ms|| tevetyv| sivpsgp| kaeiaqr| tenvfagkntdlea| mew| ktrpilsp| tkgilgfv| ftltvpserg| quality tensor for the context of the$ rrrfvqnalngngdpnnmdkavklyrklkreitfhgakeialsysagalascmgliynrmgavttesafglicat ${\tt ceqiadsqhkshrqmvtttnplirhenrmvlasttakameqmagsseqaaeamevasqarqmvqamraigthpss}$ stglkndllenlqayqkrmgvqmqrfk

SEQUENCE: 46 (A/Texas/1/77 PB1)

 $\verb|mdvnptl|| flkipaqnaisttfpytgdppyshgtgtgytmdtvnrthqysekgkwttntetgapqlnpidgplpe||$ $\tt dnepsgyaqtdcvleamafleeshpgifenscletmevvqqtrvdrltqgrqtydwtlnrnqpaatalantievf$ rsnglt an esgrlid flk dv mesm dke ei eit th fqrkrrvrdnmtkk mvtqrtigkkkqrvnkrsyliral tln $\verb|tmtkda| ergklk| rraiatpgmqirgfvyfvetlarsicekleqsglpvggnekkaklanvvrkmmtnsqdtelsf|$ $\verb|titgdntkw| nenqnprmflamity| itknqpewfrnilsiapimfsnkm| arlgkgymfeskrmklrtqipaemlas$ idlky fnestrkkie kirpllidgt asl spgmmgmfnml stvlgv silnlgqkkytkt tywwdglqssdd faliing specifically and the strength of the s ${\tt vnapnhegiqagvdrfyrtcklvginmskkksyinrtgtfeftsffyrygfvanfsmelpsfgvsginesadmsi}$ gvtviknn minndlg pataq malqlfik dyrytyrchrg dtqiqtrrsfelkkl weqtrsk agllvs dggpnlynirnlhipevclkwelmdedyqgrlcnplnpfvshkeiesvnnavvmpahgpaksmeydavatthswipkrnrsil

SEQUENCES

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SEQUENCE: 47 (A/Puerto Rico/8/34 PA) $\verb|medfvrqcfnpmivelaektm| keygedlkietn k faaict hlevcfmysdfh fin eqgesiivelgdpn all khrist hand beginning to be a simple of the contract of t$ feiiegrdrtmawtvvnsicnttgaekpkflpdlydykenrfieigvtrrevhiyylekankiksekthihifsf ${\sf tgeematkadytldees}$ rariktrlftirqemasrglwdsfrqsergeetieerfeitgtmrkladqslppnfss lenfrayvdgfepngyiegklsqmskevnariepflkttprplrlpngppcsqrskfllmdalklsiedpshege giplydaikcmrtffgwkepnvvkphekginpnyllswkqvlaelqdieneekipktknmkktsqlkwalgenma pekvdfddckdvgdlkqydsdepelrslaswiqnefnkaceltdsswieldeigedvapiehiasmrrnyftsev shcrateyimkgvyintallnascaamddfqlipmiskcrtkegrrktnlygfiikgrshlrndtdvvnfvsmef sltdprlephkwekycvleigdmlirsaigqvsrpmflyvrtngtskikmkwgmemrrcllqslqqiesmieaes ${\tt svkekdmtkeffenksetwpigespkgveessigkvcrtllaksvfnslyaspqlegfsaesrklllivqalrdn}$

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Val Asp Thr Val Leu Glu Lys Asn Val Thr Val Thr His Ser Val Asn	_	a Lys Leu L 5	eu Val Leu	_	Leu Ser Ala	_	
Leu Leu Glu Asp Asn His Asn Gly Lys Leu Cys Lys Leu Lys Gly Ile 50 Ser Glu Leu Cys Lys Leu Lys Gly Ile Ala Pro Leu Gln Leu Gly Lys Cys Ser Ile Ala Gly Trp Ile Leu Gly 80 Ser Pro Glu Cys Glu Ser Leu Phe Ser Lys Lys Lys Ser Trp Ser Tyr Ile 85 Ser Glu Asn Gly Thr Cys Tyr Pro Gly Tyr Phe 100 Ser Tyr Glu Glu Leu Arg Glu Gln Leu Ser Ser Val Ser Ser Phe 115 Ser Ser Phe 130 Phe Glu Ile Phe Pro Lys Glu Ser Ser Trp Pro Lys His Asn 130 Phe Glu Ile Phe Pro Lys Glu Ser Ser His Lys Gly Lys Ser Ser 145 Ser Ser Leu Trp Leu Thr Glu Lys Asn Gly Ser Tyr Pro 170 Ser Tyr Pro 180 Ser Ser	Ala Asp Th	_	le Gly Tyr			Asp Thr	
Ala Pro Leu Gln Leu Gly Lys Cys Ser Ile Ala Gly Trp Ile Leu Gly 80 Asn Pro Glu Cys Glu Ser Leu Phe Ser Lys Lys Ser Trp Ser Tyr Ile 95 Ala Glu Thr Pro Asn Ser Glu Asn Gly Thr Cys Tyr Pro Gly Tyr Phe 110 Ala Asp Tyr Glu Glu Leu Arg Glu Gln Leu Ser Ser Val Ser Ser Phe 125 Glu Arg Phe Glu Ile Phe Pro Lys Glu Ser Ser Trp Pro Lys His Asn 135 Val Thr Lys Gly Val Thr Ala Ala Cys Ser His Lys Gly Lys Ser Ser 160 Phe Tyr Arg Asn Leu Leu Trp Leu Thr Glu Lys Glu Lys Gly Val Leu Val	_	r Val Leu G	=	Val Thr Val		Val Asn	
Asn Pro Glu Cys Glu Ser Leu Phe Ser Lys Lys Ser Trp Ser Tyr Ile 95 Ala Glu Thr Pro Asn Ser Glu Asn Gly Thr Cys Tyr Pro Gly Tyr Phe 110 Ala Asp Tyr Glu Glu Leu Arg Glu Gln Leu Ser Ser Val Ser Ser Phe 115 Glu Arg Phe Glu Ile Phe Pro Lys Glu Ser Ser Trp Pro Lys His Asn 130 Val Thr Lys Gly Val Thr Ala Ala Cys Ser His Lys Gly Lys Ser Ser 160 Phe Tyr Arg Asn Leu Leu Trp Leu Thr Glu Lys Glu Lys Gly Val Leu Val		u Asp Asn H	_	Lys Leu Cys		Gly Ile	
Ala Glu Thr Pro Asn Ser Glu Asn Gly Thr Cys Tyr Pro Gly Tyr Phe 110 Ala Asp Tyr Glu Glu Leu Arg Glu Gln Leu Ser Ser Val Ser Ser Phe 125 Ser Phe 130 Pro Lys Glu Ser Ser Trp Pro Lys His Asn 130 Pro Lys Gly Val Thr Ala Ala Cys Ser His Lys Gly Lys Ser Ser 160 Phe Tyr Arg Asn Leu Leu Trp Leu Thr Glu Lys Glu Lys Gly Val Leu Val		u Gln Leu G 7	ly Lys Cys O	Ser Ile Ala 75	Gly Trp Ile	e Leu Gly 80	
Ala Asp Tyr Glu Glu Leu Arg Glu Gln Leu Ser Ser Val Ser Ser Phe 115	Asn Pro Gl	-	er Leu Phe		Ser Trp Ser	_	
Glu Arg Phe Glu Ile Phe Pro Lys Glu Ser Ser Trp Pro Lys His Asn 130	Ala Glu Th		er Glu Asn			-	
130			_	Gln Leu Ser		Ser Phe	
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165 170 175 Asn Leu Ser Lys Ser Tyr Val Asn Asn Lys Glu Lys Glu Val Leu Val	_	-		-	Lys Gly Lys		
	Phe Tyr Ar	-	eu Trp Leu	_	Asn Gly Ser	_	
	Asn Leu Se	_	yr Val Asn	_	_		

Leu Trp Gly Val His His Pro Ser Asn Ile Glu Asp Gln Lys Thr Ile

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Glu Gly	Arg	Ile	Asn 245	Tyr	Tyr	Trp	Thr	Leu 250	Leu	Glu	Pro	Gly	Asp 255	Thr
Ile Ile	Phe	Glu 260	Ala	Asn	Gly	Asn	Leu 265	Ile	Ala	Pro	Trp	Tyr 270	Ala	Phe
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Ser Ser 305	Leu	Pro	Phe	Gln 310	Asn	Val	His	Pro	Val 315	Thr	Ile	Gly	Glu	Cys 320
Pro Lys	Tyr	Val	Arg 325	Ser	Thr	Lys	Leu	Arg 330	Met	Val	Thr	Gly	Leu 335	Arg
Asn Ile	Pro	Ser 340	Ile	Gln	Ser	Arg	Gly 345	Leu	Phe	Gly	Ala	Ile 350	Ala	Gly
Phe Ile	Glu 355	Gly	Gly	Trp	Thr	Gly 360	Met	Ile	Asp	Gly	Trp 365	Tyr	Gly	Tyr
His His 370	Gln	Asn	Glu	Gln	Gly 375	Ser	Gly	Tyr	Ala	Ala 380	Asp	Gln	Lys	Ser
Thr Gln 385	Asn	Ala	Ile	Asn 390	Gly	Ile	Thr	Asn	Lys 395	Val	Asn	Ser	Ile	Ile 400
Glu Lys	Met	Asn	Thr 405	Gln	Phe	Thr	Ala	Val 410	Gly	Lys	Glu	Phe	Asn 415	Lys
Leu Glu	Lys	Arg 420	Met	Glu	Asn	Leu	Asn 425	Lys	Lys	Val	Asp	Asp 430	Gly	Phe
Leu Asp	Ile 435	Trp	Thr	Tyr	Asn	Ala 440	Glu	Leu	Leu	Val	Leu 445	Leu	Glu	Asn
Glu Arg 450	Thr	Leu	Asp	Phe	His 455	Asp	Ser	Asn	Val	Lys 460	Asn	Leu	Tyr	Glu
Lys Val 465	Lys	Ser	Gln	Leu 470	Lys	Asn	Asn	Ala	Lys 475	Glu	Ile	Gly	Asn	Gly 480
Cys Phe	Glu	Phe	Tyr 485	His	Lys	Cys	Asn	Asn 490	Glu	Cys	Met	Glu	Ser 495	Val
Lys Asn	Gly	Thr 500	Tyr	Asp	Tyr	Pro	Lys 505	Tyr	Ser	Glu	Glu	Ser 510	Lys	Leu
Asn Arg	Glu 515	Lys	Ile	Asp	Gly	Val 520	Lys	Leu	Glu	Ser	Met 525	Gly	Val	Tyr
Gln Ile 530	Leu	Ala	Ile	Tyr	Ser 535	Thr	Val	Ala	Ser	Ser 540	Leu	Val	Leu	Leu
Val Ser 545	Leu	Gly	Ala	Ile 550	Ser	Phe	Trp	Met	Сув 555	Ser	Asn	Gly	Ser	Leu 560
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Trp	Val	Ser 35	His	Ser	Ile	Gln	Thr 40	Gly	Ser	Gln	Asn	His 45	Thr	Gly	Ile
Cys	Asn 50	Gln	Arg	Ile	Ile	Thr 55	Tyr	Glu	Asn	Ser	Thr 60	Trp	Val	Asn	Gln
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Asp	Val	Phe 115	Val	Ile	Arg	Glu	Pro 120	Phe	Ile	Ser	CÀa	Ser 125	His	Leu	Glu
CÀa	Arg 130	Thr	Phe	Phe	Leu	Thr 135	Gln	Gly	Ala	Leu	Leu 140	Asn	Asp	ГЛа	His
Ser 145	Asn	Gly	Thr	Val	Lys 150	Asp	Arg	Ser	Pro	Tyr 155	Arg	Ala	Leu	Met	Ser 160
Cys	Pro	Ile	Gly	Glu 165	Ala	Pro	Ser	Pro	Tyr 170	Asn	Ser	Arg	Phe	Glu 175	Ser
Val	Ala	Trp	Ser 180	Ala	Ser	Ala	Cys	His 185	Asp	Gly	Met	Gly	Trp 190	Leu	Thr
Ile	Gly	Ile 195	Ser	Gly	Pro	Asp	Asp 200	Gly	Ala	Val	Ala	Val 205	Leu	Lys	Tyr
Asn	Gly 210	Ile	Ile	Thr	Glu	Thr 215	Ile	ГÀв	Ser	Trp	Arg 220	ГÀа	Arg	Ile	Leu
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Ile	Met	Thr	Asp	Gly 245	Pro	Ser	Asn	Gly	Pro 250	Ala	Ser	Tyr	Arg	Ile 255	Phe
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Val	Phe	Gly	Asp	Asn 325	Pro	Arg	Pro	Lys	330 Asp	Gly	Lys	Gly	Ser	Сув 335	Asp
Pro	Val	Thr	Val 340	Asp	Gly	Ala	Asp	Gly 345	Val	Lys	Gly	Phe	Ser 350	Tyr	Arg
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Ser 385	Asn	Phe	Leu	Val	190 390	Gln	Asp	Val	Val	Ala 395	Met	Thr	Asp	Trp	Ser 400
Gly	Tyr	Ser	Gly	Ser 405	Phe	Val	Gln	His	Pro 410	Glu	Leu	Thr	Gly	Leu 415	Asp
CAa	Met	Arg	Pro	Cha	Phe	Trp	Val	Glu	Leu	Val	Arg	Gly	Arg	Pro	Arg

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COILCIIIaCa

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Cya	Asn 50	Gln	Ser	Val	Ile	Thr 55	Tyr	Glu	Asn	Asn	Thr 60	Trp	Val	Asn	Gln
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Val	Ser	Val	Lys	Leu 85	Ala	Gly	Asn	Ser	Ser 90	Leu	CAa	Pro	Val	Ser 95	Gly
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Asp	Val	Phe 115	Val	Ile	Arg	Glu	Pro 120	Phe	Ile	Ser	CAa	Ser 125	Pro	Leu	Glu
Cys	Arg 130	Thr	Phe	Phe	Leu	Thr 135	Gln	Gly	Ala	Leu	Leu 140	Asn	Asp	Lys	His
Ser 145	Asn	Gly	Thr	Ile	Lys 150	Asp	Arg	Ser	Pro	Tyr 155	Arg	Thr	Leu	Met	Ser 160
Cys	Pro	Ile	Gly	Glu 165	Val	Pro	Ser	Pro	Tyr 170	Asn	Ser	Arg	Phe	Glu 175	Ser
Val	Ala	Trp	Ser 180	Ala	Ser	Ala	Cha	His 185	Asp	Gly	Ile	Asn	Trp 190	Leu	Thr
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Asn	Gly 210	Ile	Ile	Thr	Asp	Thr 215	Ile	Lys	Ser	Trp	Arg 220	Asn	Asn	Ile	Leu
Arg 225	Thr	Gln	Glu	Ser	Glu 230	CÀa	Ala	Cys	Val	Asn 235	Gly	Ser	CÀa	Phe	Thr 240
Val	Met	Thr	Asp	Gly 245	Pro	Ser	Asn	Gly	Gln 250	Ala	Ser	Tyr	Lys	Ile 255	Phe
Arg	Ile	Glu	Lys 260	Gly	Lys	Ile	Val	Lys 265	Ser	Val	Glu	Met	Asn 270	Ala	Pro
Asn	Tyr	His 275	Tyr	Glu	Glu	CÀa	Ser 280	СЛа	Tyr	Pro	Asp	Ser 285	Ser	Glu	Ile
Thr	Сув 290	Val	Сув	Arg	Asp	Asn 295	Trp	His	Gly	Ser	Asn 300	Arg	Pro	Trp	Val
Ser 305	Phe	Asn	Gln	Asn	Leu 310	Glu	Tyr	Gln	Ile	Gly 315	Tyr	Ile	Сув	Ser	Gly 320

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1200

1260

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Asn Lys Pho	e Ala Ala	a Ile Cys	Thr Hi:	5 Leu Glu	Val Cys Ph 45	e Met Tyr	
Ser Asp Pho	∋ His Ph⊖	e Ile Ası 55	Glu Ar	g Gly Glu	Ser Ile Il	e Val Glu	
	o Pro Asr		ı Leu Lv:	s His Ara	Phe Glu Il	e Ile Glu	
65		70	2	75		80	
Gly Arg As	Arg Ile 85	e Met Ala	Trp Th	r Val Val 90	Asn Ser Il	e Cys Asn 95	
Thr Thr Gl	y Val Glu 100	ı Lys Pro	Lys Pho		Asp Leu Ty		
Lys Glu Ası 11!		e Ile Glu	lle Gl	y Val Thr	Arg Arg Gl 125	u Val His	
Ile Tyr Ty: 130	r Leu Glu	ı Lys Ala	_	s Ile Lys	Ser Glu Ly 140	s Thr His	
Ile His Ile 145	e Phe Sei	Phe Thi	Gly Gl	ı Glu Met 155	Ala Thr Ly	s Ala Asp 160	
Tyr Thr Let	ı Asp Glı 169		Arg Ala	a Arg Ile 170	Lys Thr Ar	g Leu Phe 175	

Thr Ile Arg Gln Glu Met Ala Ser Arg Ser Leu Trp Asp Ser Phe Arg 180 $$185\$

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Arg	Ser	Leu	Ala	Ser 405	Trp	Val	Gln	Asn	Glu 410	Phe	Asn	ГÀа	Ala	Cys 415	Glu
Leu	Thr	Asp	Ser 420	Ser	Trp	Ile	Glu	Leu 425	Asp	Glu	Ile	Gly	Glu 430	Asp	Val
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Met	Phe	Leu	Tyr	Val 565	Arg	Thr	Asn	Gly	Thr 570	Ser	Lys	Ile	Lys	Met 575	ГЛа
Trp	Gly	Met	Glu 580	Met	Arg	Arg	СЛв	Leu 585	Leu	Gln	Ser	Leu	Gln 590	Gln	Ile
Glu	Ser	Met 595	Ile	Glu	Ala	Glu	Ser 600	Ser	Val	Lys	Glu	Lys	Asp	Met	Thr
Lys	Glu	Phe	Phe	Glu	Asn	Lys	Ser	Glu	Thr	Trp	Pro	Ile	Gly	Glu	Ser

_															
	610					615					620				
Pro 625	Arg	Gly	Val	Glu	Glu 630	Gly	Ser	Ile	Gly	Lys 635	Val	CÀa	Arg	Thr	Leu 640
Leu	Ala	ràs	Ser	Val 645	Phe	Asn	Ser	Leu	Tyr 650	Ala	Ser	Pro	Gln	Leu 655	Glu
Gly	Phe	Ser	Ala 660	Glu	Ser	Arg	Lys	Leu 665	Leu	Leu	Ile	Val	Gln 670	Ala	Leu
Arg	Asp	Asn 675	Leu	Glu	Pro	Gly	Thr 680	Phe	Asp	Leu	Gly	Gly 685	Leu	Tyr	Glu
Ala	Ile 690	Glu	Glu	CAa	Leu	Ile 695	Asn	Asp	Pro	Trp	Val 700	Leu	Leu	Asn	Ala
Ser 705	Trp	Phe	Asn	Ser	Phe 710	Leu	Thr	His	Ala	Leu 715	ГЛа				
) NO H: 75												
		PE: RGANI		Inf	luenz	za A	viru	ıs							
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Gly	Thr	Gly 35	Thr	Gly	Tyr	Thr	Met 40	Asp	Thr	Val	Asn	Arg 45	Thr	His	Gln
Tyr	Ser 50	Glu	Arg	Gly	Arg	Trp 55	Thr	Lys	Asn	Thr	Glu 60	Thr	Gly	Ala	Pro
Gln 65	Leu	Asn	Pro	Ile	Asp 70	Gly	Pro	Leu	Pro	Lys 75	Asp	Asn	Glu	Pro	Ser 80
Gly	Tyr	Ala	Gln	Thr 85	Asp	Cys	Val	Leu	Glu 90	Ala	Met	Ala	Phe	Leu 95	Glu
Glu	Ser	His	Pro 100	Gly	Ile	Phe	Glu	Asn 105	Ser	Сла	Ile	Glu	Thr 110	Met	Glu
Val	Val	Gln 115	Gln	Thr	Arg	Val	Asp 120	Lys	Leu	Thr	Gln	Gly 125	Arg	Gln	Thr
Tyr	Asp 130	Trp	Thr	Leu	Asn	Arg 135	Asn	Gln	Pro	Ala	Ala 140	Thr	Ala	Leu	Ala
Asn 145	Thr	Ile	Glu	Val	Phe 150	Arg	Ser	Asn	Gly	Leu 155	Ile	Ala	Asn	Glu	Ser 160
Gly	Arg	Leu	Ile	Asp 165	Phe	Leu	ГЛа	Asp	Val 170	Met	Glu	Ser	Met	Asp 175	Arg
Asp	Glu	Val	Glu 180	Val	Thr	Thr	His	Phe 185	Gln	Arg	ГЛа	Arg	Arg 190	Val	Arg
Asp	Asn	Val 195	Thr	ГÀв	ГÀв	Met	Val 200	Thr	Gln	Arg	Thr	Ile 205	Gly	Lys	Lys
Lys	His 210	Lys	Leu	Asp	Lys	Arg 215	Ser	Tyr	Leu	Ile	Arg 220	Ala	Leu	Thr	Leu
Asn 225	Thr	Met	Thr	Lys	Asp 230	Ala	Glu	Arg	Gly	Lys 235	Leu	Lys	Arg	Arg	Ala 240
Ile	Ala	Thr	Pro	Gly 245	Met	Gln	Ile	Arg	Gly 250	Phe	Val	Tyr	Phe	Val 255	Glu
Thr	Leu	Ala	Arg 260	Ser	Ile	Сув	Glu	Lys 265	Leu	Glu	Gln	Ser	Gly 270	Leu	Pro

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Met	Ile	Thr	Tyr	Ile 325	Thr	Lys	Asn	Gln	Pro 330	Glu	Trp	Phe	Arg	Asn 335	Ile
Leu	Ser	Ile	Ala 340	Pro	Ile	Met	Phe	Ser 345	Asn	Lys	Met	Ala	Arg 350	Leu	Gly
Lys	Gly	Tyr 355	Met	Phe	Glu	Ser	Lys 360	Ser	Met	Lys	Leu	Arg 365	Thr	Gln	Ile
Pro	Ala 370	Glu	Met	Leu	Ala	Asn 375	Ile	Asp	Leu	Lys	Tyr 380	Phe	Asn	Asp	Ser
Thr 385	ГЛа	Arg	ГÀа	Ile	Glu 390	LÀa	Ile	Arg	Pro	Leu 395	Leu	Ile	Asp	Gly	Thr 400
Ala	Ser	Leu	Ser	Pro 405	Gly	Met	Met	Met	Gly 410	Met	Phe	Asn	Met	Leu 415	Ser
Thr	Val	Leu	Gly 420	Val	Ser	Ile	Leu	Asn 425	Leu	Gly	Gln	Lys	Arg 430	Tyr	Thr
ГÀв	Thr	Thr 435	Tyr	Trp	Trp	Asp	Gly 440	Leu	Gln	Ser	Ser	Asp 445	Asp	Phe	Ala
Leu	Ile 450	Val	Asn	Ala	Pro	Asn 455	Tyr	Ala	Gly	Ile	Gln 460	Ala	Gly	Val	Asp
Arg 465	Phe	Tyr	Arg	Thr	Cys 470	ГÀз	Leu	Leu	Gly	Ile 475	Asn	Met	Ser	ГÀв	Lys 480
ГÀз	Ser	Tyr	Ile	Asn 485	Arg	Thr	Gly	Thr	Phe 490	Glu	Phe	Thr	Ser	Phe 495	Phe
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Gln 545	Met	Ala	Leu	Gln	Leu 550	Phe	Ile	Lys	Asp	Tyr 555	Arg	Tyr	Thr	Tyr	Arg 560
CÀa	His	Arg		Asp 565	Thr	Gln	Ile		Thr 570		Arg	Ser	Phe	Glu 575	Ile
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Asp	Gly	Gly 595	Pro	Asn	Leu	Tyr	Asn 600	Ile	Arg	Asn	Leu	His 605	Ile	Pro	Glu
Val	Cys 610	Leu	ГÀа	Trp	Glu	Leu 615	Met	Asp	Glu	Asp	Tyr 620	Gln	Gly	Arg	Leu
Cys 625	Asn	Pro	Ser	Asn	Pro 630	Phe	Val	Ser	His	Lys 635	Glu	Ile	Glu	Ser	Val 640
Asn	Asn	Ala	Val	Met 645	Met	Pro	Ala	His	Gly 650	Pro	Ala	Lys	Asn	Met 655	Glu
Tyr	Asp	Ala	Val 660	Ala	Thr	Thr	His	Ser 665	Trp	Val	Pro	Lys	Arg 670	Asn	Arg
Ser	Ile	Leu 675	Asn	Thr	Ser	Gln	Arg 680	Gly	Ile	Leu	Glu	Asp	Glu	Gln	Met
Tyr	Gln	Arg	Cya	CÀa	Asn	Leu	Phe	Glu	Lys	Phe	Phe	Pro	Ser	Ser	Ser

												COII	CIII	aca	
	690					695					700				
Tyr 705	Arg	Arg	Pro	Val	Gly 710	Ile	Ser	Ser	Met	Val 715	Glu	Ala	Met	Val	Ser 720
Arg	Ala	Arg	Ile	Asp 725	Ala	Arg	Ile	Asp	Phe 730	Glu	Ser	Gly	Arg	Ile 735	ГÀа
Lys	Glu	Glu	Phe 740	Ala	Glu	Ile	Met	Lys 745	Thr	Сла	Ser	Thr	Ile 750	Glu	Asp
Leu	Arg	Arg 755	Gln	Lys											
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ГЛа	Tyr	Thr 35	Ser	Gly	Arg	Gln	Glu 40	Lys	Asn	Pro	Ser	Leu 45	Arg	Met	Lys
Trp	Met 50	Met	Ala	Met	Lys	Tyr 55	Pro	Ile	Thr	Ala	Asp 60	ГÀа	Arg	Ile	Thr
Glu 65	Met	Ile	Pro	Glu	Arg 70	Asn	Glu	Gln	Gly	Gln 75	Thr	Leu	Trp	Ser	80 Lys
Val	Asn	Asp	Ala	Gly 85	Ser	Asp	Arg	Val	Met 90	Ile	Ser	Pro	Leu	Ala 95	Val
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ГÀа	Ile	Tyr 115	Lys	Thr	Tyr	Phe	Glu 120	Lys	Val	Glu	Arg	Leu 125	Lys	His	Gly
Thr	Phe 130	Gly	Pro	Val	His	Phe 135	Arg	Asn	Gln	Val	Lys 140	Ile	Arg	Arg	Arg
Val 145	Asp	Ile	Asn	Pro	Gly 150	His	Ala	Asp	Leu	Ser 155	Ala	Lys	Glu	Ala	Gln 160
Asp	Val	Ile	Met	Glu 165	Val	Val	Phe	Pro	Asn 170	Glu	Val	Gly	Ala	Arg 175	Ile
			180					185					190	Glu	
Leu	Gln	Asn 195	CÀa	ГЛа	Ile	Ser	Pro 200	Leu	Met	Val	Ala	Tyr 205	Met	Leu	Glu
Ī	210				-	215					220		_	Gly	
Ser 225	Ser	Val	Tyr	Ile	Glu 230	Val	Leu	His	Leu	Thr 235	Gln	Gly	Thr	Cys	Trp 240
Glu	Gln	Met	Tyr	Thr 245	Pro	Gly	Gly	Glu	Val 250	Arg	Asn	Asp	Asp	Val 255	Asp
Gln	Ser	Leu	Ile 260	Ile	Ala	Ala	Arg	Asn 265	Ile	Val	Arg	Arg	Ala 270	Ala	Val
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Ser	Ser	Phe	Ser	Phe 325	Gly	Gly	Phe	Thr	Phe 330	Lys	Arg	Thr	Ser	Gly 335	Ser
Ser	Val	Lys	Arg 340	Glu	Glu	Glu	Val	Leu 345	Thr	Gly	Asn	Leu	Gln 350	Thr	Leu
Lys	Leu	Thr 355	Val	His	Glu	Gly	Tyr 360	Glu	Glu	Phe	Thr	Met 365	Val	Gly	Lys
Arg	Ala 370	Thr	Ala	Ile	Leu	Arg 375	Lys	Ala	Thr	Arg	Arg 380	Leu	Ile	Gln	Leu
Ile 385	Val	Ser	Gly	Arg	Asp 390	Glu	Gln	Ser	Ile	Val 395	Glu	Ala	Ile	Val	Val 400
Ala	Met	Val	Phe	Ser 405	Gln	Glu	Asp	Cys	Met 410	Val	Lys	Ala	Val	Arg 415	Gly
Asp	Leu	Asn	Phe 420	Val	Asn	Arg	Ala	Asn 425	Gln	Arg	Leu	Asn	Pro 430	Met	His
Gln	Leu	Leu 435	Arg	His	Phe	Gln	Lys 440	Asp	Ala	Lys	Val	Leu 445	Phe	Leu	Asn
Trp	Gly 450	Ile	Glu	Pro	Ile	Asp 455	Asn	Val	Met	Gly	Met 460	Ile	Gly	Ile	Leu
Pro 465	Asp	Met	Thr	Pro	Ser 470	Thr	Glu	Met	Ser	Met 475	Arg	Gly	Val	Arg	Val 480
Ser	Lys	Met	Gly	Val 485	Asp	Glu	Tyr	Ser	Asn 490	Ala	Glu	Arg	Val	Val 495	Val
Ser	Ile	Asp	Arg 500	Phe	Leu	Arg	Val	Arg 505	Asp	Gln	Arg	Gly	Asn 510	Val	Leu
Leu	Ser	Pro 515	Glu	Glu	Val	Ser	Glu 520	Thr	Gln	Gly	Thr	Glu 525	Lys	Leu	Thr
Ile	Thr 530	Tyr	Ser	Ser	Ser	Met 535	Met	Trp	Glu	Ile	Asn 540	Gly	Pro	Glu	Ser
Val 545	Leu	Ile	Asn	Thr	Tyr 550	Gln	Trp	Ile	Ile	Arg 555	Asn	Trp	Glu	Thr	Val 560
ГÀа	Ile	Gln	Trp	Ser 565	Gln	Asn	Pro	Thr	Met 570	Leu	Tyr	Asn	ГÀа	Met 575	Glu
Phe	Glu	Pro	Phe 580	Gln	Ser	Leu	Val	Pro 585	Lys	Ala	Ile	Arg	Gly 590	Gln	Tyr
Ser		Phe 595		Arg	Thr		Phe 600		Gln	Met		Asp 605	Val	Leu	Gly
Thr	Phe 610	Asp	Thr	Thr	Gln	Ile 615	Ile	Lys	Leu	Leu	Pro 620	Phe	Ala	Ala	Ala
Pro 625	Pro	Lys	Gln	Ser	Arg 630	Met	Gln	Phe	Ser	Ser 635	Leu	Thr	Val	Asn	Val 640
Arg	Gly	Ser	Gly	Met 645	Arg	Ile	Leu	Val	Arg 650	Gly	Asn	Ser	Pro	Val 655	Phe
Asn	Tyr	Asn	Lys	Thr	Thr	Lys	Arg	Leu 665	Thr	Val	Leu	Gly	Lys 670	Asp	Ala
Gly	Thr	Leu 675	Thr	Glu	Asp	Pro	Asp 680	Glu	Gly	Thr	Ala	Gly 685	Val	Glu	Ser
Ala	Val 690	Leu	Arg	Gly	Phe	Leu 695	Ile	Leu	Gly	Lys	Glu 700	Asp	Arg	Arg	Tyr
Gly 705	Pro	Ala	Leu	Ser	Ile 710	Asn	Glu	Leu	Ser	Asn 715	Leu	Ala	Lys	Gly	Glu 720
Lys	Ala	Asn	Val	Leu	Ile	Gly	Gln	Gly	Asp	Val	Val	Leu	Val	Met	Lys

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Ala Phe Glu Asp Leu Arg Val Ser Ser Phe Ile Arg Gly Thr Arg Val

Leu Pro Arg Gly Lys Leu Ser Thr Arg Gly Val Gln Ile Ala Ser Asn Glu Asn Met Asp Ala Ile Val Ser Ser Thr Leu Glu Leu Arg Ser Arg Tyr Trp Ala Ile Arg Thr Arg Ser Gly Gly Asn Thr Asn Gln Gln Arg Ala Ser Ala Gly Gln Ile Ser Thr Gln Pro Thr Phe Ser Val Gln Arg Asn Leu Pro Phe Asp Lys Thr Thr Ile Met Ala Ala Phe Thr Gly Asn Thr Glu Gly Arg Thr Ser Asp Met Arg Ala Glu Ile Ile Lys Met Met Glu Ser Ala Arg Pro Glu Glu Val Ser Phe Gln Gly Arg Gly Val Phe 455 Glu Leu Ser Asp Glu Arg Ala Thr Asn Pro Ile Val Pro Ser Phe Asp Met Ser Asn Glu Gly Ser Tyr Phe Phe Gly Asp Asn Ala Glu Glu Tyr 490 Asp Asn <210> SEQ ID NO 33 <211> LENGTH: 252 <212> TYPE: PRT <213> ORGANISM: Influenza A virus <400> SEQUENCE: 33 Met Ser Leu Leu Thr Glu Val Glu Thr Tyr Val Leu Ser Ile Val Pro Ser Gly Pro Leu Lys Ala Glu Ile Ala Gln Arg Leu Glu Asn Val Phe 25 Ala Gly Lys Asn Thr Asp Leu Glu Ala Leu Met Glu Trp Leu Lys Thr Arg Pro Ile Leu Ser Pro Leu Thr Lys Gly Ile Leu Gly Phe Val Phe Thr Leu Thr Val Pro Ser Glu Arg Gly Leu Gln Arg Arg Arg Phe Val Gln Asn Ala Leu Asn Gly Asn Gly Asp Pro Asn Asn Met Asp Arg Ala Val Lys Leu Tyr Arg Lys Leu Lys Arg Glu Ile Thr Phe His Gly Ala Lys Glu Ile Ala Leu Ser Tyr Ser Ala Gly Ala Leu Ala Ser Cys Met Gly Leu Ile Tyr Asn Arg Met Gly Ala Val Thr Thr Glu Ser Ala Phe 135 Gly Leu Ile Cys Ala Thr Cys Glu Gln Ile Ala Asp Ser Gln His Lys 150 Ser His Arg Gln Met Val Thr Thr Thr Asn Pro Leu Ile Arg His Glu 170 Asn Arg Met Val Leu Ala Ser Thr Thr Ala Lys Ala Met Glu Gln Met 185 Ala Gly Ser Ser Glu Gln Ala Ala Glu Ala Met Glu Val Ala Ser Gln 200

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Ala Arg Gln Met Val Gln Ala Met Arg Ala Ile Gly Thr His Pro Ser 210 215 Ser Ser Thr Gly Leu Lys Asn Asp Leu Leu Glu Asn Leu Gln Ala Tyr Gln Lys Arg Met Gly Val Gln Met Gln Arg Phe Lys 245 <210> SEQ ID NO 34 <211> LENGTH: 470 <212> TYPE: PRT <213 > ORGANISM: Influenza A virus <400> SEQUENCE: 34 Met Asn Pro Asn Gln Lys Ile Ile Thr Ile Gly Ser Ile Ser Ile Ala Ile Gly Ile Ile Ser Leu Met Leu Gln Ile Gly Asn Ile Ile Ser Ile Trp Ala Ser His Ser Ile Gln Thr Gly Ser Gln Asn His Thr Gly Val Cys Asn Gln Arg Ile Ile Thr Tyr Glu Asn Ser Thr Trp Val Asn His Thr Tyr Val Asn Ile Asn Asn Thr Asn Val Val Ala Gly Lys Asp Lys Thr Ser Val Thr Leu Ala Gly Asn Ser Ser Leu Cys Ser Ile Ser Gly Trp Ala Ile Tyr Thr Lys Asp Asn Ser Ile Arg Ile Gly Ser Lys Gly 105 Asp Val Phe Val Ile Arg Glu Pro Phe Ile Ser Cys Ser His Leu Glu 120 Cys Arg Thr Phe Phe Leu Thr Gln Gly Ala Leu Leu Asn Asp Lys His 135 Ser Asn Gly Thr Val Lys Asp Arg Ser Pro Tyr Arg Ala Leu Met Ser Cys Pro Leu Gly Glu Ala Pro Ser Pro Tyr Asn Ser Lys Phe Glu Ser 170 Val Ala Trp Ser Ala Ser Ala Cys His Asp Gly Met Gly Trp Leu Thr 185 Ile Gly Ile Ser Gly Pro Asp Asn Gly Ala Val Ala Val Leu Lys Tyr Asn Gly Ile Ile Thr Glu Thr Ile Lys Ser Trp Lys Lys Arg Ile Leu Arg Thr Gln Glu Ser Glu Cys Val Cys Val Asn Gly Ser Cys Phe Thr Ile Met Thr Asp Gly Pro Ser Asn Gly Ala Ala Ser Tyr Lys Ile Phe Lys Ile Glu Lys Gly Lys Val Thr Lys Ser Ile Glu Leu Asn Ala Pro Asn Phe His Tyr Glu Glu Cys Ser Cys Tyr Pro Asp Thr Gly Thr Val 280 Met Cys Val Cys Arg Asp Asn Trp His Gly Ser Asn Arg Pro Trp Val 295 Ser Phe Asn Gln Asn Leu Asp Tyr Gln Ile Gly Tyr Ile Cys Ser Gly 310 315 Val Phe Gly Asp Asn Pro Arg Pro Lys Asp Gly Glu Gly Ser Cys Asn 330

Pro Val Thr Val Asp Gly Ala Asp Gly Val Lys Gly Phe Ser Tyr Lys Tyr Gly Asn Gly Val Trp Ile Gly Arg Thr Lys Ser Asn Arg Leu Arg Lys Gly Phe Glu Met Ile Trp Asp Pro Asn Gly Trp Thr Asp Thr Asp Ser Asp Phe Ser Val Lys Gln Asp Val Val Ala Ile Thr Asp Trp Ser Gly Tyr Ser Gly Ser Phe Val Gln His Pro Glu Leu Thr Gly Leu Asp Cys Ile Arg Pro Cys Phe Trp Val Glu Leu Val Arg Gly Leu Pro Arg Glu Asn Thr Thr Ile Trp Thr Ser Gly Ser Ser Ile Ser Phe Cys Gly Val Asn Ser Asp Thr Ala Asn Trp Ser Trp Pro Asp Gly Ala Glu Leu 450 455 Pro Phe Thr Ile Asp Lys 465 <210> SEO ID NO 35 <211> LENGTH: 716 <212> TYPE: PRT <213> ORGANISM: Influenza A virus <400> SEOUENCE: 35 Met Glu Asp Phe Val Arg Gln Cys Phe Asn Pro Met Ile Val Glu Leu Ala Glu Lys Ala Met Lys Glu Tyr Gly Glu Asp Leu Lys Ile Glu Thr 25 Asn Lys Phe Ala Ala Ile Cys Thr His Leu Glu Val Cys Phe Met Tyr Ser Asp Phe His Phe Ile Asn Glu Gln Gly Glu Ser Ile Val Val Glu Leu Asp Asp Pro Asn Ala Leu Leu Lys His Arg Phe Glu Ile Ile Glu Gly Arg Asp Arg Thr Met Ala Trp Thr Val Val Asn Ser Ile Cys Asn Thr Thr Gly Ala Gly Lys Pro Lys Phe Leu Pro Asp Leu Tyr Asp Tyr Lys Glu Asn Arg Phe Ile Glu Ile Gly Val Thr Arg Arg Glu Val His \$115\$ \$120\$ \$125\$Ile Tyr Tyr Leu Glu Lys Ala Asn Lys Ile Lys Ser Glu Asn Thr His Ile His Ile Phe Ser Phe Thr Gly Glu Glu Met Ala Thr Lys Ala Asp Tyr Thr Leu Asp Glu Glu Ser Arg Ala Arg Ile Lys Thr Arg Leu Phe Thr Ile Arg Gln Glu Met Ala Asn Arg Gly Leu Trp Asp Ser Phe Arg 185 Gln Ser Glu Arg Gly Glu Glu Thr Ile Glu Glu Lys Phe Glu Ile Thr Gly Thr Met Arg Arg Leu Ala Asp Gln Ser Leu Pro Pro Asn Phe Ser 215 Cys Leu Glu Asn Phe Arg Ala Tyr Val Asp Gly Phe Glu Pro Asn Gly

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Gly	Pro	Pro 275	Сув	Tyr	Gln	Arg	Ser 280	Lys	Phe	Leu	Leu	Met 285	Asp	Ala	Leu
ГÀз	Leu 290	Ser	Ile	Glu	Asp	Pro 295	Ser	His	Glu	Gly	Glu 300	Gly	Ile	Pro	Leu
Tyr 305	Asp	Ala	Ile	Lys	Cys 310	Met	Lys	Thr	Phe	Phe 315	Gly	Trp	Lys	Glu	Pro 320
Tyr	Ile	Val	Lys	Pro 325	His	Glu	Lys	Gly	Ile 330	Asn	Ser	Asn	Tyr	Leu 335	Leu
Ser	Trp	Lys	Gln 340	Val	Leu	Ser	Glu	Leu 345	Gln	Asp	Ile	Glu	Asn 350	Glu	Glu
Lys	Ile	Pro 355	Arg	Thr	Lys	Asn	Met 360	Lys	Lys	Thr	Ser	Gln 365	Leu	Lys	Trp
Ala	Leu 370	Gly	Glu	Asn	Met	Ala 375	Pro	Glu	Lys	Val	380 380	Phe	Glu	Asn	Cys
Arg 385	Asp	Ile	Ser	Asp	Leu 390	Lys	Gln	Tyr	Asp	Ser 395	Asp	Glu	Pro	Glu	Leu 400
Arg	Ser	Leu	Ser	Ser 405	Trp	Ile	Gln	Asn	Glu 410	Phe	Asn	ГÀа	Ala	Cys 415	Glu
Leu	Thr	Asp	Ser 420	Val	Trp	Ile	Glu	Leu 425	Asp	Glu	Ile	Gly	Glu 430	Asp	Val
Ala	Pro	Ile 435	Glu	His	Ile	Ala	Ser 440	Met	Arg	Arg	Asn	Tyr 445	Phe	Thr	Ala
Glu	Val 450	Ser	His	CAa	Arg	Ala 455	Thr	Glu	Tyr	Ile	Met 460	ГÀа	Gly	Val	Tyr
Ile 465	Asn	Thr	Ala	Leu	Leu 470	Asn	Ala	Ser	СЛа	Ala 475	Ala	Met	Asp	Asp	Phe 480
Gln	Leu	Ile	Pro	Met 485	Ile	Ser	Lys	Сув	Arg 490	Thr	ГÀа	Glu	Gly	Arg 495	Arg
ГÀа	Thr	Asn	Leu 500	Tyr	Gly	Phe	Ile	Ile 505	Lys	Gly	Arg	Ser	His 510	Leu	Arg
Asn	Asp	Thr 515	Asp	Val	Val	Asn	Phe 520	Val	Ser	Met	Glu	Phe 525	Ser	Leu	Thr
Asp	Pro 530	Arg	Leu	Glu	Pro	His 535	Lys	Trp	Glu	Lys	Tyr 540	CAa	Val	Leu	Glu
Ile 545	Gly	Asp	Met	Leu	Leu 550	Arg	Ser	Ala	Ile	Gly 555	Gln	Ile	Ser	Arg	Pro 560
Met	Phe	Leu	Tyr	Val 565	Arg	Thr	Asn	Gly	Thr 570	Ser	Lys	Val	Lys	Met 575	ГЛа
Trp	Gly	Met	Glu 580	Met	Arg	Arg	Cys	Leu 585	Leu	Gln	Ser	Leu	Gln 590	Gln	Ile
Glu	Ser	Met 595	Ile	Glu	Ala	Glu	Ser 600	Ser	Val	Lys	Glu	Lув 605	Asp	Met	Thr
Lys	Glu 610	Phe	Phe	Glu	Asn	Lys 615	Ser	Glu	Ala	Trp	Pro 620	Ile	Gly	Glu	Ser
Pro 625	Lys	Gly	Val	Glu	Glu 630	Gly	Ser	Ile	Gly	Lys 635	Val	СЛа	Arg	Thr	Leu 640
Leu	Ala	Lys	Ser	Val 645	Phe	Asn	Ser	Leu	Tyr 650	Ala	Ser	Pro	Gln	Leu 655	Glu

Gly Phe Ser Ala Glu Ser Arg Lys Leu Leu Leu Val Val Gln Ala Leu 665 Arg Asp Asn Leu Glu Pro Gly Thr Phe Asp Leu Gly Gly Leu Tyr Glu Ala Ile Glu Glu Cys Leu Ile As
n Asp Pro Trp Val Leu Leu As
n Ala $690 \hspace{1.5cm} 695 \hspace{1.5cm} 700 \hspace{1.5cm}$ Ser Trp Phe Asn Ser Phe Leu Thr His Ala Leu Lys <210> SEQ ID NO 36 <211> LENGTH: 757 <212> TYPE: PRT <213> ORGANISM: Influenza A virus <400> SEQUENCE: 36 Met Asp Val Asn Pro Thr Leu Leu Phe Leu Lys Val Pro Ala Gln Asn 1 $$ 5 $$ 10 $$ 15 Ala Ile Ser Thr Thr Phe Pro Tyr Thr Gly Asp Pro Pro Tyr Ser His $20 \hspace{1.5cm} 25 \hspace{1.5cm} 30 \hspace{1.5cm}$ Gly Thr Gly Thr Gly Tyr Thr Met Asp Thr Val Asn Arg Thr His Gln Tyr Ser Glu Lys Gly Lys Trp Thr Thr Asn Thr Glu Thr Gly Ala Pro 50 60 Gln Leu Asn Pro Ile Asp Gly Pro Leu Pro Glu Asp Asn Glu Pro Ser Gly Tyr Ala Gln Thr Asp Cys Val Leu Glu Ala Met Ala Phe Leu Glu Glu Ser His Pro Gly Ile Phe Glu Asn Ser Cys Leu Glu Thr Met Glu 105 Ala Val Gln Gln Thr Arg Val Asp Arg Leu Thr Gln Gly Arg Gln Thr 120 Tyr Asp Trp Thr Leu Asn Arg Asn Gln Pro Ala Ala Thr Ala Leu Ala Asn Thr Ile Glu Val Phe Arg Ser Asn Gly Leu Thr Ala Asn Glu Ser Gly Arg Leu Ile Asp Phe Leu Lys Asp Val Met Glu Ser Met Asp Lys 165 170 Glu Glu Met Glu Ile Thr Thr His Phe Gln Arg Lys Arg Arg Val Arg Asp Asn Met Thr Lys Lys Met Val Thr Gln Arg Thr Ile Gly Lys Lys Lys Gln Arg Val Asn Lys Arg Gly Tyr Leu Ile Arg Ala Leu Thr Leu Asn Thr Met Thr Lys Asp Ala Glu Arg Gly Lys Leu Lys Arg Arg Ala 225 230 235 240 Ile Ala Thr Pro Gly Met Gln Ile Arg Gly Phe Val Tyr Phe Val Glu 250 Thr Leu Ala Arg Ser Ile Cys Glu Lys Leu Glu Gln Ser Gly Leu Pro 265 Val Gly Gly Asn Glu Lys Lys Ala Lys Leu Ala Asn Val Val Arg Lys Met Met Thr Asn Ser Gln Asp Thr Glu Leu Ser Phe Thr Ile Thr Gly Asp Asn Thr Lys Trp Asn Glu Asn Gln Asn Pro Arg Met Phe Leu Ala

305					310					315					320
Met	Ile	Thr	Tyr	Ile 325	Thr	Lys	Asn	Gln	Pro 330	Glu	Trp	Phe	Arg	Asn 335	Ile
Leu	Ser	Ile	Ala 340	Pro	Ile	Met	Phe	Ser 345	Asn	Lys	Met	Ala	Arg 350	Leu	Gly
Lys	Gly	Tyr 355	Met	Phe	Glu	Ser	160 160	Arg	Met	Lys	Leu	Arg 365	Thr	Gln	Ile
Pro	Ala 370	Glu	Met	Leu	Ala	Ser 375	Ile	Asp	Leu	Lys	Tyr 380	Phe	Asn	Glu	Ser
Thr 385	Arg	Lys	Lys	Ile	Glu 390	Lys	Ile	Arg	Pro	Leu 395	Leu	Ile	Asp	Gly	Thr 400
Ala	Ser	Leu	Ser	Pro 405	Gly	Met	Met	Met	Gly 410	Met	Phe	Asn	Met	Leu 415	Ser
Thr	Val	Leu	Gly 420	Val	Ser	Ile	Leu	Asn 425	Leu	Gly	Gln	Lys	Lys 430	Tyr	Thr
Lys	Thr	Thr 435	Tyr	Trp	Trp	Asp	Gly 440	Leu	Gln	Ser	Ser	Asp 445	Asp	Phe	Ala
Leu	Ile 450	Val	Asn	Ala	Pro	Asn 455	His	Glu	Gly	Ile	Gln 460	Ala	Gly	Val	Asn
Arg 465	Phe	Tyr	Arg	Thr	Cys 470	Lys	Leu	Val	Gly	Ile 475	Asn	Met	Ser	Lys	Lys 480
Lys	Ser	Tyr	Ile	Asn 485	Lys	Thr	Gly	Thr	Phe 490	Glu	Phe	Thr	Ser	Phe 495	Phe
Tyr	Arg	Tyr	Gly 500	Phe	Val	Ala	Asn	Phe 505	Ser	Met	Glu	Leu	Pro 510	Ser	Phe
Gly	Val	Ser 515	Gly	Ile	Asn	Glu	Ser 520	Ala	Asp	Met	Ser	Ile 525	Gly	Val	Thr
Val	Ile 530	Lys	Asn	Asn	Met	Ile 535	Asn	Asn	Asp	Leu	Gly 540	Pro	Ala	Thr	Ala
Gln 545	Met	Ala	Leu	Gln	Leu 550	Phe	Ile	Lys	Asp	Tyr 555	Arg	Tyr	Thr	Tyr	Arg 560
Cys	His	Arg	Gly	Asp 565	Thr	Gln	Ile	Gln	Thr 570	Arg	Arg	Ser	Phe	Glu 575	Leu
Lys	Lys	Leu	Trp 580	Asp	Gln	Thr	Gln	Ser 585	Arg	Ala	Gly	Leu	Leu 590	Val	Ser
Asp	Gly	Gly 595	Pro	Asn	Leu	Tyr	Asn 600	Ile	Arg	Asn	Leu	His 605	Ile	Pro	Glu
Val	Cys	Leu	ГЛа	Trp	Glu	Leu 615	Met	Asp	Glu	Asn	Tyr 620	Arg	Gly	Arg	Leu
Сув 625	Asn	Pro	Leu	Asn	Pro 630	Phe	Val	Ser	His	Lys 635	Glu	Ile	Glu	Ser	Val 640
Asn	Asn	Ala	Val	Val 645	Met	Pro	Ala	His	Gly 650	Pro	Ala	Lys	Ser	Met 655	Glu
Tyr	Asp	Ala	Val 660	Ala	Thr	Thr	His	Ser 665	Trp	Ile	Pro	Lys	Arg 670	Asn	Arg
Ser	Ile	Leu 675	Asn	Thr	Ser	Gln	Arg 680	Gly	Ile	Leu	Glu	Asp 685	Glu	Gln	Met
Tyr	Gln 690	Lys	Cys	Сув	Asn	Leu 695	Phe	Glu	Lys	Phe	Phe 700	Pro	Ser	Ser	Ser
Tyr 705	Arg	Arg	Pro	Ile	Gly 710	Ile	Ser	Ser	Met	Val 715	Glu	Ala	Met	Val	Ser 720
Arg	Ala	Arg	Ile	Asp 725	Ala	Arg	Ile	Asp	Phe 730	Glu	Ser	Gly	Arg	Ile 735	Lys

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Lys Glu Glu Phe Ser Glu Ile Met Lys Ile Cys Ser Thr Ile Glu Glu Leu Arg Arg Gln Arg 755 <210> SEQ ID NO 37 <211> LENGTH: 759 <212> TYPE: PRT <213> ORGANISM: Influenza A virus <400> SEQUENCE: 37 Met Glu Arg Ile Lys Glu Leu Arg Asn Leu Met Ser Gln Ser Arg Thr Arg Glu Ile Leu Thr Lys Thr Thr Val Asp His Met Ala Ile Ile Lys Lys Tyr Thr Ser Gly Arg Gln Glu Lys Asn Pro Ser Leu Arg Met Lys Trp Met Met Ala Met Lys Tyr Pro Ile Thr Ala Asp Lys Arg Ile Thr 50 $\,$ 55 $\,$ 60 Glu Met Val Pro Glu Arg Asn Glu Gln Gly Gln Thr Leu Trp Ser Lys 65 70 75 80 Met Ser Asp Ala Gly Ser Asp Arg Val Met Val Ser Pro Leu Ala Val 90 Thr Trp Trp Asn Arg Asn Gly Pro Val Thr Ser Thr Val His Tyr Pro 100 105 Lys Val Tyr Lys Thr Tyr Phe Asp Lys Val Glu Arg Leu Lys His Gly Thr Phe Gly Pro Val His Phe Arg Asn Gln Val Lys Ile Arg Arg 135 Val Asp Ile Asn Pro Gly His Ala Asp Leu Ser Ala Lys Glu Ala Gln 155 Asp Val Ile Met Glu Val Val Phe Pro Asn Glu Val Gly Ala Arg Ile Leu Thr Ser Glu Ser Gln Leu Thr Ile Thr Lys Glu Lys Lys Glu Glu Leu Arg Asp Cys Lys Ile Ser Pro Leu Met Val Ala Tyr Met Leu Glu 200 Arg Glu Leu Val Arg Lys Thr Arg Phe Leu Pro Val Ala Gly Gly Thr Ser Ser Ile Tyr Ile Glu Val Leu His Leu Thr Gln Gly Thr Cys Trp Glu Gln Met Tyr Thr Pro Gly Gly Glu Val Arg Asn Asp Asp Val Asp Gln Ser Leu Ile Ile Ala Ala Arg Asn Ile Val Arg Arg Ala Ala Val Ser Ala Asp Pro Leu Ala Ser Leu Leu Glu Met Cys His Ser Thr Gln 280 Ile Gly Gly Thr Arg Met Val Asp Ile Leu Arg Gln Asn Pro Thr Glu 295 Glu Gln Ala Val Asp Ile Cys Lys Ala Ala Met Gly Leu Arg Ile Ser Ser Ser Phe Ser Phe Gly Gly Phe Thr Phe Lys Arg Thr Ser Gly Ser

Ser Val Lys Lys Glu Glu Glu Val Leu Thr Gly Asn Leu Gln Thr Leu

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			340					345					350		
ГÀа	Ile	Arg 355	Val	His	Glu	Gly	Tyr 360	Glu	Glu	Phe	Thr	Met 365	Val	Gly	Lys
Arg	Ala 370	Thr	Ala	Ile	Leu	Arg 375	ГÀз	Ala	Thr	Arg	Arg 380	Leu	Val	Gln	Leu
Ile 385	Val	Ser	Gly	Arg	390	Glu	Gln	Ser	Ile	Ala 395	Glu	Ala	Ile	Ile	Val 400
Ala	Met	Val	Phe	Ser 405	Gln	Glu	Asp	Cys	Met 410	Ile	Lys	Ala	Val	Arg 415	Gly
Asp	Leu	Asn	Phe 420	Val	Asn	Arg	Ala	Asn 425	Gln	Arg	Leu	Asn	Pro 430	Met	His
Gln	Leu	Leu 435	Arg	His	Phe	Gln	Lys 440	Asp	Ala	Lys	Val	Leu 445	Phe	Gln	Asn
Trp	Gly 450	Ile	Glu	His	Ile	Asp 455	Ser	Val	Met	Gly	Met 460	Val	Gly	Val	Leu
Pro 465	Asp	Met	Thr	Pro	Ser 470	Thr	Glu	Met	Ser	Met 475	Arg	Gly	Ile	Arg	Val 480
Ser	Lys	Met	Gly	Val 485	Asp	Glu	Tyr	Ser	Ser 490	Thr	Glu	Arg	Val	Val 495	Val
Ser	Ile	Asp	Arg 500	Phe	Leu	Arg	Val	Arg 505	Asp	Gln	Arg	Gly	Asn 510	Val	Leu
Leu	Ser	Pro 515	Glu	Glu	Val	Ser	Glu 520	Thr	Gln	Gly	Thr	Glu 525	Arg	Leu	Thr
Ile	Thr 530	Tyr	Ser	Ser	Ser	Met 535	Met	Trp	Glu	Ile	Asn 540	Gly	Pro	Glu	Ser
Val 545	Leu	Val	Asn	Thr	Tyr 550	Gln	Trp	Ile	Ile	Arg 555	Asn	Trp	Glu	Ala	Val 560
Lys	Ile	Gln	Trp	Ser 565	Gln	Asn	Pro	Ala	Met 570	Leu	Tyr	Asn	ГÀз	Met 575	Glu
Phe	Glu	Pro	Phe 580	Gln	Ser	Leu	Val	Pro 585	ГÀа	Ala	Ile	Arg	Ser 590	Gln	Tyr
Ser	Gly	Phe 595	Val	Arg	Thr	Leu	Phe 600	Gln	Gln	Met	Arg	Asp 605	Val	Leu	Gly
Thr	Phe 610	Asp	Thr	Thr	Gln	Ile 615	Ile	Lys	Leu	Leu	Pro 620	Phe	Ala	Ala	Ala
Pro 625	Pro	ГÀз	Gln	Ser	Arg 630	Met	Gln	Phe	Ser	Ser 635	Leu	Thr	Val	Asn	Val 640
Arg	Gly	Ser	Gly	Met 645	Arg	Ile	Leu	Val	Arg 650	Gly	Asn	Ser	Pro	Val 655	Phe
Asn	Tyr	Asn	Lys	Thr	Thr	Lys	Arg	Leu 665	Thr	Ile	Leu	Gly	Lys 670	Asp	Ala
Gly	Thr	Leu 675	Ile	Glu	Asp	Pro	Asp 680	Glu	Ser	Thr	Ser	Gly 685	Val	Glu	Ser
Ala	Val 690	Leu	Arg	Gly	Phe	Leu 695	Ile	Ile	Gly	Lys	Glu 700	Asp	Arg	Arg	Tyr
Gly 705	Pro	Ala	Leu	Ser	Ile 710	Asn	Glu	Leu	Ser	Asn 715	Leu	Ala	Lys	Gly	Glu 720
Lys	Ala	Asn	Val	Leu 725	Ile	Gly	Gln	Gly	Asp 730	Val	Val	Leu	Val	Met 735	Lys
Arg	Lys	Arg	Asp 740	Ser	Ser	Ile	Leu	Thr 745	Asp	Ser	Gln	Thr	Ala 750	Thr	Lys
Arg	Ile	Arg	Met	Ala	Ile	Asn									

Arg Ile Arg Met Ala Ile Asn 755

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Ile	Asp	Gly 35	Ile	Gly	Arg	Phe	Tyr 40	Ile	Gln	Met	CAa	Thr 45	Glu	Leu	Lys
Leu	Ser 50	Asp	Tyr	Glu	Gly	Arg 55	Leu	Ile	Gln	Asn	Ser 60	Leu	Thr	Ile	Glu
Lys 65	Met	Val	Leu	Ser	Ala 70	Phe	Asp	Glu	Arg	Arg 75	Asn	Lys	Tyr	Leu	Glu 80
Glu	His	Pro	Ser	Ala 85	Gly	Lys	Asp	Pro	Lys	Lys	Thr	Gly	Gly	Pro 95	Ile
Tyr	Arg	Arg	Val 100	Asp	Gly	Lys	Trp	Met 105	Arg	Glu	Leu	Val	Leu 110	Tyr	Asp
ГЛа	Glu	Glu 115	Ile	Arg	Arg	Ile	Trp 120	Arg	Gln	Ala	Asn	Asn 125	Gly	Glu	Asp
Ala	Thr 130	Ala	Gly	Leu	Thr	His 135	Ile	Met	Ile	Trp	His 140	Ser	Asn	Leu	Asn
Asp 145	Ala	Thr	Tyr	Gln	Arg 150	Thr	Arg	Ala	Leu	Val 155	Arg	Thr	Gly	Met	Asp 160
Pro	Arg	Met	Cys	Ser 165	Leu	Met	Gln	Gly	Ser 170	Thr	Leu	Pro	Arg	Arg 175	Ser
Gly	Ala	Ala	Gly 180	Ala	Ala	Val	Lys	Gly 185	Ile	Gly	Thr	Met	Val 190	Met	Glu
Leu	Ile	Arg 195	Met	Val	Lys	Arg	Gly 200	Ile	Asn	Asp	Arg	Asn 205	Phe	Trp	Arg
Gly	Glu 210	Asn	Gly	Arg	Lys	Thr 215	Arg	Ser	Ala	Tyr	Glu 220	Arg	Met	Cys	Asn
Ile 225	Leu	Lys	Gly	Lys	Phe 230	Gln	Thr	Ala	Ala	Gln 235	Arg	Ala	Met	Val	Asp 240
Gln	Val	Arg	Glu	Ser 245	Arg	Asn	Pro	Gly	Asn 250	Ala	Glu	Ile	Glu	Asp 255	Leu
Ile	Phe	Leu	Ala 260	Arg	Ser	Ala	Leu	Ile 265	Leu	Arg	Gly	Ser	Val 270	Ala	His
Lys	Ser	Cys 275	Leu	Pro	Ala	Cys	Val 280	Tyr	Gly	Pro	Ala	Val 285	Ser	Ser	Gly
Tyr	Asn 290	Phe	Glu	Lys	Glu	Gly 295	Tyr	Ser	Leu	Val	Gly 300	Ile	Asp	Pro	Phe
Lys 305	Leu	Leu	Gln	Asn	Ser 310	Gln	Val	Tyr	Ser	Leu 315	Ile	Arg	Pro	Asn	Glu 320
Asn	Pro	Ala	His	Lys 325	Ser	Gln	Leu	Val	Trp 330	Met	Ala	Сув	His	Ser 335	Ala
Ala	Phe	Glu	Asp 340	Leu	Arg	Leu	Leu	Ser 345	Phe	Ile	Arg	Gly	Thr 350	Lys	Val
Ser	Pro	Arg 355	Gly	ГЛа	Leu	Ser	Thr 360	Arg	Gly	Val	Gln	Ile 365	Ala	Ser	Asn
Glu	Asn	Met	Asp	Asn	Met	Gly	Ser	Gly	Thr	Leu	Glu	Leu	Arg	Ser	Gly

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	370					375					380				
Tyr 385	Trp	Ala	Ile	Arg	Thr 390	Arg	Ser	Gly	Gly	Asn 395	Thr	Asn	Gln	Gln	Arg 400
Ala	Ser	Ala	Gly	Gln 405	Thr	Ser	Val	Gln	Pro 410	Thr	Phe	Ser	Val	Gln 415	Arg
Asn	Leu	Pro	Phe 420	Glu	Lys	Ser	Thr	Ile 425	Met	Ala	Ala	Phe	Thr 430	Gly	Asn
Thr	Glu	Gly 435	Arg	Thr	Ser	Asp	Met 440	Arg	Ala	Glu	Ile	Ile 445	Arg	Met	Met
Glu	Gly 450	Ala	Lys	Pro	Glu	Glu 455	Val	Ser	Phe	Arg	Gly 460	Arg	Gly	Val	Phe
Glu 465	Leu	Ser	Asp	Glu	Lys 470	Ala	Thr	Asn	Pro	Ile 475	Val	Pro	Ser	Phe	Asp 480
Met	Ser	Asn	Glu	Gly 485	Ser	Tyr	Phe	Phe	Gly 490	Asp	Asn	Ala	Glu	Glu 495	Tyr
Asp	Asn														
			ои о												
<212	2 > T	YPE:													
				Inf:	Luenz	za A	viru	ıs							
			ICE :												
Met 1	Ser	Leu	Leu	Thr 5	Glu	Val	Glu	Thr	Tyr 10	Val	Leu	Ser	Ile	Val 15	Pro
Ser	Gly	Pro	Leu 20	ГÀз	Ala	Glu	Ile	Ala 25	Gln	Arg	Leu	Glu	30 Asp	Val	Phe
Ala	Gly	Lys 35	Asn	Thr	Asp	Leu	Glu 40	Ala	Leu	Met	Glu	Trp 45	Leu	Lys	Thr
Arg	Pro 50	Ile	Leu	Ser	Pro	Leu 55	Thr	ГÀв	Gly	Ile	Leu 60	Gly	Phe	Val	Phe
Thr 65	Leu	Thr	Val	Pro	Ser 70	Glu	Arg	Gly	Leu	Gln 75	Arg	Arg	Arg	Phe	Val 80
Gln	Asn	Ala	Leu	Asn 85	Gly	Asn	Gly	Asp	Pro 90	Asn	Asn	Met	Asp	Lys 95	Ala
Val	Lys	Leu	Tyr 100	Arg	ГÀа	Leu	ГÀа	Arg 105	Glu	Ile	Thr	Phe	His 110	Gly	Ala
ГÀа	Glu	Ile 115	Ala	Leu	Ser	Tyr	Ser 120	Ala	Gly	Ala	Leu	Ala 125	Ser	Cys	Met
Gly	Leu 130	Ile	Tyr	Asn	Arg	Met 135	Gly	Ala	Val	Thr	Thr 140	Glu	Val	Ala	Phe
Gly 145	Leu	Val	CÀa	Ala	Thr 150	CÀa	Glu	Gln	Ile	Ala 155	Asp	Ser	Gln	His	Arg 160
Ser	His	Arg	Gln	Met 165	Val	Ala	Thr	Thr	Asn 170	Pro	Leu	Ile	Arg	His 175	Glu
Asn	Arg	Met	Val 180	Leu	Ala	Ser	Thr	Thr 185	Ala	Lys	Ala	Met	Glu 190	Gln	Met
Ala	Gly	Ser 195	Ser	Glu	Gln	Ala	Ala 200	Glu	Ala	Met	Glu	Ile 205	Ala	Ser	Gln
Ala	Arg 210	Gln	Met	Val	Gln	Ala 215	Met	Arg	Ala	Ile	Gly 220	Thr	His	Pro	Ser
Ser 225	Ser	Thr	Gly	Leu	Arg 230	Asp	Asp	Leu	Leu	Glu 235	Asn	Leu	Gln	Thr	Tyr 240
Gln	Lys	Arg	Met	Gly	Val	Gln	Met	Gln	Arg	Phe	Lys				

157 158

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245 250 <210> SEQ ID NO 40 <211> LENGTH: 97 <212> TYPE: PRT <213> ORGANISM: Influenza A virus <400> SEQUENCE: 40 Met Ser Leu Leu Thr Glu Val Glu Thr Pro Ile Arg Asn Glu Trp Gly Cys Arg Cys Asn Asp Ser Ser Asp Pro Leu Val Val Ala Ala Asn Ile Ile Gly Ile Leu His Leu Ile Leu Trp Ile Leu Asp Arg Leu Phe Phe Lys Cys Val Tyr Arg Leu Phe Lys His Gly Leu Lys Arg Gly Pro Ser Thr Glu Gly Val Pro Glu Ser Met Arg Glu Glu Tyr Arg Lys Glu Gln Gln Asn Ala Val Asp Ala Asp Asp Ser His Phe Val Ser Ile Glu Leu 90 Glu <210> SEO ID NO 41 <211> LENGTH: 846 <212> TYPE: DNA <213> ORGANISM: Influenza A virus <400> SEQUENCE: 41 aatggattcc aacactgtgt caagtttcca ggtagattgc tttctttggc atatccggaa acaagttgta gaccaagaac tgagtgatgc cccattcctt gatcggcttc gccgagatca 120 gaggtcccta aggggaagag gcaatactct cggtctagac atcaaagcag ccacccatgt 180 tggaaagcaa attgtagaaa agattctgaa agaagaatct gatgaggcac ttaaaatgac 240 catggtctcc acacctgctt cgcgatacat aactgacatg actattgagg aattgtcaag 300 aaactggttc atgctaatgc ccaagcagaa agtggaagga cctctttgca tcagaatgga 360 ccaggcaatc atggagaaaa acatcatgtt gaaagcgaat ttcagtgtga tttctgaccg 420 actagagacc atagtattac taagggcttt caccgaagag ggagcaattg ttggcgaaat 480 ctcaccattg ccttcttttc caggacatac tattgaggat gtcaaaaatg caattggggt 540 cctcatcgga ggacttgaat ggaatgataa cacagttcga gtctctaaaa atctacagag attegettgg agaageagta atgagaatgg gggaeeteea ettaeteeaa aacagaaacg gaaaatggcg agaacagcta ggtcaaaagt ttgaagagat aagatggctg attgaagaag tgagacacag actaaaaaca actgaaaata gctttgaaca aataacattc atgcaagcat 780 tacaactqct qtttqaaqtq qaacaqqaqa taaqaacttt ctcatttcaq cttatttaat 840 qataaa 846 <210> SEQ ID NO 42 <211> LENGTH: 566 <212> TYPE: PRT <213> ORGANISM: Influenza A virus

<400> SEQUENCE: 42

Met Lys Thr Ile Ile Ala Leu Ser Tyr Ile Leu Cys Leu Val Phe Ala 10

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His	His	Ala 35	Val	Pro	Asn	Gly	Thr 40	Ile	Val	Lys	Thr	Ile 45	Thr	Asn	Asp
Gln	Ile 50	Glu	Val	Thr	Asn	Ala 55	Thr	Glu	Leu	Val	Gln 60	Ser	Ser	Ser	Thr
Gly 65	Gly	Ile	Сув	Asp	Ser 70	Pro	His	Gln	Ile	Leu 75	Asp	Gly	Glu	Asn	Cys
Thr	Leu	Ile	Asp	Ala 85	Leu	Leu	Gly	Asp	Pro 90	Gln	СЛа	Asp	Gly	Phe 95	Gln
Asn	Lys	ГЛа	Trp 100	Asp	Leu	Phe	Val	Glu 105	Arg	Ser	ГÀз	Ala	Tyr 110	Ser	Asn
Cys	Tyr	Pro 115	Tyr	Asp	Val	Pro	Asp 120	Tyr	Ala	Ser	Leu	Arg 125	Ser	Leu	Val
Ala	Ser 130	Ser	Gly	Thr	Leu	Glu 135	Phe	Asn	Asp	Glu	Ser 140	Phe	Asn	Trp	Thr
Gly 145	Val	Thr	Gln	Asn	Gly 150	Thr	Ser	Ser	Ser	Сув 155	Lys	Arg	Arg	Ser	Asn 160
Asn	Ser	Phe	Phe	Ser 165	Arg	Leu	Asn	Trp	Leu 170	Thr	His	Leu	Lys	Phe 175	Lys
Tyr	Pro	Ala	Leu 180	Asn	Val	Thr	Met	Pro 185	Asn	Asn	Glu	ГÀа	Phe 190	Asp	Lys
Leu	Tyr	Ile 195	Trp	Gly	Val	His	His 200	Pro	Val	Thr	Asp	Asn 205	Asp	Gln	Ile
Phe	Leu 210	Tyr	Ala	Gln	Ala	Ser 215	Gly	Arg	Ile	Thr	Val 220	Ser	Thr	Lys	Arg
Ser 225	Gln	Gln	Thr	Val	Ile 230	Pro	Asn	Ile	Gly	Ser 235	Arg	Pro	Arg	Ile	Arg 240
Asn	Ile	Pro	Ser	Arg 245	Ile	Ser	Ile	Tyr	Trp 250	Thr	Ile	Val	Lys	Pro 255	Gly
Asp	Ile	Leu	Leu 260	Ile	Asn	Ser	Thr	Gly 265	Asn	Leu	Ile	Ala	Pro 270	Arg	Gly
Tyr	Phe	Lys 275	Ile	Arg	Ser	Gly	Lys 280	Ser	Ser	Ile	Met	Arg 285	Ser	Asp	Ala
Pro	Ile 290	Gly	Lys	САв	Asn	Ser 295	Glu	Cys	Ile	Thr	Pro 300	Asn	Gly	Ser	Ile
Pro 305	Asn	Asp	ГÀа	Pro	Phe 310	Gln	Asn	Val	Asn	Arg 315	Ile	Thr	Tyr	Gly	Ala 320
Cys	Pro	Arg	Tyr	Val 325	Lys	Gln	Asn	Thr	Leu 330	Lys	Leu	Ala	Thr	Gly 335	Met
Arg	Asn	Val	Pro 340	Glu	ГÀа	Gln	Thr	Arg 345	Gly	Ile	Phe	Gly	Ala 350	Ile	Ala
Gly	Phe	Ile 355	Glu	Asn	Gly	Trp	Glu 360	Gly	Met	Val	Asp	Gly 365	Trp	Tyr	Gly
Phe	Arg 370	His	Gln	Asn	Ser	Glu 375	Gly	Ile	Gly	Gln	Ala 380	Ala	Asp	Leu	Lys
Ser 385	Thr	Gln	Ala	Ala	Ile 390	Asn	Gln	Ile	Asn	Gly 395	Lys	Leu	Asn	Arg	Leu 400
Ile	Gly	Lys	Thr	Asn 405	Glu	Lys	Phe	His	Gln 410	Ile	Glu	ГÀв	Glu	Phe 415	Ser
Glu	Val	Glu	Gly 420	Arg	Ile	Gln	Asp	Leu 425	Glu	Lys	Tyr	Val	Glu 430	Asp	Thr
Lys	Ile	Asp	Leu	Trp	Ser	Tyr	Asn	Ala	Glu	Leu	Leu	Val	Ala	Leu	Glu

		435					440					445			
Asn	Gln 450	His	Thr	Ile	Asp	Leu 455	Thr	Asp	Ser	Glu	Met 460	Asn	Lys	Leu	Phe
Glu 465	Arg	Thr	Lys	Lys	Gln 470	Leu	Arg	Glu	Asn	Ala 475	Glu	Asp	Met	Gly	Asn 480
Gly	Сла	Phe	Lys	Ile 485	Tyr	His	Lys	СЛа	Asp 490	Asn	Ala	СЛа	Ile	Gly 495	Ser
Ile	Arg	Asn	Gly 500	Thr	Tyr	Asp	His	Asp 505	Val	Tyr	Arg	Asp	Glu 510	Ala	Leu
Asn	Asn	Arg 515	Phe	Gln	Ile	Lys	Gly 520	Val	Glu	Leu	Lys	Ser 525	Gly	Tyr	Lys
Asp	Trp 530	Ile	Leu	Trp	Ile	Ser 535	Phe	Ala	Ile	Ser	Cys 540	Phe	Leu	Leu	Cys
Val 545	Ala	Leu	Leu	Gly	Phe 550	Ile	Met	Trp	Ala	Сув 555	Gln	ГÀа	Gly	Asn	Ile 560
Arg	Сув	Asn	Ile	Сув 565	Ile										
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Met 1	Asn	Pro	Asn	Gln 5	ГÀа	Ile	Ile	Thr	Ile 10	Gly	Ser	Val	Ser	Leu 15	Thr
Ile	Ser	Thr	Ile 20	CAa	Phe	Phe	Met	Gln 25	Ile	Ala	Ile	Leu	Ile 30	Thr	Thr
Val	Thr	Leu 35	His	Phe	Lys	Gln	Tyr 40	Glu	Phe	Asn	Ser	Pro 45	Pro	Asn	Asn
Gln	Val 50	Met	Leu	Сув	Glu	Pro 55	Thr	Ile	Ile	Glu	Arg 60	Asn	Ile	Thr	Glu
Ile 65	Val	Tyr	Leu	Thr	Asn 70	Thr	Thr	Ile	Glu	Lуз 75	Glu	Ile	Сув	Pro	80 FÀa
Leu	Ala	Glu	Tyr	Arg 85	Asn	Trp	Ser	Lys	Pro 90	Gln	CÀa	Asn	Ile	Thr 95	Gly
Phe	Ala	Pro	Phe 100	Ser	ГÀа	Asp	Asn	Ser 105	Ile	Arg	Leu	Ser	Ala 110	Gly	Gly
Asp	Ile	Trp 115	Val	Thr	Arg	Glu	Pro 120	Tyr	Val	Ser	CÀa	Asp 125	Pro	Asp	Lys
	130					Gly 135		-			140				
Ser 145	Asn	Asp	Thr	Val	His 150	Asp	Arg	Thr	Pro	Tyr 155	Arg	Thr	Leu	Leu	Met 160
Asn	Glu	Leu	Gly	Val 165	Pro	Phe	His	Leu	Gly 170	Thr	ГÀв	Gln	Val	Суs 175	Ile
Ala	Trp	Ser	Ser 180	Ser	Ser	CÀa	His	Asp 185	Gly	Lys	Ala	Trp	Leu 190	His	Val
CÀa	Val	Thr 195	Gly	Asp	Asp	ГЛа	Asn 200	Ala	Thr	Ala	Ser	Phe 205	Ile	Tyr	Asn
Gly	Arg 210	Leu	Val	Asp	Ser	Ile 215	Val	Ser	Trp	Ser	Lys 220	Glu	Ile	Leu	Arg
Thr 225	Gln	Glu	Ser	Glu	Сув 230	Val	СЛа	Ile	Asn	Gly 235	Thr	CAa	Thr	Val	Val 240

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Met Thr Asp Gly Ser Ala Ser Gly Lys Ala Asp Thr Lys Ile Leu Phe 250 Ile Glu Glu Gly Lys Ile Val His Thr Ser Thr Leu Ser Gly Ser Ala Gln His Val Glu Glu Cys Ser Cys Tyr Pro Arg Tyr Leu Gly Val Arg Cys Val Cys Arg Asp Asn Trp Lys Gly Ser Asn Arg Pro Ile Val Asp Ile Asn Ile Lys Asp Tyr Ser Ile Val Ser Ser Tyr Val Cys Ser Gly 305 310 315 320 Leu Val Gly Asp Thr Pro Arg Lys Asn Asp Ser Ser Ser Ser Ser His Cys Leu Asp Pro Asn Asn Glu Glu Gly Gly His Gly Val Lys Gly Trp Ala Phe Asp Asp Gly Asn Asp Val Trp Met Gly Arg Thr Ile Ser Glu Lys Leu Arg Ser Gly Tyr Glu Thr Phe Lys Val Ile Glu Gly Trp Ser Asn Pro Asn Ser Lys Leu Gln Ile Asn Arg Gln Val Ile Val Asp Arg Gly Asn Arg Ser Gly Tyr Ser Gly Ile Phe Ser Val Glu Gly Lys Ser 410 Cys Ile Asn Arg Cys Phe Tyr Val Glu Leu Ile Arg Gly Arg Lys Glu 425 Glu Thr Glu Val Leu Trp Thr Ser Asn Ser Ile Val Val Phe Cys Gly 440 Thr Ser Gly Thr Tyr Gly Thr Gly Ser Trp Pro Asp Gly Ala Asp Ile 455 Asn Leu Met Pro Ile <210> SEQ ID NO 44 <211> LENGTH: 716 <212> TYPE: PRT <213> ORGANISM: Influenza A virus <400> SEQUENCE: 44 Met Glu Asp Phe Val Arg Gln Cys Phe Asn Pro Met Ile Val Glu Leu Ala Glu Lys Ala Met Lys Glu Tyr Gly Glu Asp Pro Lys Ile Glu Thr Asn Lys Phe Ala Ala Ile Cys Thr His Leu Glu Val Cys Phe Met Tyr Ser Asp Phe His Phe Ile Asp Glu Arg Gly Glu Ser Ile Ile Val Glu Ser Gly Asp Pro Asn Ala Leu Leu Lys His Arg Phe Glu Ile Ile Glu Gly Arg Asp Arg Ile Met Ala Trp Thr Val Ile Asn Ser Ile Cys Asn Thr Thr Gly Val Glu Lys Pro Lys Phe Leu Pro Asp Leu Tyr Asp Tyr 105 Lys Glu Asn Arg Phe Ile Glu Ile Gly Val Thr Arg Arg Glu Val His 120 Ile Tyr Tyr Leu Glu Lys Ala Asn Lys Ile Lys Ser Glu Lys Thr His 135

Ile 145	His	Ile	Phe	Ser	Phe 150	Thr	Gly	Glu	Glu	Met 155	Ala	Thr	Lys	Ala	Asp 160
Tyr	Thr	Leu	Asp	Glu 165	Glu	Ser	Arg	Ala	Arg 170	Ile	Lys	Thr	Arg	Leu 175	Phe
Thr	Ile	Arg	Gln 180	Glu	Met	Ala	Ser	Lys 185	Ser	Leu	Trp	Asp	Ser 190	Phe	Arg
Gln	Ser	Glu 195	Arg	Gly	Glu	Glu	Thr 200	Ile	Glu	Glu	Lys	Phe 205	Glu	Ile	Thr
Gly	Thr 210	Met	Arg	Lys	Leu	Ala 215	Asp	Gln	Ser	Leu	Pro 220	Pro	Asn	Phe	Pro
Ser 225	Leu	Glu	Asn	Phe	Arg 230	Ala	Tyr	Val	Asp	Gly 235	Phe	Glu	Pro	Asn	Gly 240
CAa	Ile	Glu	Gly	Lys 245	Leu	Ser	Gln	Met	Ser 250	Lys	Glu	Val	Asn	Ala 255	Lys
Ile	Glu	Pro	Phe 260	Leu	Arg	Thr	Thr	Pro 265	Arg	Pro	Leu	Arg	Leu 270	Pro	Asp
Gly	Pro	Leu 275	Cys	His	Gln	Arg	Ser 280	Lys	Phe	Leu	Leu	Met 285	Asp	Ala	Leu
Lys	Leu 290	Ser	Ile	Glu	Asp	Pro 295	Ser	His	Glu	Gly	Glu 300	Gly	Ile	Pro	Leu
Tyr 305	Asp	Ala	Ile	Lys	310	Met	Lys	Thr	Phe	Phe 315	Gly	Trp	Lys	Glu	Pro 320
Asn	Ile	Val	Lys	Pro 325	His	Glu	Lys	Gly	Ile 330	Asn	Pro	Asn	Tyr	Leu 335	Met
Ala	Trp	Lys	Gln 340	Val	Leu	Ala	Glu	Leu 345	Gln	Asp	Ile	Glu	Asn 350	Glu	Glu
ГÀа	Ile	Pro 355	Arg	Thr	Lys	Asn	Met 360	Lys	Arg	Thr	Ser	Gln 365	Leu	Lys	Trp
Ala	Leu 370	Gly	Glu	Asn	Met	Ala 375	Pro	Glu	Lys	Val	380	Phe	Asp	Asp	CAa
385 Lys	Asp	Val	Gly	Asp	Leu 390	Lys	Gln	Tyr	Asp	Ser 395	Asp	Glu	Pro	Glu	Pro 400
Arg	Ser	Leu	Ala	Ser 405	Trp	Val	Gln	Asn	Glu 410	Phe	Asn	ГÀа	Ala	Cys 415	Glu
Leu	Thr	Asp	Ser 420	Ser	Trp	Ile	Glu	Leu 425	Asp	Glu	Ile	Gly	Glu 430	Aap	Val
Ala	Pro	Ile 435	Glu	His	Ile	Ala	Ser 440		Arg	Arg	Asn	Tyr 445	Phe	Thr	Ala
Glu	Val 450	Ser	His	CAa	Arg	Ala 455	Thr	Glu	Tyr	Ile	Met 460	Lys	Gly	Val	Tyr
Ile 465	Asn	Thr	Ala	Leu	Leu 470	Asn	Ala	Ser	Cys	Ala 475	Ala	Met	Asp	Asp	Phe 480
Gln	Leu	Ile	Pro	Met 485	Ile	Ser	Lys	Cys	Arg 490	Thr	Lys	Glu	Gly	Arg 495	Arg
Lys	Thr	Asn	Leu 500	Tyr	Gly	Phe	Ile	Ile 505	Lys	Gly	Arg	Ser	His 510	Leu	Arg
Asn	Asp	Thr 515	Asp	Val	Val	Asn	Phe 520	Val	Ser	Met	Glu	Phe 525	Ser	Leu	Thr
Asp	Pro 530	Arg	Leu	Glu	Pro	His 535	Lys	Trp	Glu	Lys	Tyr 540	СЛа	Val	Leu	Glu
Ile 545	Gly	Asp	Met	Leu	Leu 550	Arg	Thr	Ala	Ile	Gly 555	Gln	Val	Ser	Arg	Pro 560

Met Phe Leu Tyr Val Arg Thr Asn Gly Thr Ser Lys Ile Lys Met Lys 565 570 Trp Gly Met Glu Met Arg Arg Cys Leu Leu Gln Ser Leu Gln Gln Ile Glu Ser Met Ile Glu Ala Glu Ser Ser Val Lys Glu Lys Asp Met Thr Lys Glu Phe Phe Glu Asn Lys Ser Glu Thr Trp Pro Ile Gly Glu Ser Pro Arg Gly Val Glu Glu Gly Ser Ile Gly Lys Val Cys Arg Thr Leu Leu Ala Lys Ser Val Phe Asn Ser Leu Tyr Ala Ser Pro Gln Leu Glu Gly Phe Ser Ala Glu Ser Arg Lys Leu Leu Leu Ile Val Gln Ala Leu Arg Asp Asn Leu Glu Pro Gly Thr Phe Asp Leu Gly Gly Leu Tyr Glu 680 Ala Ile Glu Glu Cys Leu Ile Asn Asp Pro Trp Val Leu Leu Asn Ala 690 695 700 Ser Trp Phe Asn Ser Phe Leu Thr His Ala Leu Lys <210> SEQ ID NO 45 <211> LENGTH: 252 <212> TYPE: PRT <213> ORGANISM: Influenza A virus <400> SEOUENCE: 45 Met Ser Leu Leu Thr Glu Val Glu Thr Tyr Val Leu Ser Ile Val Pro Ser Gly Pro Leu Lys Ala Glu Ile Ala Gln Arg Leu Glu Asn Val Phe 25 Ala Gly Lys Asn Thr Asp Leu Glu Ala Leu Met Glu Trp Leu Lys Thr Arg Pro Ile Leu Ser Pro Leu Thr Lys Gly Ile Leu Gly Phe Val Phe Thr Leu Thr Val Pro Ser Glu Arg Gly Leu Gln Arg Arg Arg Phe Val Gln Asn Ala Leu Asn Gly Asn Gly Asp Pro Asn Asn Met Asp Lys Ala Val Lys Leu Tyr Arg Lys Leu Lys Arg Glu Ile Thr Phe His Gly Ala Lys Glu Ile Ala Leu Ser Tyr Ser Ala Gly Ala Leu Ala Ser Cys Met Gly Leu Ile Tyr Asn Arg Met Gly Ala Val Thr Thr Glu Ser Ala Phe 135 Gly Leu Ile Cys Ala Thr Cys Glu Gln Ile Ala Asp Ser Gln His Lys Ser His Arg Gln Met Val Thr Thr Asn Pro Leu Ile Arg His Glu 170 Asn Arg Met Val Leu Ala Ser Thr Thr Ala Lys Ala Met Glu Gln Met 185 Ala Gly Ser Ser Glu Gln Ala Ala Glu Ala Met Glu Val Ala Ser Gln 200 Ala Arg Gln Met Val Gln Ala Met Arg Ala Ile Gly Thr His Pro Ser 215

Ser 225	Ser	Thr	Gly	Leu	Lys 230	Asn	Asp	Leu	Leu	Glu 235	Asn	Leu	Gln	Ala	Tyr 240
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Gly	Thr	Gly 35	Thr	Gly	Tyr	Thr	Met 40	Asp	Thr	Val	Asn	Arg 45	Thr	His	Gln
Tyr	Ser 50	Glu	Lys	Gly	Lys	Trp 55	Thr	Thr	Asn	Thr	Glu 60	Thr	Gly	Ala	Pro
Gln 65	Leu	Asn	Pro	Ile	Asp 70	Gly	Pro	Leu	Pro	Glu 75	Asp	Asn	Glu	Pro	Ser 80
Gly	Tyr	Ala	Gln	Thr 85	Asp	Cys	Val	Leu	Glu 90	Ala	Met	Ala	Phe	Leu 95	Glu
Glu	Ser	His	Pro 100	Gly	Ile	Phe	Glu	Asn 105	Ser	CAs	Leu	Glu	Thr 110	Met	Glu
Val	Val	Gln 115	Gln	Thr	Arg	Val	Asp 120	Arg	Leu	Thr	Gln	Gly 125	Arg	Gln	Thr
Tyr	Asp 130	Trp	Thr	Leu	Asn	Arg 135	Asn	Gln	Pro	Ala	Ala 140	Thr	Ala	Leu	Ala
Asn 145	Thr	Ile	Glu	Val	Phe 150	Arg	Ser	Asn	Gly	Leu 155	Thr	Ala	Asn	Glu	Ser 160
Gly	Arg	Leu	Ile	Asp 165	Phe	Leu	Lys	Asp	Val 170	Met	Glu	Ser	Met	Asp 175	Lys
Glu	Glu	Ile	Glu 180	Ile	Thr	Thr	His	Phe 185	Gln	Arg	Lys	Arg	Arg 190	Val	Arg
Asp	Asn	Met 195	Thr	Lys	Lys	Met	Val 200	Thr	Gln	Arg	Thr	Ile 205	Gly	Lys	Lys
ГÀа	Gln 210	Arg	Val	Asn	Lys	Arg 215	Ser	Tyr	Leu	Ile	Arg 220	Ala	Leu	Thr	Leu
Asn 225	Thr	Met	Thr	Lys	Asp 230	Ala	Glu	Arg	Gly	Lys 235	Leu	Lys	Arg	Arg	Ala 240
Ile	Ala	Thr	Pro	Gly 245	Met	Gln	Ile	Arg	Gly 250	Phe	Val	Tyr	Phe	Val 255	Glu
Thr	Leu	Ala	Arg 260	Ser	Ile	Cys	Glu	Lys 265	Leu	Glu	Gln	Ser	Gly 270	Leu	Pro
Val	Gly	Gly 275	Asn	Glu	Lys	Lys	Ala 280	Lys	Leu	Ala	Asn	Val 285	Val	Arg	Lys
Met	Met 290	Thr	Asn	Ser	Gln	Asp 295	Thr	Glu	Leu	Ser	Phe 300	Thr	Ile	Thr	Gly
Asp 305	Asn	Thr	Lys	Trp	Asn 310	Glu	Asn	Gln	Asn	Pro 315	Arg	Met	Phe	Leu	Ala 320
Met	Ile	Thr	Tyr	Ile 325	Thr	Lys	Asn	Gln	Pro 330	Glu	Trp	Phe	Arg	Asn 335	Ile
Leu	Ser	Ile	Ala	Pro	Ile	Met	Phe	Ser	Asn	Lys	Met	Ala	Arg	Leu	Gly

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			340					345					350		
Lys	Gly	Tyr 355	Met	Phe	Glu	Ser	360 Lys	Arg	Met	Lys	Leu	Arg 365	Thr	Gln	Ile
Pro	Ala 370	Glu	Met	Leu	Ala	Ser 375	Ile	Asp	Leu	Lys	Tyr 380	Phe	Asn	Glu	Ser
Thr 385	Arg	Lys	Lys	Ile	Glu 390	Lys	Ile	Arg	Pro	Leu 395	Leu	Ile	Asp	Gly	Thr 400
Ala	Ser	Leu	Ser	Pro 405	Gly	Met	Met	Met	Gly 410	Met	Phe	Asn	Met	Leu 415	Ser
Thr	Val	Leu	Gly 420	Val	Ser	Ile	Leu	Asn 425	Leu	Gly	Gln	Lys	Lys 430	Tyr	Thr
Lys	Thr	Thr 435	Tyr	Trp	Trp	Asp	Gly 440	Leu	Gln	Ser	Ser	Asp 445	Asp	Phe	Ala
Leu	Ile 450	Val	Asn	Ala	Pro	Asn 455	His	Glu	Gly	Ile	Gln 460	Ala	Gly	Val	Asp
Arg 465	Phe	Tyr	Arg	Thr	Cys 470	Lys	Leu	Val	Gly	Ile 475	Asn	Met	Ser	Lys	Lys 480
Lys	Ser	Tyr	Ile	Asn 485	Arg	Thr	Gly	Thr	Phe 490	Glu	Phe	Thr	Ser	Phe 495	Phe
Tyr	Arg	Tyr	Gly 500	Phe	Val	Ala	Asn	Phe 505	Ser	Met	Glu	Leu	Pro 510	Ser	Phe
Gly	Val	Ser 515	Gly	Ile	Asn	Glu	Ser 520	Ala	Asp	Met	Ser	Ile 525	Gly	Val	Thr
Val	Ile 530	Lys	Asn	Asn	Met	Ile 535	Asn	Asn	Asp	Leu	Gly 540	Pro	Ala	Thr	Ala
Gln 545	Met	Ala	Leu	Gln	Leu 550	Phe	Ile	Lys	Asp	Tyr 555	Arg	Tyr	Thr	Tyr	Arg 560
Cys	His	Arg	Gly	Asp 565	Thr	Gln	Ile	Gln	Thr 570	Arg	Arg	Ser	Phe	Glu 575	Leu
Lys	Lys	Leu	Trp 580	Glu	Gln	Thr	Arg	Ser 585	Lys	Ala	Gly	Leu	Leu 590	Val	Ser
Asp	Gly	Gly 595	Pro	Asn	Leu	Tyr	Asn 600	Ile	Arg	Asn	Leu	His 605	Ile	Pro	Glu
Val	Суs 610	Leu	ГÀЗ	Trp	Glu	Leu 615	Met	Asp	Glu	Asp	Tyr 620	Gln	Gly	Arg	Leu
Суз 625	Asn	Pro	Leu	Asn	Pro 630	Phe	Val	Ser	His	Lys 635	Glu	Ile	Glu	Ser	Val 640
Asn	Asn	Ala	Val	Val 645	Met	Pro	Ala	His	Gly 650	Pro	Ala	ГÀа	Ser	Met 655	Glu
Tyr	Asp	Ala	Val 660	Ala	Thr	Thr	His	Ser 665	Trp	Ile	Pro	ГÀа	Arg 670	Asn	Arg
Ser	Ile	Leu 675	Asn	Thr	Ser	Gln	Arg 680	Gly	Ile	Leu	Glu	Asp 685	Glu	Gln	Met
Tyr	Gln 690	Lys	СЛв	Cys	Asn	Leu 695	Phe	Glu	Lys	Phe	Phe 700	Pro	Ser	Ser	Ser
Tyr 705	Arg	Arg	Pro	Val	Gly 710	Ile	Ser	Ser	Met	Val 715	Glu	Ala	Met	Val	Ser 720
Arg	Ala	Arg	Ile	Asp 725	Ala	Arg	Ile	Asp	Phe 730	Glu	Ser	Gly	Arg	Ile 735	Lys
Lys	Glu	Glu	Phe 740	Ser	Glu	Ile	Met	Lys 745	Ile	Сув	Ser	Thr	Ile 750	Glu	Glu
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Leu Arg Arg Gln Lys Gln 755

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Asn	Lys	Phe 35	Ala	Ala	Ile	Cys	Thr 40	His	Leu	Glu	Val	Сув 45	Phe	Met	Tyr
Ser	Asp 50	Phe	His	Phe	Ile	Asn 55	Glu	Gln	Gly	Glu	Ser 60	Ile	Ile	Val	Glu
Leu 65	Gly	Asp	Pro	Asn	Ala 70	Leu	Leu	Lys	His	Arg 75	Phe	Glu	Ile	Ile	Glu 80
Gly	Arg	Asp	Arg	Thr 85	Met	Ala	Trp	Thr	Val 90	Val	Asn	Ser	Ile	95 Cys	Asn
Thr	Thr	Gly	Ala 100	Glu	ГÀа	Pro	Lys	Phe 105	Leu	Pro	Asp	Leu	Tyr 110	Asp	Tyr
ГÀа	Glu	Asn 115	Arg	Phe	Ile	Glu	Ile 120	Gly	Val	Thr	Arg	Arg 125	Glu	Val	His
Ile	Tyr 130	Tyr	Leu	Glu	Lys	Ala 135	Asn	Lys	Ile	Lys	Ser 140	Glu	Lys	Thr	His
Ile 145	His	Ile	Phe	Ser	Phe 150	Thr	Gly	Glu	Glu	Met 155	Ala	Thr	Lys	Ala	Asp 160
Tyr	Thr	Leu	Asp	Glu 165	Glu	Ser	Arg	Ala	Arg 170	Ile	Lys	Thr	Arg	Leu 175	Phe
Thr	Ile	Arg	Gln 180	Glu	Met	Ala	Ser	Arg 185	Gly	Leu	Trp	Asp	Ser 190	Phe	Arg
Gln	Ser	Glu 195	Arg	Gly	Glu	Glu	Thr 200	Ile	Glu	Glu	Arg	Phe 205	Glu	Ile	Thr
Gly	Thr 210	Met	Arg	Lys	Leu	Ala 215	Asp	Gln	Ser	Leu	Pro 220	Pro	Asn	Phe	Ser
Ser 225	Leu	Glu	Asn	Phe	Arg 230	Ala	Tyr	Val	Asp	Gly 235	Phe	Glu	Pro	Asn	Gly 240
Tyr	Ile	Glu	Gly	Lys 245	Leu	Ser	Gln	Met	Ser 250	Lys	Glu	Val	Asn	Ala 255	Arg
Ile	Glu	Pro	Phe 260	Leu	Lys	Thr	Thr	Pro 265	Arg	Pro	Leu	Arg	Leu 270	Pro	Asn
Gly	Pro	Pro 275	Cys	Ser	Gln	Arg	Ser 280	Lys	Phe	Leu	Leu	Met 285	Asp	Ala	Leu
Lys	Leu 290	Ser	Ile	Glu	Asp	Pro 295	Ser	His	Glu	Gly	Glu 300	Gly	Ile	Pro	Leu
Tyr 305	Asp	Ala	Ile	Lys	Cys 310	Met	Arg	Thr	Phe	Phe 315	Gly	Trp	Lys	Glu	Pro 320
Asn	Val	Val	Lys	Pro 325	His	Glu	Lys	Gly	Ile 330	Asn	Pro	Asn	Tyr	Leu 335	Leu
Ser	Trp	Lys	Gln 340	Val	Leu	Ala	Glu	Leu 345	Gln	Asp	Ile	Glu	Asn 350	Glu	Glu
Lys	Ile	Pro 355	Lys	Thr	Lys	Asn	Met 360	Lys	Lys	Thr	Ser	Gln 365	Leu	Lys	Trp
Ala	Leu	Gly	Glu	Asn	Met	Ala	Pro	Glu	Lys	Val	Asp	Phe	Asp	Asp	Cys

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	370					375					380				
385 Lys	Asp	Val	Gly	Asp	Leu 390	Lys	Gln	Tyr	Asp	Ser 395	Asp	Glu	Pro	Glu	Leu 400
Arg	Ser	Leu	Ala	Ser 405	Trp	Ile	Gln	Asn	Glu 410	Phe	Asn	ГÀа	Ala	Cys 415	Glu
Leu	Thr	Asp	Ser 420	Ser	Trp	Ile	Glu	Leu 425	Asp	Glu	Ile	Gly	Glu 430	Asp	Val
Ala	Pro	Ile 435	Glu	His	Ile	Ala	Ser 440	Met	Arg	Arg	Asn	Tyr 445	Phe	Thr	Ser
Glu	Val 450	Ser	His	CÀa	Arg	Ala 455	Thr	Glu	Tyr	Ile	Met 460	ГЛа	Gly	Val	Tyr
Ile 465	Asn	Thr	Ala	Leu	Leu 470	Asn	Ala	Ser	Сув	Ala 475	Ala	Met	Asp	Asp	Phe 480
Gln	Leu	Ile	Pro	Met 485	Ile	Ser	Lys	СЛа	Arg 490	Thr	Lys	Glu	Gly	Arg 495	Arg
Lys	Thr	Asn	Leu 500	Tyr	Gly	Phe	Ile	Ile 505	Lys	Gly	Arg	Ser	His 510	Leu	Arg
Asn	Asp	Thr 515	Asp	Val	Val	Asn	Phe 520	Val	Ser	Met	Glu	Phe 525	Ser	Leu	Thr
Asp	Pro 530	Arg	Leu	Glu	Pro	His 535	ГЛа	Trp	Glu	Lys	Tyr 540	CÀa	Val	Leu	Glu
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Met	Phe	Leu	Tyr	Val 565	Arg	Thr	Asn	Gly	Thr 570	Ser	ГЛа	Ile	Lys	Met 575	Lys
Trp	Gly	Met	Glu 580	Met	Arg	Arg	Cha	Leu 585	Leu	Gln	Ser	Leu	Gln 590	Gln	Ile
Glu	Ser	Met 595	Ile	Glu	Ala	Glu	Ser 600	Ser	Val	ГÀЗ	Glu	Lys 605	Asp	Met	Thr
Lys	Glu 610	Phe	Phe	Glu	Asn	Lys 615	Ser	Glu	Thr	Trp	Pro 620	Ile	Gly	Glu	Ser
Pro 625	Lys	Gly	Val	Glu	Glu 630	Ser	Ser	Ile	Gly	Lys 635	Val	CAa	Arg	Thr	Leu 640
Leu	Ala	Lys	Ser	Val 645	Phe	Asn	Ser	Leu	Tyr 650	Ala	Ser	Pro	Gln	Leu 655	Glu
Gly	Phe	Ser	Ala 660	Glu	Ser	Arg	Lys		Leu		Ile	Val	Gln 670	Ala	Leu
Arg	Asp	Asn 675	Leu	Glu	Pro	Gly	Thr 680	Phe	Asp	Leu	Gly	Gly 685	Leu	Tyr	Glu
Ala	Ile 690	Glu	Glu	CÀa	Leu	Ile 695	Asn	Asp	Pro	Trp	Val 700	Leu	Leu	Asn	Ala
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Gly	Glu	Arg	Gln 20	Asn	Ala	Thr	Glu	Ile 25	Arg	Ala	Ser	Val	Gly 30	Lys	Met

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Leu	Ser 50	Asp	Tyr	Glu	Gly	Arg 55	Leu	Ile	Gln	Asn	Ser 60	Leu	Thr	Ile	Glu
Arg 65	Met	Val	Leu	Ser	Ala 70	Phe	Asp	Glu	Arg	Arg 75	Asn	ГЛа	Tyr	Leu	Glu 80
Glu	His	Pro	Ser	Ala 85	Gly	Lys	Asp	Pro	Pys	Lys	Thr	Gly	Gly	Pro 95	Ile
Tyr	Arg	Arg	Val 100	Asn	Gly	Lys	Trp	Met 105	Arg	Glu	Leu	Ile	Leu 110	Tyr	Asp
Lys	Glu	Glu 115	Ile	Arg	Arg	Ile	Trp 120	Arg	Gln	Ala	Asn	Asn 125	Gly	Asp	Asp
Ala	Thr 130	Ala	Gly	Leu	Thr	His 135	Met	Met	Ile	Trp	His 140	Ser	Asn	Leu	Asn
Asp 145	Ala	Thr	Tyr	Gln	Arg 150	Thr	Arg	Ala	Leu	Val 155	Arg	Thr	Gly	Met	Asp 160
Pro	Arg	Met	Cys	Ser 165	Leu	Met	Gln	Gly	Ser 170	Thr	Leu	Pro	Arg	Arg 175	Ser
Gly	Ala	Ala	Gly 180	Ala	Ala	Val	Lys	Gly 185	Val	Gly	Thr	Met	Val 190	Met	Glu
Leu	Val	Arg 195	Met	Ile	Lys	Arg	Gly 200	Ile	Asn	Asp	Arg	Asn 205	Phe	Trp	Arg
Gly	Glu 210	Asn	Gly	Arg	Lys	Thr 215	Arg	Ile	Ala	Tyr	Glu 220	Arg	Met	Сув	Asn
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Gln	Val	Arg	Glu	Ser 245	Arg	Asp	Pro	Gly	Asn 250	Ala	Glu	Phe	Glu	Asp 255	Leu
Thr	Phe	Leu	Ala 260	Arg	Ser	Ala	Leu	Ile 265	Leu	Arg	Gly	Ser	Val 270	Ala	His
Lys	Ser	Сув 275	Leu	Pro	Ala	Cys	Val 280	Tyr	Gly	Pro	Ala	Val 285	Ala	Ser	Gly
Tyr	Asp 290	Phe	Glu	Arg	Glu	Gly 295	Tyr	Ser	Leu	Val	Gly 300	Ile	Asp	Pro	Phe
Arg 305	Leu	Leu	Gln	Asn	Ser 310	Gln	Val	Tyr	Ser	Leu 315	Ile	Arg	Pro	Asn	Glu 320
Asn	Pro	Ala	His	Lys 325	Ser										
0.1)> SI	10 TI		10											
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	2 > T? 3 > OF			Inf	luen:	za A	viru	ıs							
< 400)> SI	EQUEI	ICE :	49											
Met 1	Ser	Leu	Leu	Thr 5	Glu	Val	Glu	Thr	Tyr 10	Val	Leu	Ser	Ile	Ile 15	Pro
Ser	Gly	Pro	Leu 20	Lys	Ala	Glu	Ile	Ala 25	Gln	Arg	Leu	Glu	Asp 30	Val	Phe
Ala	Gly	Lys 35	Asn	Thr	Asp	Leu	Glu 40	Val	Leu	Met	Glu	Trp 45	Leu	Lys	Thr
Arg	Pro 50	Ile	Leu	Ser	Pro	Leu 55	Thr	Lys	Gly	Ile	Leu 60	Gly	Phe	Val	Phe
Thr 65	Leu	Thr	Val	Pro	Ser 70	Glu	Arg	Gly	Leu	Gln 75	Arg	Arg	Arg	Phe	Val 80

Gln Asn Ala Leu Asn Gly Asn Gly Asp Pro Asn Asn Met Asp Lys Ala

90 Val Lys Leu Tyr Arg Lys Leu Lys Arg Glu Ile Thr Phe His Gly Ala Lys Glu Ile Ser Leu Ser Tyr Ser Ala Gly Ala Leu Ala Ser Cys Met Gly Leu Ile Tyr Asn Arg Met Gly Ala Val Thr Thr Glu Val Ala Phe Gly Leu Val Cys Ala Thr Cys Glu Gln Ile Ala Asp Ser Gln His Arg Ser His Arg Gln Met Val Thr Thr Thr Asn Pro Leu Ile Arg His Glu Asn Arg Met Val Leu Ala Ser Thr Thr Ala Lys Ala Met Glu Gln Met Ala Gly Ser Ser Glu Gln Ala Ala Glu Ala Met Glu Val Ala Ser Gln 200 205 Ala Arg Gln Met Val Gln Ala Met Arg Thr Ile Gly Thr His Pro Ser 215 Ser Ser Ala Gly Leu Lys Asn Asp Leu Leu Glu Asn Leu Gln Ala Tyr 230 Gln Lys Arg Met Gly Val Gln Met Gln Arg Phe Lys 245 <210> SEQ ID NO 50 <211> LENGTH: 566 <212> TYPE: PRT <213> ORGANISM: Influenza A virus <400> SEQUENCE: 50 Met Lys Ala Ile Leu Val Val Leu Leu Tyr Thr Phe Ala Thr Ala Asn 10 Ala Asp Thr Leu Cys Ile Gly Tyr His Ala Asn Asn Ser Thr Asp Thr 25 Val Asp Thr Val Leu Glu Lys Asn Val Thr Val Thr His Ser Val Asn Leu Leu Glu Asp Lys His Asn Gly Lys Leu Cys Lys Leu Arg Gly Val Ala Pro Leu His Leu Gly Lys Cys Asn Ile Ala Gly Trp Ile Leu Gly Asn Pro Glu Cys Glu Ser Leu Ser Thr Ala Ser Ser Trp Ser Tyr Ile Val Glu Thr Pro Ser Ser Asp Asn Gly Thr Cys Tyr Pro Gly Asp Phe Ile Asp Tyr Glu Glu Leu Arg Glu Gln Leu Ser Ser Val Ser Ser Phe Glu Arg Phe Glu Ile Phe Pro Lys Thr Ser Ser Trp Pro Asn His Asp 135 Ser Asn Lys Gly Val Thr Ala Ala Cys Pro His Ala Gly Ala Lys Ser 155 Phe Tyr Lys Asn Leu Ile Trp Leu Val Lys Lys Gly Asn Ser Tyr Pro Lys Leu Ser Lys Ser Tyr Ile Asn Asp Lys Gly Lys Glu Val Leu Val Leu Trp Gly Ile His His Pro Ser Thr Ser Ala Asp Gln Gln Ser Leu

													CIII		
		195					200					205			
Tyr	Gln 210	Asn	Ala	Asp	Thr	Tyr 215	Val	Phe	Val	Gly	Ser 220	Ser	Arg	Tyr	Ser
Lys 225	Lys	Phe	Lys	Pro	Glu 230	Ile	Ala	Ile	Arg	Pro 235	Lys	Val	Arg	Asp	Gln 240
Glu	Gly	Arg	Met	Asn 245	Tyr	Tyr	Trp	Thr	Leu 250	Val	Glu	Pro	Gly	Asp 255	ГЛа
Ile	Thr	Phe	Glu 260	Ala	Thr	Gly	Asn	Leu 265	Val	Val	Pro	Arg	Tyr 270	Ala	Phe
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Pro	ГЛа	Tyr	Val	125 325	Ser	Thr	ГХа	Leu	Arg 330	Leu	Ala	Thr	Gly	Leu 335	Arg
Asn	Ile	Pro	Ser 340	Ile	Gln	Ser	Arg	Gly 345	Leu	Phe	Gly	Ala	Ile 350	Ala	Gly
Phe	Ile	Glu 355	Gly	Gly	Trp	Thr	Gly 360	Met	Val	Asp	Gly	Trp 365	Tyr	Gly	Tyr
His	His 370	Gln	Asn	Glu	Gln	Gly 375	Ser	Gly	Tyr	Ala	Ala 380	Asp	Leu	Lys	Ser
Thr 385	Gln	Asn	Ala	Ile	390	Glu	Ile	Thr	Asn	Lys 395	Val	Asn	Ser	Val	Ile 400
Glu	Lys	Met	Asn	Thr 405	Gln	Phe	Thr	Ala	Val 410	Gly	Lys	Glu	Phe	Asn 415	His
Leu	Glu	Lys	Arg 420	Ile	Glu	Asn	Leu	Asn 425	Lys	Lys	Val	Aap	Asp 430	Gly	Phe
Leu	Asp	Ile 435	Trp	Thr	Tyr	Asn	Ala 440	Glu	Leu	Leu	Val	Leu 445	Leu	Glu	Asn
Glu	Arg 450		Leu	_	_	His	_				Lys 460		Leu	Tyr	Glu
Lys 465			Ser			Lys			Ala	Lys 475	Glu	Ile	Gly	Asn	Gly 480
	Phe	Glu	Phe	Tyr 485		ГÀа	Cys	Asp	Asn 490		Cys	Met	Glu	Ser 495	
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Gln	Ile 530		Ala	Ile	Tyr	Ser 535		Val	Ala	Ser	Ser 540		Val	Leu	Val
Val 545		Leu	Gly	Ala	Ile 550	Ser	Phe	Trp	Met	Сув 555		Asn	Gly	Ser	Leu 560
	Cys	Arg	Ile	Сув 565						555					500
				203											

The invention claimed is:

- 1. A reassortant influenza A virus comprising six backbone viral segments, an HA segment and an NA segment, wherein two, three, four, five, or six backbone viral segments are from a donor strain, wherein the donor strain is selected from the 5 group consisting of 105p30 and PR8-X, wherein (a) at least one donor segment is selected from the group consisting of SEQ ID NOs 9-12 and SEQ ID NOs 17-22, (b) at least two donor segments are selected from the group consisting of SEQ ID NOs 9-13 and SEQ ID NOs: 17-22, or (c) at least two donor segments are selected from the group consisting of SEQ ID NOs 9-12 and 14 and SEQ ID NOs 17-22, and at least one viral segment is derived from a second influenza strain.
- 2. The reassortant influenza A virus of claim 1 wherein at $_{15}$ least one backbone viral segment comprises the sequence of SEQ ID NO: 17 or SEQ ID NO: 20.
- 3. The reassortant influenza A virus of claim 1, wherein the virus comprises backbone segments from two or more donor
- 4. The reassortant influenza A virus of claim 3, wherein the PB1 and the PB2 viral segments are from the same donor
- 5. The reassortant influenza A virus of claim 4, wherein the PB1 viral segment has at least 95% identity to SEQ ID NO: 18 25 and the PB2 viral segment has at least 95% identity to SEQ ID
- 6. The reassortant influenza A virus of claim 5, wherein the virus further comprises a viral segment having at least 95% identity to a sequence selected from the group consisting of 30 SEQ ID NOs 17-22.
- 7. The reassortant influenza A virus of claim 2, wherein the virus comprises the PB2 segment of SEQ ID NO: 19, the PB1 segment of SEQ ID NO: 18 and the NP segment of SEQ ID NO: 20.
- 8. The reassortant influenza A virus of claim 1, wherein the virus has the HA segment from a pandemic influenza strain.
- 9. A method of preparing a reassortant influenza virus comprising steps of
 - construct(s) which encode(s) the viral segments required to produce an influenza virus wherein at least one backbone viral segment(s) is/are from a 105p30 and/or a PR8 X influenza strain, wherein (a) at least one donor segment is selected from the group consisting of 45 SEQ ID NOs 9-12 and SEQ ID NOs 17-22, (b) two or more backbone viral segments are selected from the group consisting of SEQ ID NOs 9-13 and SEQ ID NOs 17-22, or (c) two or more backbone viral segments are selected from the group consisting of SEQ ID NOs 9-12, 50 SEQ ID NO: 14 and SEQ ID NOs 17-22, and wherein at least one viral segment is derived from a second influenza strain; and
 - (ii) culturing the culture host in order to produce reassortant virus.
- 10. The method of claim 9, further comprising the step (iii) of purifying the reassortant virus obtained in step (ii).
- 11. The method of claim 9 wherein the at least one viral segment from the second influenza strain is the HA segment.
 - 12. A method for producing influenza viruses comprising: 60
 - (a) infecting a culture host with the reassortant influenza virus of claim 1;
 - (b) culturing the host from step (a) to produce the virus; and
 - (c) purifying the virus obtained in step (b).

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- 13. A method of preparing a vaccine, comprising steps of
- (x) preparing a virus by the method of claim 12 and
- (y) preparing vaccine from the virus.
- 14. The method of claim 13, wherein
- (i) the culture host is an embryonated hen egg, or
- (ii) the culture host is a eukarvotic cell.
- 15. The method of claim 14, wherein the culture host is (ii) and the eukaryotic cell is an MDCK, Vero or PerC6 cell.
- 16. The method of claim 15, wherein the eukaryotic cell grows
- (i) adherently or
 - (ii) in suspension.
- 17. The method of claim 15, wherein the eukaryotic cell is cell line MDCK 33016 (DSM ACC2219).
- 18. The method of claim 13, wherein step (b) includes inactivating the virus.
- 19. The method of claim 13, wherein the vaccine is a whole virion vaccine, a split virion vaccine, a surface antigen vaccine or a virosomal vaccine.
- 20. The method of claim 13, wherein the vaccine contains 20 less than 10 ng of residual host cell DNA per dose.
- 21. A composition comprising six backbone viral segments, an HA segment and an NA segment, wherein two, three, four, five, or six donor polypeptides are encoded by influenza A backbone viral segments from at least one donor strain, wherein the at least one donor strain is selected from the group consisting of 105p30 and PR8 X, wherein (a) at least one donor segment is selected from the group consisting of SEQ ID NOs 9-12 and SEQ ID NOs 17-22, (b) at least two donor segments are selected from the group consisting of SEQ ID NOs 9-13 and SEQ ID NOs 17-22, or (c) at least two donor segments are selected from the group consisting of SEQ ID NOs 9-12, SEQ ID NO: 14 and SEQ ID NOs 17-22, and a hemagglutinin polypeptide encoded by the HA segment that is not from influenza strains 105p30 (SEQ ID NO: 23) or 35 PR8 X (SEQ ID NO: 15).
 - 22. The reassortant influenza A virus of claim 1, comprising at least one donor segment is selected from the group consisting of SEQ ID NOs 9-12.
- 23. The reassortant influenza A virus of claim 1, compris-(i) introducing into a culture host one or more expression 40 ing at least two donor segments are selected from (a) the group consisting of SEQ ID NOs 9-13 and SEQ ID NOs 17-22, or (b) the group consisting of SEQ ID NOs 9-12, SEQ ID NO: 14 and SEQ ID NOs 17-22.
 - 24. The method of preparing a reassortant influenza virus of claim 9, wherein at least one donor segment is selected from the group consisting of SEQ ID NOs 9-12.
 - 25. The method of preparing a reassortant influenza virus of claim 9, wherein at least two of the two or more backbone viral segments are selected from (a) the group consisting of SEQ ID NOs 9-13 and SEQ ID NOs 17-22, or (b) the group consisting of SEQ ID NOs 9-12, SEQ ID NO: 14 and SEQ ID NO 17-22.
 - 26. The composition of claim 21, wherein at least one donor segment is selected from the group consisting of SEQ 55 ID NOs 9-12.
 - 27. The composition of claim 21, wherein at least two donor segments are selected from (a) the group consisting of SEQ ID NOs: 9-13, or (b) the group consisting of SEQ ID NOs 9-12 and SEQ ID NO: 14.
 - 28. The composition of claim 26, wherein the composition is an influenza vaccine.
 - 29. The composition of claim 27, wherein the composition is an influenza vaccine.